Contents lists available at ScienceDirect

### **Neural Networks**

journal homepage: www.elsevier.com/locate/neunet

# Effect of diverse recoding of granule cells on optokinetic response in a cerebellar ring network with synaptic plasticity

### Sang-Yoon Kim, Woochang Lim\*

Institute for Computational Neuroscience and Department of Science Education, Daegu National University of Education, Daegu 42411, Republic of Korea

### ARTICLE INFO

ABSTRACT

Article history: Received 26 March 2020 Received in revised form 12 November 2020 Accepted 24 November 2020 Available online 28 November 2020

Keywords: Optokinetic response Cerebellar ring network Diverse recoding Effective long-term depression Effective motor learning We consider a cerebellar ring network for the optokinetic response (OKR), and investigate the effect of diverse recoding of granule (GR) cells on OKR by varying the connection probability  $p_c$  from Golgi to GR cells. For an optimal value of  $p_c^*$  (= 0.06), individual GR cells exhibit diverse spiking patterns which are in-phase, anti-phase, or complex out-of-phase with respect to their population-averaged firing activity. Then, these diversely-recoded signals via parallel fibers (PFs) from GR cells are effectively depressed by the error-teaching signals via climbing fibers from the inferior olive which are also in-phase ones. Synaptic weights at in-phase PF-Purkinje cell (PC) synapses of active GR cells are strongly depressed via strong long-term depression (LTD), while those at anti-phase and complex out-of-phase PF-PC synapses are weakly depressed through weak LTD. This kind of "effective" depression (i.e., strong/weak LTD) at the PF-PC synapses causes a big modulation in firings of PCs, which then exert effective inhibitory coordination on the vestibular nucleus (VN) neuron (which evokes OKR). For the firing of the VN neuron, the learning gain degree  $\mathcal{L}_g$ , corresponding to the modulation gain ratio, increases with increasing the learning cycle, and it saturates at about the 300th cycle. By varying  $p_c$  from  $p_c^*$ , we find that a plot of saturated learning gain degree  $\mathcal{L}_{g}^{*}$  versus  $p_{c}$  forms a bell-shaped curve with a peak at  $p_c^*$  (where the diversity degree in spiking patterns of GR cells is also maximum). Consequently, the more diverse in recoding of GR cells, the more effective in motor learning for the OKR adaptation. © 2020 Published by Elsevier Ltd.

### 1. Introduction

The cerebellum receives information from the sensory systems, the spinal cord and other parts of the brain and then regulates motor movements. For a smoothly integrated body movement, the cerebellum activates a large set of spatially separated muscles in a precise order and timing. Thus, the cerebellum plays an essential role in fine motor control (i.e., precise spatial and temporal motor control) for coordinating voluntary movements such as posture, balance, and locomotion, resulting in smooth and balanced muscular activity (Ito, 1984, 2002a, 2012). Moreover, it is also involved in higher cognitive functions such as time perception and language processing (Ito, 2002a, 2012). Animals and humans with damaged cerebella are still able to initiate movements, but these movements become slow, inexact, and uncoordinated (Gilman, Bloedel, & Lechtenberg, 1981; Manto, 2010).

The spatial information of movements (e.g., amplitude or velocity) is called "gain", while the temporal information of

\* Corresponding author. E-mail addresses: sykim@icn.re.kr (S.-Y. Kim), wclim@icn.re.kr (W. Lim).

https://doi.org/10.1016/j.neunet.2020.11.014 0893-6080/© 2020 Published by Elsevier Ltd. movements (e.g., initiation or termination) is called "timing" (Yamazaki & Nagao, 2012). The goal of cerebellar motor learning is to perform precise gain and temporal control for movements. The cerebellar mechanisms for gain and timing control for eve movements have been studied in the two types of experimental paradigms; (1) gain control for the optokinetic response (OKR) and the vestibulo-ocular reflex (Ito, 1984, 1998) and (2) timing control for the eyeblink conditioning (Christian & Thompson, 2003; Mauk & Donegan, 1997). Here, we are concerned about gain adaptation of OKR. When the eye tracks a moving object with the stationary head, OKR may be seen. When the moving object is out of the field of vision, the eye moves back rapidly to the original position where it first saw. In this way, OKR consists of two consecutive slow and fast phases. Experimental works on OKR in vertebrates such as rabbits, mice, and zebrafishes have been done in diverse aspects (Harvey, De'Sperati, & Strata, 1997; Huang & Neuhauss, 2008; Iwashita, Kanai, Funabiki, Matsuda, & Hirano, 2001; Matsuno et al., 2016; Nagao, 1983, 1988; Scheetz et al., 2017; Tabata, Shimizu, Wada, Miura, & Kawano, 2010).

In the Marr–Albus–Ito theory for cerebellar computation (Albus, 1971; Ito, 1984; Marr, 1969), the cerebellum is considered to act as a simple perceptron (i.e., pattern associator) which associates input [mossy fiber (MF)] patterns with output [Purkinje







cell (PC)] patterns. The input patterns become more sparse and less similar to each other via recoding process in the granular layer, consisting of the granule (GR) and the Golgi (GO) cells. Then, the recoded inputs are fed into the PCs via the parallel fibers (PFs) (i.e., the axons of GR cells). In addition to the PF recoded signals, the PCs also receive the error-teaching signals through the climbing-fiber (CF) from the inferior olive (IO). The PF-PC synapses are assumed to be the only synapses at which motor learning occurs. Thus, synaptic plasticity may occur at the PF-PC synapses (i.e., their synaptic strengths may be potentiated or depressed). Marr in Marr (1969) assumes that a Hebbian type of long-term potentiation (LTP) (i.e., increase in synaptic strengths) occurs at the PF-PC synapses when both the PF and the CF signals are conjunctively excited (Brindley, 1964; Hebb, 1949). This Marr's theory (which directly relates the cerebellar function to its structure) represents a milestone in the history of cerebellum (Strata, 2009). In contrast to Marr's learning via LTP, Albus in Albus (1971) assumes that synaptic strengths at PF-PC synapses are depressed [i.e., an anti-Hebbian type of long-term depression (LTD) occurs] in the case of conjunctive excitations of both the PF and the CF signals. In the case of Albus' learning via LTD, PCs learn when to stop their inhibition (i.e. when to disinhibit) rather than when to fire. In several later experimental works done by Ito et al., clear evidences for LTD were obtained (Ito, 2000; Ito, Sakurai, & Tongroach, 1982; Sakurai, 1987). Thus, LTD became established as a unique type of synaptic plasticity for cerebellar motor learning (Ito, 1989, 2001, 2002b; Ito & Kano, 1982).

In addition to experimental works on the OKR (Harvey et al., 1997; Huang & Neuhauss, 2008; Iwashita et al., 2001; Matsuno et al., 2016; Nagao, 1983, 1988; Scheetz et al., 2017; Tabata et al., 2010), computational works have also been performed (Gomi & Kawato, 1992; Yamazaki & Nagao, 2012). The Marr-Albus model of the cerebellum was also reformulated to incorporate dynamical responses in terms of the adaptive filter model (used in the field of engineering control) (Dean, Porrill, Ekerot, & Jörntell, 2010; Fujita, 1982). The cerebellar structure may be mapped onto an adaptive filter structure. Through analysis-synthesis process of the adaptive filter model. the (time-varying) filter inputs (i.e., MF "context" signals for the post-eye-movement) are analyzed into diverse component signals (i.e., diversely recoded PF signals). Then, they are weighted (i.e., synaptic plasticity at PF-PC synapses) and recombined to generate the filter output (i.e., firing activity of PCs). The filter is adaptive because its weights are adjusted by an error-teaching signal (i.e., CF signal), employing the covariance learning rule (Sejnowski, 1977). Using this adaptive filter model, gain adaptation of OKR was successfully simulated (Gomi & Kawato, 1992). Recently, Yamazaki and Nagao in Yamazaki and Nagao (2012) employed a spiking network model, which was originally introduced for Pavlovian delay eyeblink conditioning (Yamazaki & Tanaka, 2007a). As elements in the spiking network, leaky integrate-and-fire neuron models were used, and parameter values for single cells and synaptic currents were adopted from physiological data. Through a largescale computer simulation, some features of OKR adaptation were successfully reproduced.

However, the effects of diverse recoding of GR cells on the OKR adaption in previous computational works are still needed to be more clarified in several dynamical aspects. First of all, dynamical classification of diverse PF signals (corresponding to the recoded outputs of GR cells) must be completely done for clear understanding their association with the error-teaching CF signals. Then, based on such dynamical classification of diverse spiking patterns of GR cells, synaptic plasticity at PF-PC synapses and subsequent learning progress could be more clearly understood. As a result, understanding on the learning gain and the learning progress for the OKR adaptation is expected to be so much improved.

To this end, we consider a cerebellar spiking ring network for the OKR adaptation, and first make a dynamical classification of diverse spiking patterns of GR cells (i.e., diverse PF signals) by changing the connection probability  $p_c$  from GO to GR cells in the granular layer. An instantaneous whole-population spike rate  $R_{GR}(t)$  (which is obtained from the raster plot of spikes of individual neurons) may well describe collective firing activity in the whole population of GR cells (Brunel, 2000; Brunel & Hakim, 1999, 2008; Brunel & Hansel, 2006; Brunel & Wang, 2003; Geisler, Brunel, & Wang, 2005; Kim & Lim, 2014; Wang, 2010).  $R_{GR}(t)$  is in-phase with respect to the sinusoidally-modulating MF input signal for the post-eye-movement, although it has a central flattened plateau due to inhibitory inputs from GO cells.

The whole population of GR cells is divided into GR clusters. These GR clusters show diverse spiking patterns which are in-phase, anti-phase, and complex out-of-phase relative to the instantaneous whole-population spike rate  $R_{GR}(t)$ . Each spiking pattern is characterized in terms of the "conjunction" index, denoting the resemblance (or similarity) degree between the spiking pattern and the instantaneous whole-population spike rate  $R_{GR}(t)$  (corresponding to the population-averaged firing activity). To quantify the degree of diverse recoding of GR cells, we introduce the diversity degree  $\mathcal{D}$ , given by the relative standard deviation in the distribution of conjunction indices of all spiking patterns. We mainly consider an optimal case of  $p_c^*$  (= 0.06) where the spiking patterns of GR clusters are the most diverse. In this case,  $\mathcal{D}^* \simeq 1.613$  which is a quantitative measure for diverse recoding of GR cells in the granular layer. We also investigate dynamical origin of these diverse spiking patterns of GR cells. It is thus found that, diverse total synaptic inputs (including both the excitatory MF inputs and the inhibitory inputs from the presynaptic GO cells) into the GR clusters result in production of diverse spiking patterns (i.e. outputs) in the GR clusters.

Next, based on dynamical classification of diverse spiking patterns of GR clusters, we employ a refined rule for synaptic plasticity (constructed from the experimental result in Safo & Regehr, 2008) and make an intensive investigation on the effect of diverse recoding of GR cells on synaptic plasticity at PF-PC synapses and the subsequent learning process. PCs (corresponding to the output of the cerebellar cortex) receive both the diversely-recoded PF signals from GR cells and the error-teaching CF signals from the IO neuron. We also note that the CF signals are in-phase with respect to the instantaneous whole-population spike rate  $R_{GR}(t)$ . In this case, CF signals may be regarded as "instructors", while PF signals can be considered as "students". Then, in-phase PF student signals are strongly depressed (i.e., their synaptic weights at PF-PC synapses are greatly decreased through strong LTD) by the in-phase CF instructor signals. On the other hand, out-of-phase PF student signals are weakly depressed (i.e., their synaptic weights at PF-PC synapses are a little decreased via weak LTD) due to the phase difference between the student PF and the instructor CF signals. In this way, the student PF signals are effectively (i.e., strongly/weakly) depressed by the error-teaching instructor CF signals.

During learning cycles, the "effective" depression (i.e., strong/weak LTD) at PF-PC synapses may cause a big modulation in firing activities of PCs, which then exerts effective inhibitory coordination on vestibular nucleus (VN) neuron (which evokes OKR eye-movement). For the firing activity of VN neuron, the learning gain degree  $\mathcal{L}_g$ , corresponding to the modulation gain ratio (i.e., normalized modulation divided by that at the 1st cycle), increases with learning cycle, and it eventually becomes saturated.

Saturation in the learning progress is clearly shown in the IO system. During the learning cycle, the IO neuron receives both the excitatory sensory signal for a desired eye-movement and



**Fig. 1.** (a) Optokinetic response when the eye tracks stripe slip. External sensory signals: (b1) firing rate  $f_{MF}(t)$  of the mossy-fiber (MF) context signal for the post-eye-movement and (b2) firing rate  $f_{DS}(t)$  of the inferior-olive (IO) desired signal (DS) for a desired eye-movement.

the inhibitory signal from the VN neuron (representing a realized eye-movement). We introduce the learning progress degree  $\mathcal{L}_p$ , given by the ratio of the cycle-averaged inhibitory input from the VN neuron to the cycle-averaged excitatory input of the desired sensory signal. With increasing cycle, the cycle-averaged inhibition (from the VN neuron) increases (i.e.,  $\mathcal{L}_p$  increases), and converges to the constant cycle-averaged excitation (through the desired signal). Thus, at about the 300th cycle, the learning progress degree becomes saturated at  $\mathcal{L}_p = 1$ . At this saturated stage, the cycle-averaged excitatory and inhibitory inputs to the IO neuron become balanced, and we get the saturated learning gain degree  $\mathcal{L}_g^*$ ( $\simeq 1.608$ ) in the VN.

By changing  $p_c$  from  $p_c^*$  (= 0.06), we also investigate the effect of diverse recoding of GR cells on the OKR adaptation. Thus, the plot of saturated learning gain degree  $\mathcal{L}_g^*$  versus  $p_c$  is found to form a bell-shaped curve with a peak ( $\mathcal{L}_g^* \simeq 1.608$ ) at  $p_c^*$ . With increasing or decreasing  $p_c$  from  $p_c^*$ , the diversity degree  $\mathcal{D}$  in firing activities of GR cells also forms a bell-shaped curve with a maximum value ( $\mathcal{D}^* \simeq 1.613$ ) at  $p_c^*$ . We note that both the saturated learning gain degree  $\mathcal{L}_g^*$  and the diversity degree  $\mathcal{D}$  have a strong correlation with the Pearson's correlation coefficient  $r \simeq 0.9998$  (Pearson, 1895). Consequently, the more diverse the recoding of GR cells, the more effective the motor learning for the OKR adaptation.

This paper is organized as follows: In Section 2, we describe the cerebellar ring network for the OKR, composed of the granular layer, the Purkinje-molecular layer, and the VN-IO part. The governing equations for the population dynamics in the ring network are also presented, along with a refined rule for the synaptic plasticity at the PF-PC synapses. Then, in main Section 3, we investigate the effect of diverse recoding of GR cells on motor learning for the OKR adaptation by changing  $p_c$ . Finally, we give summary and discussion in Section 4. In Appendix B, glossary for various terms characterizing the cerebellar model is given to help readers keep track of them.

### 2. Cerebellar ring network with synaptic plasticity

In this section, we describe our cerebellar ring network with synaptic plasticity for the OKR adaptation. Fig. 1(a) shows OKR which may be seen when the eye tracks successive stripe slip with the stationary head. When each moving stripe is out of the field of vision, the eye moves back quickly to the original position where it first saw. Thus, OKR is composed of two consecutive slow and fast phases (i.e., slow tracking eye-movement and fast reset saccade). It takes 2 s (corresponding to 0.5 Hz) for one complete slip of each stripe. Slip of the visual image across large portions of the retina is the stimulus that produces the adaptation of the optokinetic system.

### 2.1. MF context signal and IO desired signal

There are two types of sensory signals which transfer the retinal slip information from the retina to their targets by passing intermediate pre-cerebellar nuclei (PCN). In the 1st case, the retinal slip information first passes the pretectum in the midbrain, then passes the nucleus reticularis tegmentis pontis (NRTP) in the pons, and finally it is transferred to the granular layer (consisting of GR and GO cells) in the cerebellar cortex via MF sensory signal containing "context" for the post-eye-movement. The MF context signals are modeled in terms of Poisson spike trains which modulate sinusoidally at the stripe-slip frequency  $f_s = 0.5$  Hz (i.e., one-cycle period: 2 s) with the peak firing rate of 30 Hz (i.e., 30 spikes/s) (Yamazaki & Nagao, 2012). The firing frequency  $f_{\rm MF}$  of Poisson spike trains for the MF context signal is given by

$$f_{\rm MF}(t) = -\bar{f}_{\rm MF}\cos(2\pi f_{\rm s}t) + \bar{f}_{\rm MF}; \quad \bar{f}_{\rm MF} = 15 \text{ Hz},$$
 (1)

which is shown in Fig. 1(b1).

In the 2nd case, the retinal slip information passes only the pretectum, and then (without passing NRTP) it is directly fed into to the IO via a sensory signal for a "desired" eye-movement. As in the MF context signals, the IO desired signals are also modeled in terms of the same kind of sinusoidally modulating Poisson spike trains at the stripe-slip frequency  $f_s = 0.5$  Hz. The firing frequency  $f_{DS}$  of Poisson spike trains for the IO desired signal (DS) is given by:

$$f_{\rm DS}(t) = -\bar{f}_{\rm DS}\cos(2\pi f_{\rm s}t) + \bar{f}_{\rm DS}; \quad \bar{f}_{\rm DS} = 1.5 \text{ Hz},$$
 (2)

which is shown in Fig. 1(b2). In this case, the peak firing rate for the IO desired signal is reduced to 3 Hz to satisfy low mean firing rates ( $\sim 1.5$  Hz) of individual IO neurons (i.e., corresponding to 1/10 of the peak firing rate of the MF signal) (Llinás, 2014; Mathy et al., 2009).

#### 2.2. Architecture for cerebellar ring network

As in the famous small-world ring network (Strogatz, 2001; Watts & Strogatz, 1998), we develop a one-dimensional ring network with a simple architecture, which is in contrast to the twodimensional square-lattice network (Yamazaki & Nagao, 2012; Yamazaki & Tanaka, 2007a). This kind of ring network has advantage for computational and analytical efficiency, and its visual representation may also be easily made.

Here, we employ such a cerebellar ring network for the OKR. Fig. 2(a) shows the box diagram for the cerebellar network. The granular layer, corresponding to the input layer of the cerebellar cortex, consists of the excitatory GR cells and the inhibitory GO cells. On the other hand, the Purkinje-molecular layer, corresponding to the output layer of the cerebellar cortex, is composed of the inhibitory PCs and the inhibitory BCs (basket cells). The MF



**Fig. 2.** Cerebellar ring network. (a) Box diagram for the cerebellar network. Lines with triangles and circles denote excitatory and inhibitory synapses, respectively. GR (granule cell), GO (Golgi cell), and PF (parallel fiber) in the granular layer, PC (Purkinje cell) and BC (basket cell) in the Purkinje-molecular layer, and other parts for VN (vestibular nuclei), IO(inferior olive), PCN(pre-cerebellar nuclei), MF (mossy fiber), and CF (climbing fiber). (b) Schematic diagram for granular-layer ring network with concentric inner GR and outer GO rings. Numbers represent granular layer zones (bounded by dotted lines) for  $N_C = 32$ . In each *I*th zone ( $I = 1, ..., N_C$ ), there exists the *I*th GR cluster on the inner GR ring. Each GR cluster consists of GR cells (solid circles), and it is bounded by 2 glomeruli (stars). On the outer GO ring in the *I*th zone, there exists the *I*th GO cell (diamonds). (c) Schematic diagram for Purkinje-molecular-layer ring network with concentric inner PC and outer BC rings. Numbers represent the Purkinje-molecular-layer zones (bounded by dotted lines) for  $N_{PC} = 16$ . In each *J*th zone, there exist the *J*th PC (solid circle) on the inner PC ring and the *J*th BC (solid triangle) on the outer BC ring.

context signal for the post-eye-movement is fed from the PCN (pre-cerebellar nuclei) to the GR cells. They are diversely recoded via inhibitory coordination of GO cells on GR cells in the granular layer. Then, these diversely-recoded outputs are fed via PFs to the PCs and the BCs in the Purkinje-molecular layer.

The PCs receive another excitatory error-teaching CF signals from the IO, along with the inhibitory inputs from BCs. Then, depending on the type of PF signals (i.e., in-phase or out-of-phase PF signals), diverse PF (student) signals are effectively depressed by the (in-phase) error-teaching (instructor) CF signals. Such effective depression at PF-PC synapses causes a large modulation in firing activities of PCs (principal output cells in the cerebellar cortex). Then, the VN neuron generates the final output of the cerebellum (i.e., it evokes OKR eye-movement) through receiving both the inhibitory inputs from the PCs and the excitatory inputs via MFs. This VN neuron also provides inhibitory inputs for the realized eye-movement to the IO neuron which also receives the excitatory desired signals for a desired eye-movement from the PCN. Then, the IO neuron supplies excitatory error-teaching CF signals to the PCs.

Fig. 2(b) shows a schematic diagram for the granular-layer ring network with concentric inner GR and outer GO rings. Numbers represent granular-layer zones (bounded by dotted lines). That is, the numbers 1, 2, ..., and  $N_C$  denote the 1st, the 2nd, ..., and the  $N_{\rm C}$ th granular-layer zones, respectively. Hence, the total number of granular-layer zones is  $N_C$ ; Fig. 2(b) shows an example for  $N_C = 32$ . In each *I*th zone  $(I = 1, ..., N_C)$ , there exists the *I*th GR cluster on the inner GR ring. Each GR cluster consists of  $N_{GR}$ excitatory GR cells (solid circles). Then, location of each GR cell may be represented by the two indices (I, i) which represent the *i*th GR cell in the *I*th GR cluster, where  $i = 1, ..., N_{GR}$ . Here, we consider the case of  $N_{\rm C} = 2^{10}$  and  $N_{\rm GR} = 50$ , and hence the total number of GR cells is 51,200. In this case, the Ith zone covers the angular range of  $(I - 1) \theta_{GR}^* < \theta < I \theta_{GR}^* (\theta_{GR}^* = 0.35^\circ)$ . On the outer GO ring in each Ith zone, there exists the Ith inhibitory GO cell (diamond), and hence the total number of GO cells is  $N_{\rm C}$ .

We note that each GR cluster is bounded by 2 glomeruli (corresponding to the axon terminals of the MFs) (stars). GR cells within each GR cluster share the same inhibitory and excitatory synaptic inputs through their dendrites which contact the two glomeruli at both ends of the GR cluster. Each glomerulus receives inhibitory inputs from nearby 81 (clockwise side: 41 and counter-clockwise side: 40) GO cells with a random connection probability  $p_c$  (= 0.06). Hence, on average, about 5 GO cell axons innervate each glomerulus. Thus, each GR cell receives about 10 inhibitory inputs through 2 dendrites which synaptically contact the glomeruli at both boundaries. In this way, each GR cell in the GR cluster shares the same inhibitory synaptic inputs from nearby GO cells through the intermediate glomeruli at both ends.

Also, each GR cell shares the same two excitatory inputs via the two glomeruli at both boundaries, because a glomerulus receives an excitatory MF input. Here, we take into consideration stochastic variability of synaptic transmission from a glomerulus to GR cells, and supply independent Poisson spike trains with the same firing rate to each GR cell for the excitatory MF signals. In this GR-GO feedback system, each GO cell receives excitatory synaptic inputs via PFs from GR cells in the nearby 49 (central side: 1, clockwise side: 24 and counter-clockwise side: 24) GR clusters with a random connection probability 0.1. Thus, 245 PFs (i.e. GR cell axons) innervate a GO cell.

Fig. 2(c) shows a schematic diagram for the Purkinjemolecular-layer ring network with concentric inner PC and outer BC rings. Numbers denote the Purkinje-molecular-layer zones (bounded by dotted lines). In each *J*th zone  $(J = 1, ..., N_{PC})$ , there exist the Jth PC (solid circles) on the inner PC ring and the Jth BC (solid triangles) on the outer BC ring. Here, we consider the case of  $N_{PC} = 16$ , and hence the total numbers of PC and BC are 16, respectively. In this case, each Jth ( $J = 1, ..., N_{PC}$ ) zone covers the angular range of  $(J - 1) \theta_{PC}^* < \theta < J \theta_{PC}^*$ , where  $\theta_{PC}^* \simeq 22.5^\circ$  (corresponding to about 64 zones in the granular-layer ring network). We note that diversely-recoded PFs innervate PCs and BCs. Each PC (BC) in the *I*th Purkinje-molecular-layer zone receives excitatory synaptic inputs via PFs from all the GR cells in the 288 GR clusters (clockwise side: 144 and counter-clockwise side: 144 when starting from the angle  $\theta = (J - 1) \theta_{PC}^*$  in the granular-layer ring network). Thus, each PC (BC) is synaptically connected via PFs to the 14,400 GR cells (which corresponds to about 28% of the total GR cells). In addition to the PF signals, each PC also receives inhibitory inputs from nearby 3 BCs (central side: 1, clockwise side: 1 and counter-clockwise side: 1) and excitatory error-teaching CF signal from the IO.

Outside the cerebellar cortex, for simplicity, we consider just one VN neuron and one IO neuron. Both excitatory inputs via 100 MFs and inhibitory inputs from all the 16 PCs are fed into the VN neuron. Then, the VN neuron evokes the OKR eye-movement and supplies inhibitory input for the realized eye-movement to the IO neuron. One additional excitatory desired signal from the PCN is also fed into the IO neuron. Then, through integration of both excitatory and inhibitory inputs, the IO neuron provides excitatory error-teaching CF signals to the PCs.

### 2.3. Leaky integrate-and-fire neuron model with afterhyperpolarization current

As elements of the cerebellar ring network, we choose leaky integrate-and-fire (LIF) neuron models which incorporate additional afterhyperpolarization (AHP) currents that determine refractory periods (Gerstner & Kistler, 2002). This LIF neuron model is one of the simplest spiking neuron models. Due to its simplicity, it can be easily analyzed and simulated. Thus, it has been very popularly used as a neuron model.

The following equations govern dynamics of states of individual neurons in the X population:

$$C_X \frac{dv_i^{(X)}}{dt} = -I_{L,i}^{(X)} - I_{AHP,i}^{(X)} + I_{ext}^{(X)} - I_{syn,i}^{(X)}, \quad i = 1, \dots, N_X,$$
(3)

where  $N_X$  is the total number of neurons in the X population, X = GR and GO in the granular layer, X = PC and BC in the Purkinje-molecular layer, and in the other parts X = VN and IO. In Eq. (1),  $C_X$  (pF) represents the membrane capacitance of the cells in the X population, and the state of the *i*th neuron in the X population at a time t (ms) is characterized by its membrane potential  $v_i^{(X)}$  (mV). The time-evolution of  $v_i^{(X)}(t)$  is governed by 4 types of currents (pA) into the *i*th neuron in the X population; the leakage current  $I_{L,i}^{(X)}$ , the AHP current  $I_{AHP,i}^{(X)}$ , the external constant current  $I_{ext}^{(X)}$  (independent of *i*), and the synaptic current  $I_{syn,i}^{(X)}$ . We note that the equation for a single LIF neuron model

[without the AHP current and the synaptic current in Eq. (3)] describes a simple parallel resistor-capacitor (RC) circuit. Here, the leakage term is due to the resistor and the integration of the external current is due to the capacitor which is in parallel to the resistor. Thus, in Eq. (3), the 1st type of leakage current  $I_{L,i}^{(X)}$  for the *i*th neuron in the *X* population is given by:

$$I_{L,i}^{(X)} = g_L^{(X)} (v_i^{(X)} - V_L^{(X)}),$$
(4)

where  $g_L^{(X)}$  and  $V_L^{(X)}$  are conductance (nS) and reversal potential for the leakage current, respectively. When the membrane potential  $v_i^{(X)}$  reaches a threshold  $v_{th}^{(X)}$  at a time  $t_{f,i}^{(X)}$ , the *i*th neuron fires a spike. After firing (i.e.,  $t \ge t_{f,i}^{(X)}$ ), the 2nd type of AHP current  $I_{AHP,i}^{(X)}$  follows:

$$I_{AHP,i}^{(X)} = g_{AHP}^{(X)}(t) \left( v_i^{(X)} - V_{AHP}^{(X)} \right) \quad \text{for } t \ge t_{f,i}^{(X)}.$$
(5)

Here,  $V_{AHP}^{(X)}$  is the reversal potential for the AHP current, and the conductance  $g_{AHP}^{(X)}(t)$  is given by an exponential-decay function:

$$g_{AHP}^{(X)}(t) = \bar{g}_{AHP}^{(X)} e^{-(t-t_{f,i}^{(X)})/\tau_{AHP}^{(X)}},$$
(6)

where  $\bar{g}_{AHP}^{(X)}$  and  $\tau_{AHP}^{(X)}$  are the maximum conductance and the decay time constant for the AHP current. As  $\tau_{AHP}^{(X)}$  increases, the refractory period becomes longer.

The 3rd type of external constant current  $I_{ext}^{(X)}$  for the cellular spontaneous discharge is supplied to only the PCs and the VN neuron because of their high spontaneous firing rates (Häusser & Clark, 1997; Thach, 1968). In Appendix A, Table A.1 shows the parameter values for the capacitance  $C_X$ , the leakage current  $I_L^{(X)}$ , the AHP current  $I_{AHP}^{(X)}$ , and the external constant current  $I_{ext}^{(X)}$ . These values are adopted from physiological data (Yamazaki & Nagao, 2012; Yamazaki & Tanaka, 2007a).

### 2.4. Synaptic currents

The 4th type of synaptic current  $I_{syn,i}^{(X)}$  into the *i*th neuron in the X population consists of the following 3 kinds of synaptic currents:

$$I_{syn,i}^{(X)} = I_{AMPA,i}^{(X,Y)} + I_{NMDA,i}^{(X,Y)} + I_{GABA,i}^{(X,Z)}.$$
(7)

Here,  $I_{\text{AMPA},i}^{(X,Y)}$  and  $I_{\text{NMDA},i}^{(X,Y)}$  are the excitatory AMPA ( $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid) receptor-mediated and NMDA (N-methyl-D-aspartate) receptor-mediated currents from the pre-synaptic source Y population to the post-synaptic *i*th neuron in the target X population. On the other hand,  $I_{GABA,i}^{(\Lambda,L)}$ is the inhibitory GABA<sub>A</sub> ( $\gamma$ -aminobutyric acid type A) receptormediated current from the pre-synaptic source Z population to the post-synaptic *i*th neuron in the target X population.

Similar to the case of the AHP current, the R (= AMPA, NMDA, or GABA) receptor-mediated synaptic current  $I_{R,i}^{(T,S)}$  from the presynaptic source *S* population to the *i*th post-synaptic neuron in the target *T* population is given by:

$$I_{R,i}^{(T,S)} = g_{R,i}^{(T,S)}(t) (v_i^{(T)} - V_R^{(S)}),$$
(8)

where  $g_{(R,i)}^{(T,S)}(t)$  and  $V_R^{(S)}$  are synaptic conductance and synaptic reversal potential (determined by the type of the pre-synaptic source *S* population), respectively. We get the synaptic conductance  $g_{R,i}^{(T,S)}(t)$  from:

$$g_{R,i}^{(T,S)}(t) = \bar{g}_R^{(T)} \sum_{j=1}^{N_S} J_{ij}^{(T,S)} w_{ij}^{(T,S)} s_j^{(T,S)}(t),$$
(9)

where  $\bar{g}_{R}^{(T)}$  and  $J_{ij}^{(T,S)}$  are the maximum conductance and the synaptic weight of the synapse from the *j*th pre-synaptic neuron in the source *S* population to the *i*th post-synaptic neuron in the target *T* population, respectively. The inter-population synaptic connection from the source *S* population (with  $N_s$  neurons) to the target *T* population is given by the connection weight matrix  $W^{(T,S)}$  (= { $w_{ij}^{(T,S)}$ }) where  $w_{ij}^{(T,S)} = 1$  if the *j*th neuron in the source *S* population is pre-synaptic to the *i*th neuron in the target *T* population; otherwise  $w_{ij}^{(T,S)} = 0$ .

The post-synaptic ion channels are opened due to the binding of neurotransmitters (emitted from the source *S* population) to receptors in the target *T* population. The fraction of open ion channels at time *t* is represented by  $s^{(T,S)}$ . The time course of  $s_j^{(T,S)}(t)$  of the *j*th neuron in the source *S* population is given by a sum of exponential-decay functions  $E_R^{(T,S)}(t - t_j^{(j)})$ :

$$s_{j}^{(T,S)}(t) = \sum_{f=1}^{F_{j}^{(S)}} E_{R}^{(T,S)}(t - t_{f}^{(j)}),$$
(10)

where  $t_f^{(j)}$  and  $F_j^{(s)}$  are the *f* th spike time and the total number of spikes of the *j*th neuron in the source *S* population, respectively. The exponential-decay function  $E_R^{(T,S)}(t)$  (which corresponds to contribution of a pre-synaptic spike occurring at t = 0 in the absence of synaptic delay) is given by:

$$E_{R}^{(T,S)}(t) = e^{-t/\tau_{R}^{(T)}} \Theta(t) \quad \text{or}$$
(11a)

$$= (A_1 e^{-t/\tau_{R,1}} + A_2 e^{-t/\tau_{R,2}})\Theta(t),$$
(11b)

where  $\Theta(t)$  is the Heaviside step function:  $\Theta(t) = 1$  for  $t \ge 0$  and 0 for t < 0. Depending on the source and the target populations,  $E_R^{(T,S)}(t)$  may be a type-1 single exponential-decay function of Eq. (11a) or a type-2 dual exponential-decay function of Eq. (11b). In the type-1 case, there exists one synaptic decay time constant  $\tau_R^{(T)}$  (determined by the receptor on the post-synaptic target *T* population), while in the type-2 case, two synaptic decay time constants,  $\tau_{R,1}^{(T)}$  and  $\tau_{R,2}^{(T)}$  exist. In most cases, the type-1 single exponential-decay function of Eq. (11a) appears, except for the two synaptic currents  $I_{GABA}^{(CR,GO)}$  and  $I_{NMDA}^{(CO,CR)}$ . In Appendix A, Tables A.2, A.3, and A.4 show the parameter where for the two synaptic decay target  $\tau_{R,2}^{(T)}$  and  $\tau_{R,2}^{(T)}$  exponential-decay function of Eq. (11a) appears, except for the two synaptic currents  $I_{GABA}^{(CR,GO)}$  and  $I_{NMDA}^{(CO,CR)}$ .

In Appendix A, Tables A.2, A.3, and A.4 show the parameter values for the maximum conductance  $\bar{g}_{R}^{(T)}$ , the synaptic weight  $J_{ij}^{(T,S)}$ , the synaptic reversal potential  $V_{R}^{(S)}$ , the synaptic decay time constant  $\tau_{R}^{(T)}$ , and the amplitudes  $A_{1}$  and  $A_{2}$  for the type-2 exponential-decay function in the granular layer, the Purkinje-molecular layer, and the other parts for the VN and IO, respectively. These values are adopted from physiological data (Yamazaki & Nagao, 2012; Yamazaki & Tanaka, 2007a).

### 2.5. Synaptic plasticity

We use a rule for synaptic plasticity, based on the experimental result in Safo and Regehr (2008). This rule is a refined one for the LTD in comparison to the rule used in Yamazaki and Nagao (2012) and Yamazaki and Tanaka (2007a), the details of which will be explained below.

The coupling strength of the synapse from the pre-synaptic neuron j in the source S population to the post-synaptic neuron *i* in the target *T* population is  $J_{ij}^{(T,S)}$ . Initial synaptic strengths are given in Tables A.2, A.3, and A.4. Here, we assume that learning occurs only at the PF-PC synapses. Hence, only the synaptic strengths  $J_{ij}^{(PC,PF)}$  of PF-PC synapses may be modifiable, while synaptic strengths of all the other synapses are static. [Here, the index *j* for the PFs corresponds to the two indices (M, m) for GR cells representing the *m*th (1 < m < 50) cell in the *M*th (1 < M < 50)2<sup>10</sup>) GR cluster.] Synaptic plasticity at PF-PC synapses have been so much studied in diverse experimental (Chen & Thompson, 1995: Coesmans. Weber. De Zeeuw. & Hansel. 2004: De Schutter. 1995: Gallimore, Kim, Tanaka-Yamamoto, & De Schutter, 2018: Ito, 1989: Ito & Kano, 1982: Ito et al., 1982: Lev-Ram, Mehta, Kleinfeld, & Tsien, 2003; Molnár, 2014; Safo & Regehr, 2008; Sakurai, 1987; Steuber et al., 2007; Wang, Denk, & Häusser, 2000; Yang & Lisberger, 2014) and computational (Achard & De Schutter, 2008; Albus, 1971; Bouvier et al., 2018; Buonomano & Mauk, 1994; Gerstner & van Hemmen, 1992; Kenvon, Medina, & Mauk, 1998; Medina, Garcia, Nores, Taylor, & Mauk, 2000; Roberts, 2007; Yamazaki & Nagao, 2012; Yamazaki & Tanaka, 2007a) works.

With increasing time t, synaptic strength for each PF-PC synapse is updated with the following multiplicative rule (depending on states) (Safo & Regehr, 2008):

$$J_{ij}^{(PC,PF)}(t) \to J_{ij}^{(PC,PF)}(t) + \Delta J_{ij}^{(PC,PF)}(t),$$
(12)

where

$$\Delta J_{ij}^{(\text{PC,PF})}(t) = \Delta \text{LTD}_{ij}^{(1)} + \Delta \text{LTD}_{ij}^{(2)} + \Delta \text{LTP}_{ij}, \qquad (13)$$

$$\Delta \text{LTD}_{ij}^{(1)} = -\delta_{\text{LTD}} \cdot J_{ij}^{(\text{PC,PF})}(t) \cdot CF_i(t) \cdot \sum_{\Delta t=0}^{-r_i} \Delta J_{\text{LTD}}(\Delta t), \quad (14)$$

$$\Delta \text{LTD}_{ij}^{(2)} = -\delta_{LTD} \cdot J_{ij}^{(\text{PC,PF})}(t) \cdot [1 - CF_i(t)] \cdot PF_{ij}(t) \cdot D_i(t)$$
$$\times \sum_{\Delta t=0}^{\Delta t_i^*} \Delta J_{LTD}(\Delta t), \qquad (15)$$

$$\Delta \text{LTP}_{ij} = \delta_{LTP} \cdot [J_0^{(\text{PC},\text{PF})} - J_{ij}^{(\text{PC},\text{PF})}(t)] \cdot [1 - CF_i(t)] \cdot PF_{ij}(t) \\ \times [1 - D_i(t)].$$
(16)

Here,  $J_0^{(PC,PF)}$  is the initial value (= 0.006) for the synaptic strength of PF-PC synapses. Synaptic modification (LTD or LTP) occurs, depending on the relative time difference  $\Delta t = t_{CF}$  (CF activation time) -  $t_{PF}$  (PF activation time)] between the spiking times of the error-teaching instructor CF and the diversely-recoded student PFs. In Eqs. (14)–(16),  $CF_i(t)$  represents a spike train of the CF signal coming into the *i*th PC. When  $CF_i(t)$  activates at a time t,  $CF_i(t) = 1$ ; otherwise,  $CF_i(t) = 0$ . This instructor CF firing causes LTD at PF-PC synapses in conjunction with earlier ( $\Delta t > 0$ ) student PF firings in the range of  $t_{CF} - \Delta t_r^* < t_{PF} < t_{CF}$  ( $\Delta t_r^* \simeq$ 277.5 ms), which corresponds to the major LTD in Eq. (14).

We next consider the case of  $CF_i(t) = 0$ , corresponding to Eqs. (15) and (16). Here,  $PF_{ij}(t)$  denotes a spike train of the PF signal from the *j*th pre-synaptic GR cell to the *i*th post-synaptic PC. When  $PF_{ij}(t)$  activates at time *t*,  $PF_{ij}(t) = 1$ ; otherwise,  $PF_{ij}(t) = 0$ . In the case of  $PF_{ij}(t) = 1$ , PF firing may give rise to LTD or LTP, depending on the presence of earlier CF firings in an effective range. If CF firings exist in the range of  $t_{PF} + \Delta t_i^* < t_{CF} < t_{PF}$  ( $\Delta t_i^* \simeq -117.5 \text{ ms}$ ),  $D_i(t) = 1$ ; otherwise  $D_i(t) = 0$ . When both  $PF_{ij}(t) = 1$  and  $D_i(t) = 1$ , the PF firing causes another LTD at PF-PC synapses in association with earlier ( $\Delta t < 0$ ) CF firings [see Eq. (15)]. The likelihood for occurrence of earlier CF firings



**Fig. 3.** Time window for the LTD at the PF-PC synapse. Plot of synaptic modification  $\Delta J_{LTD}(\Delta t)$  for LTD versus  $\Delta t$  [see Eq. (17)].

within the effective range is very low because mean firing rates of the CF signals (corresponding to output firings of individual IO neurons) are ~ 1.5 Hz (Llinás, 2014; Mathy et al., 2009). Hence, this 2nd type of LTD is a minor one. On the other hand, in the case of  $D_i(t) = 0$  (i.e., absence of earlier associated CF firings), LTP occurs due to the PF firing alone [see Eq. (16)]. The update rate  $\delta_{LTD}$  for LTD in Eqs. (14) and (15) is 0.005, while the update rate  $\delta_{LTP}$  for LTP in Eqs. (16) is 0.0005 (=  $\delta_{LTD}/10$ ).

In the case of LTD in Eqs. (14) and (15), the synaptic modification  $\Delta J_{LTD}(\Delta t)$  varies depending on the relative time difference  $\Delta t$  (=  $t_{CF} - t_{PF}$ ). We employ the following time window for the synaptic modification  $\Delta J_{LTD}(\Delta t)$  (Safo & Regehr, 2008):

$$\Delta J_{LTD}(\Delta t) = A + B \cdot e^{-(\Delta t - t_0)^2 / \sigma^2},$$
(17)

where A = -0.12, B = 0.4,  $t_0 = 80$ , and  $\sigma = 180$ . Fig. 3 shows the time window for  $\Delta J_{LTD}(\Delta t)$ . As shown well in Fig. 3, LTD occurs in an effective range of  $\Delta t_l^* < \Delta t < \Delta t_r^*$ . We note that a peak exists at  $t_0 = 80$  ms, and hence peak LTD occurs when PF firing precedes CF firing by 80 ms. A CF firing causes LTD in conjunction with earlier PF firings in the black region ( $0 < \Delta t < \Delta t_r^*$ ), and it also gives rise to another LTD in association with later PF firings in the gray region ( $\Delta t_l^* < \Delta t < 0$ ). The effect of CF firing on earlier PF firings is much larger than that on later PF firings. However, outside the effective range (i.e.,  $\Delta t > \Delta t_r^*$  or  $< \Delta t_l^*$ ), PF firings alone lead to LTP, due to absence of effectively associated CF firings.

Finally, we discuss the advantages of our refined rule for synaptic plasticity in comparison to the synaptic rule in Yamazaki and Nagao (2012) and Yamazaki and Tanaka (2007a). Our rule is constructed from the experimental result in Safo and Regehr (2008). In the presence of a CF firing, a major LTD ( $\Delta$ LTD<sup>(1)</sup>) takes place in association with earlier PF firings in the range of  $t_{\rm CF} - \Delta t_r^* < t_{\rm PF} < t_{\rm CF}$  ( $\Delta t_r^* \simeq 277.5$  ms), while a minor LTD  $(\Delta LTD^{(2)})$  occurs in association with later PF firings in the range of  $t_{CF} < t_{PF} < t_{CF} - \Delta t_l^*$  ( $\Delta t_l^* \simeq -117.5$  ms). The magnitude of LTD changes depending on  $\Delta t$  (=  $t_{CF} - t_{PF}$ ); a peak LTD occurs for  $\Delta t = 80$  ms. On the other hand, the rule in Yamazaki and Nagao (2012) and Yamazaki and Tanaka (2007a) considers only the major LTD in conjunction with earlier PF firings in the range of  $t_{\rm CF}$  – 50  $< t_{\rm PF} < t_{\rm CF}$ , the magnitude of major LTD is equal, independently of  $\Delta t$ , and minor LTD in association with later PF firings is not considered. Outside the effective range of LTD, PF firings alone result in LTP in both rules. However, we also note that some features of OKR were successfully reproduced by using the simple synaptic rule with only the major LTD in Yamazaki and Nagao (2012) and Yamazaki and Tanaka (2007a).

### 2.6. Numerical method for integration

Numerical integration of the governing equation (3) for the time-evolution of states of individual neurons, along with the update rule for synaptic plasticity of Eq. (12), is done by employing the 2nd-order Runge–Kutta method with the time step 1 ms. For each realization, we choose random initial points  $v_i^{(X)}(0)$  for the *i*th neuron in the *X* population with uniform probability in the range of  $v_i^{(X)}(0) \in (V_L^{(X)} - 5.0, V_L^{(X)} + 5.0)$ ; the values of  $V_L^{(X)}$  are given in Table A.1.

## 3. Effect of diverse spiking patterns of GR clusters on motor learning for the OKR adaption

In this section, we study the effect of diverse recoding of GR cells on motor learning for the OKR adaptation by varying the connection probability  $p_c$  from the GO to the GR cells. We mainly consider an optimal case of  $p_c^* = 0.06$  where the spiking patterns of GR clusters are the most diverse. In this case, we first make dynamical classification of diverse spiking patterns of the GR clusters. Then, we make an intensive investigation on the effect of diverse recoding of GR cells on synaptic plasticity at PF-PC synapses and the subsequent learning process in the PC-VN-IO system. Finally, we vary  $p_c$  from the optimal value  $p_c^*$ , and study dependence of the diversity degree  $\mathcal{D}$  of spiking patterns and the saturated learning gain degree  $\mathcal{L}_g^*$  on  $p_c$ . Both  $\mathcal{D}$  and  $\mathcal{L}_g^*$  are found to form bell-shaped curves with peaks at  $p_c^*$ , and they have strong correlation with the Pearson's coefficient  $r \simeq 0.9998$ . As a result, the more diverse the recoding of GR cells, the more effective the motor learning for the OKR adaptation.

### 3.1. Firing activity in the whole population of GR cells

As shown in Fig. 2, recoding process is performed in the granular layer (corresponding to the input layer of the cerebellar cortex), consisting of GR and GO cells. In the GR-GO feedback system, GR cells (principal output cells in the granular layer) receive excitatory context signals for the post-eye-movement via the sinusoidally-modulating MFs [see Fig. 1(b1)] and make recoding of context signals. In this recoding process, GO cells make effective inhibitory coordination for diverse recoding of GR cells. Thus, diversely recoded signals are fed into the PCs (principal output cells in the cerebellar cortex) via PFs. Due to this type of diverse recoding of GR cells, the cerebellum was recently reinterpreted as a liquid state machine with powerful discriminating/separating capability (i.e., different input signals are transformed into more different ones via recoding process) rather than the simple perceptron in the Marr-Albus-Ito theory (Maass, Natschläger, & Markram, 2002; Yamazaki & Tanaka, 2007b).

We first consider the firing activity in the whole population of GR cells for  $p_c^* = 0.06$ . Collective firing activity may be well visualized in the raster plot of spikes which is a collection of spike trains of individual neurons. Such raster plots of spikes are fundamental data in experimental neuroscience. As a population quantity showing collective firing behaviors, we use an instantaneous whole-population spike rate  $R_{GR}(t)$  which may be obtained from the raster plots of spikes (Brunel, 2000; Brunel & Hakim, 1999, 2008; Brunel & Hansel, 2006; Brunel & Wang, 2003; Geisler et al., 2005; Kim & Lim, 2014; Wang, 2010). To obtain a smooth instantaneous whole-population spike rate, we employ the kernel density estimation (kernel smoother) (Shimazaki & Shinomoto, 2010). Each spike in the raster plot is convoluted (or blurred) with a kernel function  $K_h(t)$  [such as a smooth Gaussian function in Eq. (19)], and then a smooth estimate of instantaneous whole-population spike rate  $R_{GR}(t)$  is obtained by averaging the



**Fig. 4.** Firing activity of GR cells in an optimal case of  $p_c$  (connection probability from GO to GR cells) = 0.06. (a) Raster plots of spikes of  $10^3$  randomly chosen GR cells. (b) Instantaneous whole-population spike rate  $R_{GR}(t)$  in the whole population of GR cells. Band width for  $R_{GR}(t)$ : h = 10 ms. Plots of the activation degrees (c1) A(t) in the whole population of GR cells and (c2)  $A^{(G)}(t)$  in the *G* spiking group [*G* : in-phase (solid curve) and out-of-phase (dotted curve)]. Plots of (d1) instantaneous individual firing rate  $f_{GR}^{(i)}(t)$  for the active GR cells and (d2) instantaneous population spike rate  $f_{GR}^{(p)}(t)$  in the whole population of GR cells. Bin size for (c1)-(d2):  $\Delta t = 10$  ms.

convoluted kernel function over all spikes of GR cells in the whole population:

$$R_{\rm GR}(t) = \frac{1}{N} \sum_{i=1}^{N} \sum_{s=1}^{n_i} K_h(t - t_s^{(i)}), \tag{18}$$

where  $t_s^{(i)}$  is the sth spiking time of the *i*th GR cell,  $n_i$  is the total number of spikes for the *i*th GR cell, and N is the total number of GR cells (i.e.,  $N = N_c \cdot N_{\text{GR}} = 51,200$ ). Here, we use a Gaussian kernel function of band width h:

$$K_{h}(t) = \frac{1}{\sqrt{2\pi}h} e^{-t^{2}/2h^{2}}, \quad -\infty < t < \infty.$$
(19)

Throughout the paper, the band width h of  $K_h(t)$  is 10 ms.

Fig. 4(a) shows a raster plot of spikes of  $10^3$  randomly chosen GR cells. At the initial and the final stage of the cycle, GR cells fire sparse spikes, because the firing rates of Poisson spikes for the MF are low. On the other hand, at the middle stage, the firing rates for the MF are relatively high, and hence spikes of GR cells become relatively dense. Fig. 4(b) shows the instantaneous whole-population spike rate  $R_{GR}(t)$  in the whole population of GR cells.  $R_{GR}(t)$  is basically in proportion to the sinusoidally-modulating inputs via MFs. However, it has a different waveform with a central plateau. At the initial stage, and at the final stage, it decreases slowly. In comparison to the MF signal, the top part of  $R_{GR}(t)$  becomes lowered and flattened, due to the effect of inhibitory GO cells. Thus, a central plateau emerges.

We next consider the activation degree of GR cells. To examine it, we divide the whole learning cycle (2000 ms) into 200 bins (bin size: 10 ms). Then, we get the activation degree  $A_i$  for the active GR cells in the *i*th bin:

$$A_i = \frac{N_{a,i}}{N},\tag{20}$$

where  $N_{a,i}$  and N are the number of active GR cells in the *i*th bin and the total number of GR cells, respectively. Fig. 4(c1) shows a plot of the activation degree A(t) in the whole population of GR cells. It is nearly symmetric, and has double peaks with a central valley at the middle stage; its values at both peaks are about 0.94 and the central minimum value is about 0.65.

Presence of the central valley in A(t) is in contrast to the central plateau in  $R_{GR}(t)$ . Appearance of such a central valley may

be understood as follows: The whole population of GR cells can be decomposed into two types of in-phase and out-of-phase spiking groups. Spiking patterns of in-phase (out-of-phase) GR cells are in-phase (out-of-phase) with respect to  $R_{GR}(t)$  (representing the population-averaged firing activity in the whole population of GR cells); details will be given in Figs. 5 and 6. Then, the activation degree  $A_i^{(G)}$  of active GR cells in the *G* spiking group in the *i*th bin is given by:

$$A_i^{(G)} = \frac{N_{a,i}^{(G)}}{N},$$
(21)

where  $N_{a,i}^{(G)}$  is the number of active GR cells in the *G* spiking group  $(G = i \text{ and } o \text{ for the in-phase and the out-of-phase spiking groups, respectively) in the$ *i* $th bin. The sum of <math>A_i^{(G)}(t)$  over the in-phase and the out-of-phase spiking groups is just the activation degree  $A_i(t)$  in the whole population. Fig. 4(c2) shows plots of activation degree  $A^{(G)}(t)$  in the in-phase (solid line) and the out-of-phase (dotted curve) spiking groups. In the case of in-phase (G = i) spiking group,  $A^{(i)}(t)$  has a central plateau, while  $A^{(o)}(t)$  has double peaks with a central valley in the case of out-of-phase (G = o) spiking group. Hence, small contribution of out-of-phase spiking group at the middle stage leads to emergence of the central valley in A(t) in the whole population.

We note again that, in the whole population the activation degree A(t) with a central valley is in contrast to  $R_{GR}(t)$  with a central plateau. To understand this discrepancy, we consider the bin-averaged instantaneous individual firing rates  $f_{GR}^{(i)}$  of active GR cells:

$$f_{\rm GR}^{(i)} = \frac{N_{\rm s,i}}{N_{a,i} \ \Delta t},\tag{22}$$

where  $N_{s,i}$  is the number of spikes of GR cells in the *i*th bin,  $N_{a,i}$  is the number of active GR cells in the *i*th bin, and the bin size  $\Delta t$  is 10 ms. Fig. 4(d1) shows a plot of  $f_{GR}^{(i)}(t)$  for the active GR cells. We note that active GR cells fire spikes at higher firing rates at the middle stage because  $f_{GR}^{(i)}(t)$  has a central peak. Then, the bin-averaged instantaneous population spike rate  $f_{GR}^{(p)}$  is given by the product of the activation degree  $A_i$  of Eq. (20) and the instantaneous individual firing rate  $f_{GR}^{(i)}$  of Eq. (22):

$$f_{\rm GR}^{(p)} = A_i f_{\rm GR}^{(i)} = \frac{N_{\rm s,i}}{N \ \Delta t}.$$
(23)



**Fig. 5.** Diverse spiking patterns in the GR clusters in the optimal case of  $p_c^* = 0.06$ . Raster plots of spikes and instantaneous cluster spike rates  $R_{GR}^{(I)}(t)$  for diverse spiking patterns. Five in-phase spiking patterns in the *I*th GR clusters; I = (a1) 594, (a2) 543, (a3) 663, (a4) 332, and (a5) 399. Two anti-phase spiking patterns in the *I*th GR cluster; I = (b1) 49 and (b2) 101. Six complex out-of-phase spiking patterns in the *I*th GR clusters; I = (c1) 192, (c2) 91, (c3) 773, (c4) 382, (c5) 705, and (c6) 349.  $C^{(I)}$  represents the conjunction index of the spiking pattern in the *I*th GR cluster.



**Fig. 6.** Characterization of diverse spiking patterns in the GR clusters in the optimal case of  $p_c^* = 0.06$ . (a) Distribution of conjunction indices  $\{c^{(l)}\}$  for the GR clusters in the whole population. (b) Fraction of spiking groups. Distribution of conjunction indices  $\{c^{(l)}\}$  for the (c1) in-phase, (c2) anti-phase, and (c3) complex out-of-phase spiking groups. Bin size for the histograms in (a) and in (c1)–(c3) is 0.1. (d) Bar diagram for the ranges of conjunction indices  $\{c^{(l)}\}$  for the in-phase, anti-phase, and complex out-of-phase spiking groups.

The instantaneous population spike rate  $f_{GR}^{(p)}(t)$  in Fig. 4(d2) has a central plateau, as in the case of  $R_{GR}(t)$ . We note that both  $f_{GR}^{(p)}(t)$  and  $R_{GR}(t)$  correspond to bin-based estimate and kernel-based smooth estimate for the instantaneous whole-population spike rate for the GR cells, respectively (Kim & Lim, 2014). In this way, although the activation degree A(t) of GR cells are lower

at the middle stage, their population spike rate becomes nearly the same as that in the neighboring parts (i.e., central plateau is formed), due to the higher individual firing rates.

### 3.2. Dynamical classification of spiking patterns of GR clusters

There are  $N_C$  (= 2<sup>10</sup>) GR clusters.  $N_{GR}$  (= 50) GR cells in each GR cluster share the same inhibitory and excitatory inputs via their dendrites which synaptically contact the two glomeruli (i.e., terminals of MFs) at both ends of the GR cluster [see Fig. 2(b)]; nearby inhibitory GO cell axons innervate the two glomeruli. Hence, GR cells in each GR cluster show similar firing behaviors. Similar to the case of  $R_{GR}(t)$  in Eq. (18), the firing activity of the *I*th GR cluster is characterized in terms of its instantaneous cluster spike rate  $R_{GR}^{(I)}(I = 1, ..., N_C)$ :

$$R_{GR}^{(l)}(t) = \frac{1}{N_{GR}} \sum_{i=1}^{N_{GR}} \sum_{s=1}^{N_{GR}} K_h(t - t_s^{(l,i)}),$$
(24)

where  $t_s^{(l,i)}$  is the sth spiking time of the *i*th GR cell in the *I*th GR cluster and  $n_i^{(l)}$  is the total number of spikes for the *i*th GR cell in the *I*th GR cluster.

We introduce the conjunction index  $C^{(l)}$  of each GR cluster, representing the degree for the conjunction (association) of the spiking behavior  $[R_{GR}^{(l)}(t)]$  of each *I*th GR cluster with that of the whole population  $[R_{GR}(t)$  in Fig. 4(b)] [i.e., denoting the degree for the resemblance (similarity) between  $R_{GR}^{(l)}(t)$  and  $R_{GR}(t)$ ]. The conjunction index  $C^{(l)}$  is given by the cross-correlation at the zero-time lag [i.e.,  $Corr_{GR}^{(l)}(0)$ ] between  $R_{GR}^{(l)}(t)$  and  $R_{GR}(t)$ :

$$Corr_{GR}^{(I)}(\tau) = \frac{\Delta R_{GR}(t+\tau)\Delta R_{GR}^{(I)}(t)}{\sqrt{\Delta R_{GR}^2(t)}\sqrt{\Delta R_{GR}^{(I)}^2(t)}},$$
(25)

where  $\Delta R_{GR}(t) = R_{GR}(t) - \overline{R_{GR}(t)}$ ,  $\Delta R_{GR}^{(I)}(t) = R_{GR}^{(I)}(t) - \overline{R_{GR}^{(I)}(t)}$ , and the overline denotes the time average over a cycle. We note

that  $C^{(l)}$  represents well the phase difference (shift) between the spiking patterns  $[R_{GR}^{(l)}(t)]$  of GR clusters and the firing behavior  $[R_{GR}(t)]$  in the whole population.

In all the 2<sup>10</sup> GR clusters, we obtain their conjunction indices  $C^{(l)}$ , make intensive examination of the phase difference of  $R_{GR}^{(l)}(t)$  with respect to  $R_{GR}(t)$ , and thus classify the whole GR clusters into the in-phase, the anti-phase, and the complex out-of-phase spiking groups. Fig. 5 shows examples for diverse spiking patterns of GR clusters. This type of diversity arises from inhibitory coordination of GO cells on the firing activity of GR cells in the GR-GO feedback system in the granular layer.

Five examples for "in-phase" spiking patterns in the *I*th (I =594, 543, 663, 332, and 399) GR clusters are given in Figs. 5(a1)-5(a5), respectively. Raster plot of spikes of  $N_{GR}(=50)$  GR cells and the corresponding instantaneous cluster spike rate  $R_{CR}^{(l)}(t)$ are shown, along with the value of  $C^{(I)}$  in each case of the *I*th GR cluster. In all these cases, the instantaneous cluster spike rates  $R_{GR}^{(l)}(t)$  are in-phase relative to the instantaneous wholepopulation spike rate  $R_{GR}(t)$ . Among them, in the case of I = 594 with the maximum conjunction index  $C_{max}$  (= 0.85),  $R_{GR}^{(594)}(t)$  with a central plateau is the most similar (in-phase) to  $R_{GR}(t)$ . In the next case of I = 543 with  $C^{(I)} = 0.48$ ,  $R_{GR}^{(543)}(t)$  has a central sharp peak, and hence its similarity degree relative to  $R_{CR}(t)$  decreases. The remaining two cases of I = 663 and 332 (with more than one central peaks) may be regarded as ones developed from the case of I = 594. With increasing the number of peaks in the central part, the value of  $C^{(l)}$  decreases, and hence the resemblance degree relative to  $R_{GR}(t)$  is reduced. The final case of I = 399 with double peaks can be considered as one evolved form the case of I = 543. In this case, the value of  $C^{(I)}$  is reduced to 0.40.

Based on the examples in Figs. 5(a1)–5(a5), spiking patterns which have central plateau, central sharp peak, and two or more central peaks in the middle part of cycle are considered as in-phase spiking patterns relative to the instantaneous whole-population spike rate  $R_{GR}(t)$ . We make an intensive examination of the instantaneous cluster spike rates  $R_{GR}^{(I)}$  of the GR clusters with  $C^{(I)} < 0.40$ , and determine the higher threshold  $C_h^*$  ( $\simeq 0.39$ ) between the in-phase and the complex out-of-phase spiking patterns. For  $C^{(I)} > C_h^*$  in-phase spiking patterns such as ones in Figs. 5(a1)–5(a5) appear. On the other hand, when passing the higher threshold  $C_h^*$  from the above, complex out-of-phase spiking patterns (with  $C^{(I)} < C_h^*$ ) emerge. These complex out-of-phase spiking patterns have left-skewed (right-skewed) peaks near the 1st (3rd) quartile of cycle [i.e., near t = 500 (1500) ms], explicit examples of which will be given below in Figs. 5(c1)–5(c4). Thus, all the GR clusters, exhibiting in-phase spiking patterns, constitute the in-phase spiking group where the range of  $C^{(I)}$  is  $(C_h^*, C_{max})$ ;  $C_{max} = 0.85$  and  $C_h^* \simeq 0.39$ .

Next, we consider the "anti-phase" spiking patterns. Two examples for the anti-phase spiking patterns in the *I*th (I = 49 and 101) GR clusters are given in Figs. 5(b1) and 5(b2), respectively. We note that, in both cases, the instantaneous cluster spike rates  $R_{GR}^{(I)}(t)$  are anti-phase with respect to  $R_{GR}(t)$  in the whole population. In the case of I = 49 with the minimum conjunction index  $C_{min}$  (= -0.57),  $R_{GR}^{(49)}(t)$  is the most anti-phase relative to  $R_{GR}(t)$ , and it has double peaks near the 1st and the 3rd quartiles and a central deep valley at the middle of the cycle. The case of I = 101 with  $C^{(I)}$  (= -0.23) may be regarded as evolved from the case of I = 49. It has an increased (but still negative) value of  $C^{(I)}$  (= -0.23) due to the risen central shallow valley.

Based on the examples in Figs. 5(b1)-5(b2), spiking patterns  $[R_{GR}^{(l)}(t)]$  which have double peaks near the 1st and the 3rd quartile and a central valley at the middle of cycle are regarded as antiphase spiking patterns with respect to  $R_{GR}(t)$ . Like the case of the above in-phase spiking patterns, through intensive examination

of  $R_{GR}^{(l)}$  of the GR clusters with  $C^{(l)} > -0.23$ , we determine the lower threshold  $C_l^*$  ( $\simeq -0.20$ ) between the anti-phase and the complex out-of-phase spiking patterns. For  $C^{(l)} < C_l^*$  antiphase spiking patterns such as ones in Figs. 5(b1)–5(b2) exist. In contrast, when passing the lower threshold  $C_l^*$  from the below, complex out-of-phase spiking patterns (with  $C^{(l)} > C_l^*$ ) appear. These complex out-of-phase spiking patterns have a central peak which is transformed from the central valley at the middle of cycle, along with double peaks near the 1st and the 3rd quartile. An explicit example will be given below in Fig. 5(c5). Thus, all the GR clusters, showing anti-phase spiking patterns, form the anti-phase spiking group where the range of  $C^{(l)}$  is  $(C_{min}, C_l^*)$ ;  $C_{min} = -0.57$  and  $C_l^* \simeq -0.20$ .

As discussed above, in the range of  $C_l^*$  ( $\simeq -0.20$ )  $< C^{(l)} <$  $C_{h}^{*}$  ( $\simeq$  0.39), a 3rd type of complex "out-of-phase" spiking patterns appear between the in-phase and the anti-phase spiking patterns. Figs. 5(c1)-5(c6) show six examples for the complex out-of-phase spiking patterns in the *I*th (I = 192, 91, 773, 382, 705, and 349) GR clusters. The cases of I = 192 and 91 seem to be developed from the in-phase spiking pattern in the *I*th (I = 594 or 543) GR cluster. In the case of I = 192,  $R_{GR}^{(192)}$  has a left-skewed peak near the 1st quartile of the cycle, while in the case of I = 91, (91)  $\hat{R}_{CR}^{(91)}$  has a right-skewed peak near the 3rd quartile. Hence, the values of  $C^{(I)}$  for I = 192 and 91 are reduced to 0.12 and 0.10, respectively. In the next two cases of I = 773 and 382, they seem to be developed from the cases of I = 192 and 91, respectively. The left-skewed (right-skewed) peak in the case of I = 192 (91) is bifurcated into double peaks, which leads to more reduction of conjunction indices;  $C^{(773)} = 0.08$  and  $C^{(382)} = 0.05$ . In the remaining case of I = 705, it seems to be evolved from the antiphase spiking pattern in the *I* = 101 case. The central valley for I = 101 is transformed into a central peak. Thus,  $R_{GR}^{(705)}$  has three peaks, and its value of  $C^{(705)}$  is a little increased to -0.18. As  $C^{(I)}$  is more increased toward the zero,  $R_{GR}^{(I)}$  becomes more complex, as shown in Fig. 5(cf6) in the zero, cf. 240 with  $\sigma^{(349)}_{GR} = 0.07$ . as shown in Fig. 5(c6) in the case of I = 349 with  $C^{(349)} = -0.07$ .

Results on characterization of the diverse in-phase, anti-phase, and complex out-of-phase spiking patterns are shown in Fig. 6. Fig. 6(a) shows the plot of the fraction of conjunction indices  $C^{(I)}$  in the whole GR clusters.  $C^{(I)}$  increases slowly from the negative to the peak at 0.55, and then it decreases rapidly. For this distribution { $C^{(I)}$ }, the range is (-0.57, 0.85), the mean is 0.32, and the standard deviation is 0.516. Then, the diversity degree D for the spiking patterns [ $R^{(I)}_{GR}(t)$ ] of all the GR clusters is given by:

### $\mathcal{D}$ = Relative Standard Deviation for the Distribution { $\mathcal{C}^{(l)}$ }, (26)

where the relative standard deviation is just the standard deviation divided by the mean. In the optimal case of  $p_c^* = 0.06$ ,  $\mathcal{D}^* \simeq 1.613$ , which is just a quantitative measure for the diverse recoding performed via feedback cooperation between the GR and the GO cells in the granular layer. As will be seen later in Fig. 18(b) for the plot of  $\mathcal{D}$  versus  $p_c$ ,  $\mathcal{D}^*$  is just the maximum, and hence spiking patterns of GR clusters at  $p_c^*$  is the most diverse.

We decompose the whole GR clusters into the in-phase, antiphase, and complex out-of-phase spiking groups. Fig. 6(b) shows the fraction of spiking groups. The in-phase spiking group with  $C_h^*$  ( $\simeq 0.39$ )  $< C^{(l)} < C_{max}$  (= 0.85) is a major one with fraction 50.2%, while the anti-phase spiking group with  $C_{min}$  (= -0.57)  $< C^{(l)} < C_l^*$  ( $\simeq -0.20$ ) is a minor one with fraction 5.8%. Between them ( $C_l^* < C_l^{(l)} < C_h^*$ ), the complex out-of-phase spiking group with fraction 44% exists. In this case, the spiking-group ratio, given by the ratio of the fraction of the in-phase spiking group to that of the out-of-phase spiking group (consisting of both the anti-phase and complex out-of-phase spiking groups), is  $\mathcal{R}^* \simeq 1.008$ . Thus, in the optimal case of  $p_c^* = 0.06$ , the fractions between the in-phase and the out-of-phase spiking groups are well balanced. Under such good balance between the in-phase



**Fig. 7.** Dynamical origin for the diverse spiking patterns in the GR clusters in the optimal case of  $p_c^* = 0.06$ . In-phase spiking patterns for I = (a1) 594 and (a2) 543, anti-phase spiking pattern for (a3) I = 49, and complex out-of-phase spiking patterns for I = (a4) 192 and (a5) 91. In (a1)-(a5), top panel: raster plots of spikes in the sub-population of pre-synaptic GO cells innervating the *I*th GR cluster, 2nd panel: plots of  $f_X^{(I)}(t)$ : bin-averaged instantaneous spike rates of the MF signals (X = MF) into the *I*th GR cluster (gray line) and bin-averaged instantaneous sub-population of pre-synaptic GO cells (X = GO) innervating the *I*th GR cluster (black line);  $\langle \cdots \rangle_r$  represents the realization average (number of realizations is 100), 3rd panel: time course of  $\langle g_{10}^{(I)}(t) \rangle_r$ : conductance of total synaptic inputs (including both the excitatory and inhibitory inputs) into the *I*th GR cluster, and bottom panel: plots of  $R_{GR}^{(I)}(t)$ : instantaneous cluster spike rate in the *I*th GR cluster. (b) Plot of cluster-averaged conductance  $\langle g_{tot}^{(w)}(t) \rangle_r$  of total synaptic inputs into the GR clusters versus *t*. (c) Distribution of conjunction indices  $\{C_{in}^{(I)}\}$  for the conductances of total synaptic inputs into the GR clusters.

and the out-of-phase spiking groups, spiking patterns of the GR clusters are the most diverse.

Figs. 6(c1)-6(c3) also show the plots of the fractions of conjunction indices  $C^{(I)}$  of the GR clusters in the in-phase, antiphase, and complex out-of-phase spiking groups, respectively. The ranges for the distributions  $\{C^{(l)}\}$  in the three spiking groups are also given in the bar diagram in Fig. 6(d). In the case of inphase spiking group, the distribution  $\{C^{(l)}\}$  with a peak at 0.55 has only positive values in the range of  $(C_h^*, C_{max})$   $(C_{max} = 0.85)$ and  $C_h^* \simeq 0.39$ ), and its mean and standard deviations are 0.538 and 0.181, respectively. On the other hand, in the case of the antiphase spiking group, the distribution  $\{C^{(l)}\}\$  with a peak at -0.25has only negative values in the range of  $(C_{min}, C_l^*)$   $(C_{min} = -0.57)$ and  $C_1^* \simeq -0.20$ ), and its mean and standard deviations are -0.331 and 0.135, respectively. Between the in-phase and the anti-phase spiking groups, there exists an intermediate complex out-of-phase spiking group. In this case, the range for the distribution  $\{\mathcal{C}^{(l)}\}\$  with a peak at 0.35 is  $(\mathcal{C}_l^*, \mathcal{C}_h^*)$ , and the mean and

the standard deviation are 0.174 and 0.242, respectively. As will be seen in the next subsection, these in-phase, anti-phase, and complex out-of-phase spiking groups play their own roles in the synaptic plasticity at PF-PC synapses, respectively.

Finally, we study the dynamical origin of diverse spiking patterns in the *I*th GR clusters. As examples, we consider two inphase spiking patterns for I = 594 and 543 [see the spiking patterns in Figs. 5(a1) and 5(a2)], one anti-phase spiking pattern for I = 49 [see the spiking pattern Fig. 5(b1)], and two complex out-of-phase spiking patterns for I = 192 and 91 [see the spiking patterns in Figs. 5(c1) and 5(c2)]. In Fig. 7, (a1)–(a5) correspond to the cases of I = 594, 543, 49, 192, and 91, respectively.

Diverse recodings for the MF signals are made in the GR layer, composed of excitatory GR and inhibitory GO cells (i.e., in the GR-GO cell feedback loop). In this case, spiking activities of GR cells are controlled by two types of synaptic input currents (i.e., excitatory synaptic inputs through MF signals and inhibitory synaptic inputs from randomly connected GO cells). Then, we make investigations on the dynamical origin of diverse spiking patterns of the GR clusters (shown in Fig. 5) through analysis of total synaptic inputs into the GR clusters. Synaptic current is given by the product of synaptic conductance g and potential difference [see Eq. (8)]. Here, synaptic conductance determines the time-course of synaptic current. Hence, it is enough to consider the time-course of synaptic of synaptic strength per synapse, the number of synapses  $M_{syn}$ , and the fraction s of open (post-synaptic) ion channels [see Eq. (9)]. Here, the synaptic strength per synapse is given by the product of maximum synaptic conductance  $\bar{g}$  and synaptic weight J, and the time-course of s(t) is given by a summation for exponential-decay functions over pre-synaptic spikes, as shown in Eqs. (9) and (10).

We make an approximation of the fraction s(t) of open ion channels (i.e., contributions of summed effects of pre-synaptic spikes) by the bin-averaged spike rate  $f_X^{(I)}(t)$  of pre-synaptic neurons (X = MF and GO);  $f_{MF}^{(I)}(t)$  is the bin-averaged spike rate of the MF signals into the *I*th GR cluster and  $f_{GO}^{(I)}(t)$  is the bin-averaged spike rate of the pre-synaptic GO cells innervating the *I*th GR cluster. Then, the conductance  $g_X^{(I)}(t)$  of synaptic input from X (= MF or GO) into the *I*th GR cluster ( $I = 1, ..., N_C$ ) is given by:

$$g_X^{(I)}(t) \simeq M_f^{(R)} \cdot f_X^{(I)}(t).$$
 (27)

Here, the multiplication factor  $M_f^{(R)}$  [= maximum synaptic conductance  $\bar{g}_R \times$  synaptic weight  $J^{(GR,X)} \times$  number of synapses  $M_{\text{syn}}^{(GR,X)}$ ] varies depending on X and the receptor R on the post-synaptic GR cells. In the case of excitatory synaptic currents into the *I*th GR cluster with AMPA receptors via the MF signal,  $M_f^{(AMPA)} = 2.88$ ;  $\bar{g}_{AMPA} = 0.18$ ,  $J^{(GR,MF)} = 8.0$ , and  $M_{\text{syn}}^{(GR)} = 2$ . In contrast, in the case of the *I*th GR cluster with NMDA receptors,  $\bar{g}_{\text{NMDA}} = 0.025$ , and hence  $M_f^{(\text{NMDA})} = 0.4$ , which is much less than  $M_f^{(AMPA)}$ . For the inhibitory synaptic current from presynaptic GO cells to the *I*th GR cluster with GABA receptors,  $M_f^{(GABA)} = 2.72$ ;  $\bar{g}_{GABA} = 0.028$ ,  $J^{(GR,GO)} = 10$ , and  $M_{\text{syn}}^{(GR,GO)} = 9.7$ . Then, the conductance  $g_{tot}^{(I)}$  of total synaptic inputs (including both the excitatory and the inhibitory input) into the *I*th GR cluster is given by:

$$g_{tot}^{(l)}(t) = g_{MF}^{(l)} - g_{GO}^{(l)} = g_{AMPA}^{(l)} + g_{NMDA}^{(l)} - g_{GO}^{(l)}$$
  
= 3.28 f\_{MF}^{(l)}(t) - 2.72 f\_{GO}^{(l)}(t). (28)

Total synaptic input with conductance  $g_{tot}^{(I)}(t)$  is fed into GR cells in the *I*th GR cluster, and then the corresponding output, given by the instantaneous cluster spike rate  $R_{GR}^{(I)}(t)$ , emerges. Through averaging  $g_{tot}^{(I)}(t)$  over all the GR clusters, we obtain the clusteraveraged conductance  $g_{tot}^{(w)}(t)$  of total synaptic inputs into the GR clusters:

$$g_{tot}^{(w)}(t) = \frac{1}{N_C} \sum_{l=1}^{N_C} g_{tot}^{(l)}(t).$$
(29)

The cluster-averaged total synaptic input with  $g_{tot}^{(w)}(t)$  gives rise to the cluster-averaged output, given by the instantaneous whole-population spike rate  $R_w(t) = \frac{1}{N_C} \sum_{l=1}^{N_C} R_{GR}^{(l)}(t)$ ].

In Figs. 7(a1)–7(a5), the top panels show the raster plots of spikes in the sub-populations of pre-synaptic GO cells innervating the *I*th GR clusters. We obtain bin-averaged (sub-population) spike rates  $f_{GO}^{(I)}(t)$  from the raster plots. The bin-averaged spike rate of pre-synaptic GO cells in the *i*th bin is given by  $\frac{n_i^{(s)}}{N_{pre} \Delta t}$ , where  $n_i^{(s)}$  is the number of spikes in the *i*th bin,  $\Delta t$  (= 10 ms) is the bin size, and  $N_{pre}$  (= 10) is the number of pre-synaptic

GO cells. Via an average over 100 realizations, we obtain the realization-averaged (bin-averaged) spike rate of pre-synaptic GO cells  $\langle f_{GO}^{(l)}(t) \rangle_r$  because  $N_{pre}$  (= 10) is small;  $\langle \cdots \rangle_r$  represent a realization-average. The 2nd panels show  $\langle f_{GO}^{(l)}(t) \rangle_r$  (black line) and  $\langle f_{MF}^{(l)}(t) \rangle_r$  (gray line). We note that  $\langle f_{GO}^{(l)}(t) \rangle$  changes depending on *I*, while  $\langle f_{MF}^{(l)}(t) \rangle$  is independent of *I*. In contrast to the spiking activity of GR cells [which exhibit random repetition of transitions between active (bursting) and inactive (silent) states (see Fig. 4(a))], GO cells exhibit relatively regular spikings, which may be well seen in slightly-modified sinusoidal-like bin-averaged spike rate  $\langle f_{GO}^{(l)}(t) \rangle_r$  (Heine, Highstein, & Blazquez, 2010; Yamazaki & Tanaka, 2007a). Then, we may get the realization-averaged conductance  $\langle g_{lot}^{(l)}(t) \rangle_r$  of total synaptic inputs in Eq. (28), which is shown in the 3rd panels. These conductances  $\langle g_{lot}^{(l)}(t) \rangle_r$  of total synaptic inputs relatively regular patterns.

We note that the shapes of  $\langle g_{tot}^{(I)}(t) \rangle_r$  (corresponding to the total input into the *I*th GR cluster) in the 3rd panels are nearly the same as those of  $R_{GR}^{(I)}(t)$  (corresponding to the output of the *I*th GR cluster) in the bottom panels. Hence, we expect that inphase (out-of-phase) inputs into the GR clusters may result in generation of in-phase (out-of-phase) outputs (i.e., responses) in the GR clusters. To confirm this point clearly, similar to case of the spiking patterns  $[R_{GR}^{(I)}(t)]$  (i.e., the outputs) in the GR clusters, we introduce the conjunction index for the total synaptic input into the *I*th GR cluster between  $\langle g_{tot}^{(I)}(t) \rangle_r$  (conductance of total synaptic inputs  $\langle g_{tot}^{(w)}(t) \rangle_r$ . Fig. 7(b) shows the plot of  $\langle g_{tot}^{(w)}(t) \rangle_r$  versus *t*. We also note that the shape of  $\langle g_{tot}^{(w)}(t) \rangle_r$  is similar to the instantaneous whole-population spike rate  $R_{GR}(t)$  in Fig. 4(b).

As in the case of the conjunction index  $C^{(l)}$  for the spiking patterns (i.e. outputs) in the *I*th GR cluster [see Eq. (25)], the conjunction index  $C_{in}^{(l)}$  for the total synaptic input is given by the cross-correlation at the zero-time lag (i.e.,  $Corr_{in}^{(l)}(0)$ ) between  $\langle g_{tot}^{(l)}(t) \rangle_r$  and  $\langle g_{tot}^{(w)}(t) \rangle_r$ :

$$Corr_{\rm in}^{(l)}(\tau) = \frac{\Delta \langle g_{tot}^{(w)}(t+\tau) \rangle_r \Delta \langle g_{tot}^{(l)}(t) \rangle_r}{\sqrt{\Delta \langle g_{tot}^{(w)}(t) \rangle_r^2} \sqrt{\Delta \langle g_{tot}^{(l)}(t) \rangle_r^2}},$$
(30)

where  $\Delta \langle \underline{g_{tot}^{(w)}(t)} \rangle_r = \langle g_{tot}^{(w)}(t) \rangle_r - \overline{\langle g_{tot}^{(w)}(t) \rangle_r}$ ,  $\Delta \langle g_{tot}^{(l)}(t) \rangle_r = \langle g_{tot}^{(l)}(t) \rangle_r - \langle g_{tot}^{(l)}(t) \rangle_r$ , and the overline represents the time average. Thus, we have two types of conjunction indices,  $C^{(l)}$  [output conjunction index: given by  $Corr_{GR}^{(l)}(0)$ ] and  $C_{in}^{(l)}$  [input conjunction index: given by  $Corr_{in}^{(l)}(0)$ ] for the output and the input in the *I*th GR cluster, respectively.

Fig. 7(c) shows the plot of fraction of input conjunction indices  $\{C_{in}^{(l)}\}$  in the whole GR clusters. We note that the distribution of input conjunction indices in Fig. 7(c) is nearly the same as that of output conjunction indices in Fig. 6(a).  $C_{in}^{(l)}$  increases slowly from the negative value to the peak at 0.55, and then it decreases rapidly. In this distribution of  $\{C_{in}^{(l)}\}$ , the range is (-0.57, 0.85), the mean is 0.321, and the standard deviation is 0.516. Then, we obtain the diversity degree  $\mathcal{D}_{in}$  for the total synaptic inputs  $\{\langle g_{lot}^{(l)}(t) \rangle_r\}$  of all the GR clusters:

 $\mathcal{D}_{in} = \text{Relative Standard Deviation for the Distribution of } \{\mathcal{C}_{in}^{(l)}\}.$ (31)

Hence,  $\mathcal{D}_{in} \simeq 1.607$  for the synaptic inputs, which is nearly the same as  $\mathcal{D}^* (\simeq 1.613)$  for the spiking patterns of GR cells. Consequently, diverse total synaptic inputs into the GR clusters lead to generation of diverse outputs (i.e., spiking patterns) of the GR cells.

# 3.3. Effect of diverse recoding in GR clusters on synaptic plasticity at PF-PC synapses

Based on dynamical classification of spiking patterns of GR clusters, we investigate the effect of diverse recoding in the GR clusters on synaptic plasticity at PF-PC synapses. As shown in the above subsection, MF context input signals for the post-eye-movement are diversely recoded in the granular layer (corresponding to the input layer of the cerebellar cortex). The diversely-recoded in-phase and out-of-phase PF (student) signals (corresponding to the outputs from the GR cells) are fed into the PCs (i.e., principal cells of the cerebellar cortex) and the BCs in the Purkinje-molecular layer (corresponding to the output layer of the cerebellar cortex). The PCs also receive in-phase error-teaching CF (instructor) signals from the IO, along with the inhibitory inputs from the BCs. Then, the synaptic weights at the PF-PC synapses vary depending on the relative phase difference between the PF (student) signals and the CF (instructor) signals.

We first consider the change in normalized synaptic weights  $\tilde{J}$  of active PF-PC synapses during learning in the optimal case of  $p_c^* = 0.06$ ;

$$\tilde{J}_{ij}(t) = \frac{J_{ij}^{(PC,PF)}(t)}{J_0^{(PC,PF)}}.$$
(32)

where the initial synaptic strength ( $J_0^{(PC,PF)} = 0.006$ ) is the same for all PF-PC synapses. Figs. 8(a1)–8(a5) show cycle-evolution of distribution of  $\tilde{J}(t)$  of active PF-PC synapses. With increasing the learning cycle, normalized synaptic weights  $\tilde{J}(t)$  are decreased due to LTD at PF-PC synapses, and eventually their distribution seems to be saturated at about the 300th cycle. We note that in-phase PF signals are strongly depressed (i.e., strong LTD) by the in-phase CF signals, while out-of-phase PF signals are weakly depressed (i.e., weak LTD) due to the phase difference between the PF and the CF signals. As shown in Fig. 4(c2), the activation degree  $A^{(G)}$  of the in-phase spiking group (G = i) is dominant at the middle stage of the cycle, while at the other parts of the cycle, the activation degrees of the in-phase (G = i) and the outof-phase (G = o) spiking groups are comparable. Consequently, strong LTD occurs at the middle stage, while at the initial and final stages somewhat less LTD takes place due to contribution of both the out-of-phase spiking group (with weak LTD) and the in-phase spiking group.

To more clearly examine the above cycle evolutions, we get the bin-averaged (normalized) synaptic weight in each *i*th bin (bin size:  $\Delta t = 100$  ms):

$$\langle \tilde{J}(t) \rangle_i = \frac{1}{N_{\text{s},i}} \sum_{f=1}^{N_{\text{s},i}} \tilde{J}_{i,f}(t),$$
(33)

where  $J_{i,f}$  is the normalized synaptic weight of the *f*th active PF signal in the *i*th bin, and  $N_{s,i}$  is the total number of active PF signals in the *i*th bin. Figs. 8(b1)–8(b5) show cycle-evolution of bin-averaged (normalized) synaptic weights  $\langle \tilde{J}(t) \rangle$  of active PF signals. In each cycle,  $\langle \tilde{J}(t) \rangle$  forms a well-shaped curve. With increasing the cycle, the well curve comes down, its modulation [= (maximum – minimum)/2] increases, and saturation seems to occur at about the 300th cycle.

We also get the cycle-averaged mean  $\langle \tilde{J}(t) \rangle$  via time average of  $\langle \tilde{J}(t) \rangle$  over a cycle:

$$\overline{\langle \tilde{J} \rangle} = \frac{1}{N_b} \sum_{i=1}^{N_b} \langle \tilde{J}(t) \rangle_i, \tag{34}$$

N.

where  $N_b$  is the number of bins for cycle averaging, and the overbar represents the time average over a cycle. Figs. 8(c) and

8(d) show plots of the cycle-averaged mean  $\langle \tilde{J}(t) \rangle$  and the modulation  $\mathcal{M}_J$  for  $\langle \tilde{J}(t) \rangle$  versus cycle. The cycle-averaged mean  $\langle \tilde{J}(t) \rangle$  decreases from 1 to 0.372 due to LTD at PF-PC synapses. However, strength of the LTD varies depending on the stages of the cycle. At the middle stage, strong LTD occurs, due to dominant contribution of in-phase active PF signals. On the other hand, at the initial and the final stage, somewhat less LTD takes place, because both the out-of-phase spiking group (with weak LTD) and the in-phase spiking group make contributions together. As a result, with increasing cycle, the middle-stage part comes down more rapidly than the initial and final parts, and hence the modulation  $\mathcal{M}_I$  increases from 0 to 0.112.

We now decompose the whole active PF signals into the in-phase and the out-of-phase active PF signals, and make an intensive investigation on their effect on synaptic plasticity at PF-PC synapses. Fig. 9 shows cycle-evolution of distributions of normalized synaptic weights  $\tilde{J}^{(G)}$  of active PF signals in the *G* spiking group [(a1)-(a5) in-phase (G = i) and (b1)-(b5) out-of-phase (G = o)] in the optimal case of  $p_c^* = 0.06$ ; the out-of-phase spiking groups. With increasing learning cycle, normalized synaptic weights  $\tilde{J}^{(G)}$  for the in-phase and the out-of-phase PF signals are decreased, and saturated at about the 300th cycle.

We note that the strength of LTD varies distinctly depending on the type of spiking group. The student PF signals (corresponding to the axons of the GR cells) are classified as in-phase or out-of-phase PF signals with respect to the "reference" signal  $R_{GR}(t)$ . Here,  $R_{GR}(t)$  is the instantaneous whole-population spike rate of Eq. (18)(denoting the population-averaged firing activity in the whole population of the GR cells). It is basically in proportion to the sinusoidal MF context signal  $f_{MF}(t)$  of Eq. (1), although its top part becomes lowered and flattened due to inhibitory coordination of GO cells. The PF student signals (coming from the GR clusters) are characterized in terms of their conjunction indices  $C^{(l)}$  of Eq. (25) between the instantaneous cluster spike rate  $R_{CR}^{(l)}(t)$  and the reference signal  $R_{GR}(t)$ . The ranges of  $\{C^{(l)}\}$  for the in-phase and the out-of-phase spiking groups are (0.39, 0.85) and (-0.57, 0.39), respectively, as shown in Fig. 6(d). The range for the out-of-phase spiking group is broader than that for the in-phase spiking group. These student PF signals are depressed by the instructor CF signals (coming from the IO neuron). We also note that the instructor CF signals are in-phase with the reference signal  $R_{GR}(t)$ , because the CF signals are also basically in proportion to the sinusoidal IO desired signal  $f_{DS}(t)$  of Eq. (2) which is in-phase with  $f_{MF}(t)$ . Hence, in-phase student PF signals are strongly depressed by the instructor CF signals, because they are "well-matched" with the in-phase CF signals. On the other hand, out-of-phase student PF signals are weakly depressed by the instructor CF signals, because they are "ill-matched" with the in-phase CF signals.

In the case of active in-phase PF signals, the distribution of their normalized synaptic weights  $\tilde{J}^{(G)}(t)$  (G = i) forms the bottom dense bands in Figs. 8(a1)-8(a5), due to strong LTD at the in-phase PF-PC synapses. The (vertical) widths of these bottom bands are narrow because of the narrow range of  $\{C^{(I)}\}$ . On the other hand, in the case of active out-of-phase PF signals, the distribution of  $\{J^{(G)}(t)\}$  (*G* = *o*) constitutes the upper sparse wellshaped parts in Figs. 8(a1)-8(a5), because of weak LTD at the out-of-phase PF-PC synapses. Due to the broad range of  $\{C^{(I)}\}$ , the heights of the well-shaped parts for the out-of-phase PF signals are higher than those of the bottom bands for the inphase PF signals with the narrow range. Moreover, the shapes for the distributions of  $\{\tilde{J}^{(G)}(t)\}\$  are consistent with the activation degrees  $A^{(G)}$  of Eq. (21) of the active PF signals in the G spiking group which is also shown in Fig. 4(c2). The activation degree  $A^{(o)}$ (G = o) for the out-of-phase spiking group has a small minimum



**Fig. 8.** Change in synaptic weights of active PF-PC synapses during learning in the optimal case of  $p_c^* = 0.06$ . (a1)–(a5) Cycle-evolution of distribution of normalized synaptic weights  $\tilde{J}$  of active PF signals. (b1)–(b5) Cycle-evolution of bin-averaged (normalized) synaptic weights  $\langle \tilde{J} \rangle$  of active PF signals. Bin size:  $\Delta t = 100$  ms. Plots of (c) cycle-averaged mean  $\langle \tilde{J} \rangle$  and (d) modulation  $\mathcal{M}_I$  for  $\langle \tilde{J} \rangle$  versus cycle.



**Fig. 9.** Change in synaptic weights of active PF-PC synapses in each spiking group during learning in the optimal case of  $p_c^* = 0.06$ . Cycle-evolution of distributions of normalized synaptic weights  $\tilde{J}^{(G)}$  of active PF signals in the *G* spiking group [G: (a1)–(a5) in-phase and (b1)–(b5) out-of-phase]. Cycle-evolution of distributions of normalized synaptic weights  $\tilde{J}^{(G)}$  of active out-of-phase PF signals in the *G* spiking group [G: (c1)–(c5) anti-phase and (d1)–(d5) complex out-of-phase].

in the middle stage of cycle, which leads to the well-shaped distributions of  $\{\tilde{J}^{(G)}(t)\}\ (G = o)$ . Hence, the in-phase PF signals (with strong LTD) make a large contribution in the middle stage of cycle, while at the initial and the final stage, contributions of both the out-of-phase PF signals (with weak LTD) and the in-phase PF signals are comparable.

In the above way, effective depression (i.e., strong/weak LTD) at PF-PC synapses occurs, depending on the spiking type (inphase or out-of-phase) of the active PF signals; strong (weak) LTD takes place for the in-phase (out-of-phase) PF signals. However, contributions of these in-phase and out-of-phase spiking groups vary depending on the stages of cycle. Fig. 4(c2) shows the activation degree  $A^{(G)}$  of the in-phase (G = i) and the out-of-phase (G = o) spiking groups. In the middle stage of cycle, the in-phase spiking group has a larger activation degree  $A^{(i)}$ . On the other hand, at the initial and the final stage, the activation degrees of the in-phase and the out-of-phase spiking groups are comparable. Hence, strong LTD takes place in the middle stage, due to a large contribution of the in-phase spiking group. Thus, the in-phase spiking group makes a big contribution to formation of the minimum of bin-averaged (normalized) synaptic weights  $\langle \tilde{J}(t) \rangle$  in Figs. 8(b1)–8(b5). In contrast, at the initial and the final stage of cycle, less LTD occurs because of comparable contributions of both the out-of-phase spiking group with weak LTD and the inphase spiking group with strong LTD. Hence, maxima of  $\langle \tilde{J}(t) \rangle$  appear at the initial and the final stage of cycle because both the (weakly-depressed) out-of-phase and the (strongly-depressed) in-phase spiking groups contribute together.

In this way, the in-phase and the out-of-phase spiking groups play their own roles in formation of modulation of  $\langle \tilde{J}(t) \rangle$ . The minimum of  $\langle \tilde{J}(t) \rangle$  in the middle stage of cycle is formed via a large contribution of the in-phase spiking group with strong LTD,



**Fig. 10.** Optimal case of  $p_c^* = 0.06$ . (a1)–(a5) Cycle-evolution of bin-averaged (normalized) synaptic weights  $\langle \tilde{J}^{(G)} \rangle$  of active PF signals in the *G* spiking-group. Plots of (b) cycle-averaged mean  $\overline{\tilde{J}^{(G)}}$  and (c) modulation  $\mathcal{M}_J^{(G)}$  for  $\tilde{J}^{(G)}$  versus cycle. (d1)–(d5) Cycle-evolution of firing fraction  $F^{(G)}$  of active PF signals in the *G* spiking group. (e1)–(e5) Cycle-evolution of weighted bin-averaged synaptic weights  $\langle W_J^{(G)} \rangle$  of active PF signals in the *G* spiking group, in the whole population of GR cells are also given. (f1) Plots of cycle-averaged means  $\overline{\langle W_J^{(G)} \rangle}$  and  $\overline{\tilde{J}}$  for  $\langle W_J^{(G)} \rangle$  and  $\langle \tilde{J} \rangle$  versus cycle. (f2) Plots of percentage contributions  $PC^{(G)}$  of the *G* spiking group (i.e.,  $\overline{\langle W_J^{(G)} \rangle / \tilde{J} \rangle$ ) versus cycle. Left, right, and middle vertical gray bands in each cycle in (e1)–(e5) denote the initial, final, and middle stages, respectively. Plots of stage-averaged values  $\overline{\langle W_J^{(G)} \rangle}$  and  $\overline{\tilde{J}}$  versus cycle at the (g1) middle, (g2) initial, and (g3) final stages of a cycle. Plots of percentage contributions  $PC^{(G)}$  of the *G* spiking group versus cycle at the (h1) middle, (h2) initial, and (h3) final stages of a cycle.

while formation of the maxima of  $\langle \tilde{J}(t) \rangle$  in the initial and the final stage is made via comparable contributions of the out-of-phase spiking group with weak LTD and the in-phase spiking group with strong LTD. Consequently, this kind of constructive interplay between the in-phase (strong LTD) and the out-of-phase (weak LTD) spiking groups leads to a big modulation of  $\langle \tilde{J}(t) \rangle$ , as shown in Fig. 8(d). (More detailed discussion on this point will be given below in Fig. 10.)

We also make further decomposition of the out-of-phase PF signals into the anti-phase and the complex out-of-phase ones. Fig. 9 also shows cycle-evolution of distributions of normalized synaptic weights  $\tilde{J}^{(G)}$  of active out-of-phase PF signals in the G spiking group [G: (c1)-(c5) anti-phase and (d1)-(d5) complex out-of-phase]. As the learning cycle is increased, normalized synaptic weights  $\tilde{J}^{(G)}$  for the anti-phase and the complex outof-phase PF signals are decreased and saturated at about the 300th cycle. In the case of anti-phase PF signals, weak depression occurs, and they constitute the top part for the out-of-phase PF signals in Figs. 9(b1)-9(b5). On the other hand, in the case of complex out-of-phase PF signals, intermediate LTD takes place, and they form the bottom part for the out-of-phase PF signals in Figs. 9(b1)-9(b5). These anti-phase (weak LTD) and complex outof-phase (intermediate LTD) spiking groups make contribution to formation of maxima of  $\langle \tilde{J}(t) \rangle$  at the initial and the final stage of cycle, together with the in-phase spiking group with strong LTD.

Figs. 10(a1)-10(a5) show cycle-evolutions of bin-averaged (normalized) synaptic weights  $\langle \tilde{J}^{(G)}(t) \rangle$  of active PF signals in the *G* spiking-group [i.e., corresponding to the bin-averages for the distributions of  $\tilde{J}^{(G)}(t)$  in Figs. 9(a1)-9(a5) and Figs. 9(b1)-9(b5)]; *G*: in-phase (solid circles) and out-of-phase (open circles). In the case of in-phase PF signals, they are strongly depressed without modulation. On the other hand, in the case of out-of-phase PF signals, they are more weakly depressed in comparison with the case at the middle stage.

The cycle-averaged means  $\langle \tilde{J}^{(G)}(t) \rangle$  and modulations  $\mathcal{M}_{J}^{(G)}$  for  $\langle \tilde{J}^{(G)}(t) \rangle$  are given in Figs. 10(b) and 10(c), respectively. Both cycleaveraged means in the cases of the in-phase and the out-of-phase PF signals decrease, and saturations occur at about the 300th cycle. In comparison with the case of out-of-phase PF signals (open circles), the cycle-averaged means in the case of in-phase PF signals (solid circles) are more reduced; the saturated limit value in the case of in-phase (out-of-phase) PF signals is 0.199 (0.529). In contrast, modulation occurs only for the out-of-phase PF signals (open circles), it increases with cycle, and becomes saturated at about the 300th cycle where the saturated value is 0.087.

In addition to the above bin-averaged (normalized) synaptic weights  $\langle \tilde{J}^{(G)}(t) \rangle$ , we need another information on firing fraction  $F^{(G)}(t)$  of active PF signals in the *G* (in-phase or out-of-phase) spiking group to obtain the contribution of each spiking group to the bin-averaged synaptic weights  $\langle \tilde{J}(t) \rangle$  of active PF signals in the whole population. The firing fraction  $F_i^{(G)}$  of active PF signals in the *G* spiking group in the *i*th bin is given by:

$$F_i^{(G)} = \frac{N_{s,i}^{(G)}}{N_{s,i}},\tag{35}$$

where  $N_{s,i}$  is the total number of active PF signals in the *i*th bin and  $N_{s,i}^{(G)}$  is the number of active PF signals in the *G* spiking group in the *i*th bin. We note that  $F^{(G)}(t)$  is the same, independently of the learning cycle, because firing activity of PF signals depends only on the GR and GO cells in the granular layer.

Figs. 10(d1)–10(d5) show the firing fraction  $F^{(G)}(t)$  of active PF signals in the *G* spiking group. The firing fraction  $F^{(G)}(t)$  for the inphase (*G* = *i*) active PF signals (solid circles) forms a bell-shaped curve, while  $F^{(G)}(t)$  for the out-of-phase (*G* = *o*) active PF signals (open circles) forms a well-shaped curve. The bell-shaped curve for the in-phase PF signal is higher than the well-shaped curve for the out-of-phase PF signal. For the in-phase PF signals, the firing fraction  $F^{(i)}(t)$  is about 0.94 (i.e., 94%) at the middle stage, and

about 0.51 (i.e., 51%) at the initial and the final stage. On the other hand, for the out-of-phase PF signals, the firing fraction  $F^{(0)}(t)$  is about 0.49 (i.e., 49%) at the initial and the final stage, and about 0.06 (i.e., 6%) at the middle stage. Consequently, the fraction of in-phase active PF signals is dominant at the middle stage, while at the initial and the final stage, the fractions of both in-phase and out-of-phase active PF signals are nearly the same.

The weighted bin-averaged synaptic weight  $\langle W_I^{(G)} \rangle_i$  for each G spiking group in the *i*th bin is given by the product of the firing fraction  $F_i^{(G)}$  and the bin-averaged (normalized) synaptic weight  $\langle \tilde{I}^{(G)} \rangle_i$ :

$$\langle W_j^{(G)} \rangle_i = F_i^{(G)} \, \langle \tilde{J}^{(G)} \rangle_i, \tag{36}$$

where the firing fraction  $F_i^{(G)}$  plays a role of a weighting function for  $\langle \tilde{J}^{(G)} \rangle_i$ . Then, the bin-averaged (normalized) synaptic weight  $\langle \tilde{J} \rangle_i$  of active PF signals in the whole population in the *i*th bin [see Eq. (33)] is given by the sum of weighted bin-averaged synaptic weights  $\langle W_{l}^{(G)} \rangle_{i}$  of all spiking groups:

$$\tilde{\langle J} \rangle_i = \sum_{G}^{\text{all spiking groups}} \langle W_J^{(G)} \rangle_i.$$
(37)

Hence,  $\langle W_I^{(G)}(t) \rangle$  represents contribution of the *G* spiking group to  $\langle \tilde{I}(t) \rangle$  of active PF signals in the whole population.

Figs. 10(e1)–10(e5) show cycle-evolution of weighted bin-averaged synaptic weights  $\langle W_I^{(G)}(t) \rangle$  of active PF signals in the *G* spiking group [G: in-phase (solid circles) and out-of-phase (open circles)]. In the case of in-phase PF signals, the bin-averaged (normalized) synaptic weights  $\langle \tilde{J}^{(G)}(t) \rangle$  are straight horizontal lines, the firing fraction  $F^{(G)}(t)$  is a bell-shaped curve, and hence their product leads to a bell-shaped curve for the weighted bin-averaged synaptic weight  $\langle W_J^{(G)}(t) \rangle$ . With increasing the cycle, the horizontal straight lines for  $\langle \tilde{J}^{(G)}(t) \rangle$  come down rapidly, while there is no change with cycle in  $F^{(G)}(t)$ . Hence, the bell-shaped curves for  $\langle W_l^{(G)}(t) \rangle$  also come down quickly, their modulations also are reduced in a fast way, and they become saturated at about the 300th cycle.

On the other hand, in the case of out-of-phase PF signals, the bin-averaged (normalized) synaptic weights  $\langle \tilde{I}^{(G)}(t) \rangle$  lie on a well-shaped curve, the firing fraction  $F^{(G)}(t)$  also is a well-shaped curve, and then their product results in a well-shaped curve for the weighted bin-averaged synaptic weight  $\langle W_l^{(G)}(t) \rangle$ . With increasing the cycle, the well-shaped curves for  $\langle \tilde{J}^{(G)}(t) \rangle_i$  come down slowly, while there is no change with cycle in  $F^{(G)}(t)$ . Hence, the well-shaped curves for  $\langle W_{l}^{(G)}(t) \rangle$  also come down gradually, their modulations also are reduced little by little, and eventually they get saturated at about the 300th cycle.

For comparison, bin-averaged (normalized) synaptic weights  $\langle I(t) \rangle$  of active PF signals in the whole population (crosses) are also given in Figs. 10(e1)-10(e5), and they form a well-shaped curve. According to Eq. (37), the sum of the values at the solid circle (in-phase) and the open circle (out-of-phase) at a time t in each cycle is just the value at the cross (whole population). At the middle stage of each cycle, contributions of in-phase PF signals (solid circles) are dominant [i.e., contributions of out-of-phase PF signals (open circles) are negligible], while at the initial and the final stage, contributions of out-of-phase PF signals are larger than those of in-phase PF signals (both contributions must be considered). Consequently,  $\langle \tilde{J}(t) \rangle$  of active PF signals in the whole population becomes more reduced at the middle stage than at the initial and the final stage, due to the dominant effect (i.e., strong LTD) of in-phase active PF signals at the middle stage, which results in a well-shaped curve for  $\langle I(t) \rangle$  in the whole population.

We make a quantitative analysis for contribution of  $\langle W_I^{(G)}(t) \rangle$ in each *G* spiking group to  $\langle \tilde{J}(t) \rangle$  in the whole population. Fig. 10(f1) shows plots of cycle-averaged weighted synaptic weight  $\overline{\langle W_J^{(G)}(t) \rangle}$  (i.e., time average of  $\langle W_J^{(G)}(t) \rangle$  over a cycle) in the *G* spiking-group [*G*: in-phase (solid circles) and out-ofphase (open circles)] and cycle-averaged synaptic weight  $\langle \tilde{J}(t) \rangle$  of Eq. (34) in the whole population (crosses) versus cycle. The cycleaveraged weighted synaptic weights  $\overline{\langle W_J^{(G)}(t) \rangle}$  in the in-phase spiking group (solid circles) are larger than those in the out-ofphase spiking group (open circles), and their sums correspond to the cycle-averaged synaptic weight  $\langle \tilde{J}(t) \rangle$  in the whole population (crosses). With increasing cycle, both  $\langle W_I^{(G)}(t) \rangle$  and  $\langle \tilde{J}(t) \rangle$  become saturated at about the 300th cycle. In the in-phase spiking group  $\overline{\langle W_J^{(G)}(t) \rangle}$  decreases rapidly from 0.722 to 0.198, while  $\overline{\langle W_J^{(G)}(t) \rangle}$  in the out-of-phase spiking group decreases slowly from 0.273 to 0.174. Thus, the saturated values of  $\langle W_j^{(G)}(t) \rangle$  in both the in-phase and the out-of-phase spiking groups become close. The percentage contribution  $PC^{(G)}$  of  $\langle W_j^{(G)}(t) \rangle$  in the *G* spiking

group to  $\langle \tilde{I}(t) \rangle$  in the whole population is given by:

$$PC^{(G)}(\%) = \frac{\overline{\langle W_J^{(G)}(t) \rangle}}{\overline{\langle \tilde{J}(t) \rangle}} \times 100.$$
(38)

Fig. 10(f2) shows a plot of  $PC^{(G)}$  versus cycle [G: in-phase (solid circles) and out-of-phase (open circles)].  $PC^{(G)}$  of the in-phase spiking group decreases from 72.2% to 53.2%, while  $PC^{(G)}$  of the out-of-phase spiking group increases from 27.3% to 46.8%. Thus, the saturated values of  $PC^{(G)}$  of both the in-phase and the out-ofphase spiking groups get close.

We are particularly interested in the left, the right, and the middle vertical gray bands in each cycle in Figs. 10(e1)-10(e5) which denote the initial (0 < t < 100 ms), the final (1900 < t <2000 ms), and the middle (900 < t < 1100 ms) stages, respectively. In the case of in-phase (out-of-phase) spiking group, the maximum (minimum) of  $\langle W_{I}^{(G)}(t) \rangle$  appears at the middle stage, while the minimum (maximum) occurs at the initial and the final stage. Figs. 10(g1)-10(g3) show plots of stage-averaged weighted synaptic weight  $\overline{\langle W_{l}^{(G)}(t) \rangle}$  [i.e., time average of  $\langle W_{l}^{(G)}(t) \rangle$  over a stage] in the G spiking-group [G: in-phase (solid circles) and out-of-phase (open circles)] and stage-averaged synaptic weight  $\langle I(t) \rangle$  [i.e., time average of  $\langle I(t) \rangle$  over the stage] in the whole population (crosses) versus cycle in the middle, the initial, and the final stages, respectively. The sum of the values of  $\langle W_I^{(G)}(t) \rangle$ at a time t in the in-phase and the out-of-phase spiking groups corresponds to the value of  $\langle \tilde{J}(t) \rangle$  in the whole population. As the cycle is increased, both  $\overline{\langle W_J^{(G)}(t) \rangle}$  and  $\overline{\langle \tilde{J}(t) \rangle}$  become saturated at about the 300th cycle. Figs. 10(h1)–10(h3) also show plots of percentage contribution  $PC^{(G)}$  of the *G* spiking group (i.e., ratio of the stage-averaged weighted synaptic weight  $\overline{\langle W_{I}^{(G)}(t) \rangle}$  in the G spiking group to the stage-averaged synaptic weight  $\langle \tilde{I}(t) \rangle$  in the whole population) in the middle, the initial, and the final stages, respectively [G: in-phase (solid circles) and out-of-phase (open circles)].

In the case of in-phase spiking group,  $\langle W_{I}^{(G)}(t) \rangle$  decreases rapidly with cycle in all the 3 stages, while in the case of outof-phase spiking group, it also decreases in a relatively slow way with cycle. At the middle stage,  $\overline{\langle W_J^{(G)}(t) \rangle}$  in the in-phase spiking group (solid circles) is much higher than that in the out-of-phase spiking group (open circles), and it decreases rapidly from 0.944 to 0.266. In this case, the percentage contribution  $PC^{(G)}$  of the inphase spiking group increases from 94.4% to 97.0%. Consequently,

the contribution of in-phase spiking group is dominant at the middle stage, which leads to strong LTD at the PF-PC synapses. On the other hand, at the initial and the final stage, with increasing cycle  $\langle W_j^{(G)}(t) \rangle$  in the out-of-phase spiking group becomes larger than that in the in-phase spiking group. The percentage contribution  $PC^{(G)}$  of the out-of-phase spiking group increases from 49.2% to 70.2%, while that of the in-phase spiking group decreases from 50.8% to 29.8%. As a result, the contribution of out-of-phase spiking group at the initial and the final stage is larger than that of in-phase spiking group, which results in weak LTD at the PF-PC synapses.

In the above way, under good balance between the in-phase and the out-of-phase spiking groups (i.e., spiking-group ratio  $\mathcal{R}^* \simeq 1.008$ ) in the optimal case of  $p_c^* = 0.06$ , effective depression (i.e., strong/weak LTD) at the PF-PC synapses causes a big modulation in synaptic plasticity, which also leads to large modulations in firing activities of the PCs and the VN neuron (i.e., emergence of effective learning process). Hence, diverse recoding in the granular layer (i.e., appearance of diverse spiking patterns in the GR clusters) which results in effective (strong/weak) LTD at the PF-PC synapses is essential for effective motor learning for the OKR adaptation, which will be discussed in the following subsection.

### 3.4. Effect of PF-PC synaptic plasticity on subsequent learning process in the PC-VN-IO system

As a result of diverse recoding in the GR clusters, strong LTD occurs at the middle stage of a cycle due to dominant contribution of the in-phase spiking group. On the other hand, at the initial and the final stage, somewhat less LTD takes place due to contribution of both the out-of-phase spiking group (with weak LTD) and the in-phase spiking group. Due to this kind of effective (strong/weak) LTD at the PF-PC synapses, a big modulation in synaptic plasticity at the PF-PC synapses (i.e., big modulation in bin-averaged normalized synaptic weight  $\langle \tilde{J}(t) \rangle$ ) emerges. In this subsection, we investigate the effect of PF-PC synaptic plasticity with a big modulation on the subsequent learning process in the PC-VN-IO system.

Fig. 11 shows change in firing activity of PCs during learning in the optimal case of  $p_c^* = 0.06$ . Cycle-evolutions of raster plots of spikes of 16 PCs and the corresponding instantaneous population spike rates  $R_{PC}(t)$  are shown in Figs. 11(a1)–11(a5) and Figs. 11(b1)–11(b5), respectively. Since the number of PCs is small,  $R_{PC}(t)$  seems to be a little rough. To get a smooth estimate for  $R_{PC}(t)$ , we make 100 realizations. Realization-averaged smooth instantaneous population spike rates  $\langle R_{PC}(t) \rangle_r$  are given in Figs. 11(c1)–11(c5);  $\langle \cdots \rangle_r$  denotes realization average and the number of realizations is 100.  $\langle R_{PC}(t) \rangle_r$  seems to be saturated at about the 300th cycle.

With increasing the learning cycle, raster plots of spikes of all the 16 PCs become more and more sparse at the middle stage, which may be clearly seen in the instantaneous population spike rate  $\langle R_{PC}(t) \rangle_r$ . Due to the effect of synaptic plasticity at the PF-PC synapses, the minimum of  $\langle R_{PC}(t) \rangle_r$  appears at the middle stage, while the maximum occurs at the initial and the final stage. Thus, the modulation of  $\langle R_{PC}(t) \rangle_r$  increases with increasing the cycle.

In-phase PF (student) signals are strongly depressed by the inphase CF (instructor) signals, while out-of-phase PF signals are weakly pressed due to the phase difference between the out-ofphase PF signals and the in-phase CF signals. Fraction of in-phase PF signals at the middle stage of a cycle is dominant. On the other hand, at the initial and the final stage, fraction of out-of-phase PF signals are larger than that of in-phase PF signals. Thus, binaveraged normalized synaptic weights  $\langle \tilde{J}(t) \rangle$  form a well-shaped curve, as shown in Figs. 8(b1)-8(b5). That is, strong LTD occurs at the middle stage, while at the initial and the final stages, weak LTD takes place. As a result of this kind of effective depression (strong/weak LTD) at the (excitatory) PF-PC synapses,  $\langle R_{PC}(t) \rangle_r$  becomes lower at the middle stage (strong LTD) than at the initial and the final stage (weak LTD). Thus,  $\langle R_{PC}(t) \rangle_r$  forms a well-shaped curve with a minimum at the middle stage.

Figs. 11(d1) and 11(d2) show plots of cycle-averaged mean  $\overline{\langle R_{PC}(t) \rangle_r}$  (i.e., time average of  $\langle R_{PC}(t) \rangle_r$  over a cycle) and modulation  $\mathcal{M}_{PC}$  of  $\langle R_{PC}(t) \rangle_r$  versus cycle, respectively. Due to LTD at the PF-PC synapses, the cycle-averaged mean  $\overline{\langle R_{PC}(t) \rangle_r}$  decreases monotonically from 86.1 Hz, and it becomes saturated at 51.7 Hz at about the 300th cycle. On the other hand, the modulation  $\mathcal{M}_{PC}$  increases monotonically from 2.6 Hz, and it get saturated at 24.1 Hz at about the 300th cycle. Consequently, a big modulation occurs in  $\mathcal{M}_{PC}$  due to the effective depression (strong/weak LTD) at the PF-PC synapses. These PCs (principal cells of the cerebellar cortex) exert effective inhibitory coordination on the VN neuron which evokes OKR eye-movement.

Fig. 12 shows change in firing activity of the VN neuron which produces the final output of the cerebellum during learning in the optimal case of  $p_c^* = 0.06$ . Cycle-evolutions of raster plots of spikes of the VN neuron (i.e., collection of spike trains for all the realizations; number of realizations is 100) and the bin-averaged instantaneous individual firing rates  $f_{\rm VN}(t)$  [i.e., the number of spikes of the VN neuron in a bin divided by the bin width ( $\Delta t = 100 \text{ ms}$ )] are shown in Figs. 12(a1)–12(a5) and Figs. 12(b1)–12(b5), respectively.  $f_{\rm VN}(t)$  seems to be saturated at about the 300th cycle.

In contrast to the case of PCs, as the cycle is increased, raster plots of spikes of the VN neuron become more and more dense at the middle stage, which may be clearly seen in the instantaneous individual firing rates  $f_{VN}(t)$ . Due to the effective inhibitory coordinations of PCs on the VN neuron, the maximum of  $f_{VN}(t)$ appears at the middle stage, while the minimum occurs at the initial and the final stage. Thus,  $f_{VN}(t)$  forms a bell-shaped curve.

Figs. 12(c1) and 12(c2) show plots of cycle-averaged mean  $f_{VN}(t)$  of  $f_{VN}(t)$  [i.e., time average of  $f_{VN}(t)$  over a cycle] and modulation  $\mathcal{M}_{VN}$  of  $f_{VN}(t)$  versus cycle, respectively. Due to the decreased inhibitory inputs from the PCs, the cycle-averaged mean  $f_{VN}(t)$  increases monotonically from 44.3 Hz, and it becomes saturated at 71.5 Hz at about the 300th cycle. Also, the modulation of  $f_{VN}(t)$  increases from 20.2 Hz, and it gets saturated at 32.5 Hz at about the 300th cycle. As a result of effective inhibitory coordination of PCs, a big modulation occurs in  $\mathcal{M}_{VN}$ .

The learning gain degree  $\mathcal{L}_g$ , corresponding to the modulation gain ratio, is given by the normalized modulation of  $f_{VN}(t)$  divided by that at the 1st cycle:

$$\mathcal{L}_g = \frac{\mathcal{M}_{VN}}{\mathcal{M}_{VN} \text{ at the 1st cycle}},\tag{39}$$

where  $\mathcal{M}_{\rm VN}$  at the 1st cycle is 20.2 Hz. Fig. 12(d) shows a plot of  $\mathcal{L}_g$  versus cycle.  $\mathcal{L}_g$  increases monotonically from 1, and it becomes saturated at about the 300th cycle. Thus, we get the saturated learning gain degree  $\mathcal{L}_g^*$  ( $\simeq$  1.608). As will be seen in the next subsection,  $\mathcal{L}_g^*$  ( $\simeq$  1.608) is the largest one among the available ones. Hence, in the optimal case of  $p_c^* = 0.06$ where spiking patterns of GR clusters with the diversity degree  $\mathcal{D}^*$  ( $\simeq$  1.613) are the most diverse, motor learning for the OKR adaptation with the saturated learning gain degree  $\mathcal{L}_g^*$  ( $\simeq$  1.608) is the most effective.

Learning progress may be seen clearly in the IO system. During the learning cycle, the IO neuron receives both the excitatory desired signal for a desired eye-movement and the inhibitory signal from the VN neuron (denoting a realized eye-movement). We introduce the learning progress degree  $\mathcal{L}_p$ , given by the ratio of the cycle-averaged inhibitory input from the VN neuron to the



**Fig. 11.** Change in firing activity of PCs during learning in the optimal case of  $p_c^* = 0.06$ . (a1)-(a5) Raster plots of spikes of PCs and (b1)-(b5) instantaneous population spike rates  $R_{PC}(t)$ . (c1)-(c5) Realization-averaged instantaneous population spike rates  $\langle R_{PC}(t) \rangle_r$ ; number of realizations is 100. Plots of (d1) cycle-averaged mean  $\overline{\langle R_{PC}(t) \rangle_r}$  and (d2) modulations  $\mathcal{M}_{PC}$  for  $\langle R_{PC}(t) \rangle_r$  versus cycle.



**Fig. 12.** Change in firing activity of the VN neuron during learning in the optimal case of  $p_c^* = 0.06$ . (a1)-(a5) Raster plots of spikes of the VN neuron (i.e., collection of spike trains for all the realizations; number of realizations is 100) and (b1)-(b5) bin-averaged instantaneous individual firing rate  $f_{VN}(t)$ ; the bin size is  $\Delta t = 100$  ms. Plots of (c1) cycle-averaged mean  $\overline{f_{VN}(t)}$  and (c2) modulation  $M_{VN}$  for  $f_{VN}(t)$  versus cycle. (d) Plot of learning gain degree  $\mathcal{L}_g$  versus cycle.



**Fig. 13.** Change in firing activity of the IO neuron during learning in the optimal case of  $p_c^* = 0.06$ . Plots of (a1) realization-average for the cycle-averaged inhibitory synaptic current from the VN neuron  $(\langle \overline{l}_{GABA}^{(10,VN)} \rangle_r)$  (open circles) and realization-average for the magnitude of the cycle-averaged excitatory synaptic current through the IO desired signal  $(\langle \overline{l}_{AMPA}^{(10,VN)} \rangle_r)$  (crosses) versus cycle; number of realizations  $\langle \cdots \rangle_r$  is 100. (a2) Plot of learning progress degree  $\mathcal{L}_p$  versus cycle. (b1)-(b5) Raster plots of spikes of the lone unor (i.e., collection of spike trains for all the realizations; number of realizations is 100) and (c1)-(c5) bin-averaged instantaneous individual firing rate  $f_{10}(t)$ ; the bin size is  $\Delta t = 400$  ms. (d) Plot of cycle-averaged individual firing rate  $f_{10}(t)$  versus cycle.

magnitude of the cycle-averaged excitatory input via the desired signal:

$$\mathcal{L}_p = \frac{\overline{I_{\mathsf{GABA}}^{(\mathsf{IO},\mathsf{VN})}}}{|\overline{I_{\mathsf{AMPA}}^{(\mathsf{IO},\mathsf{DS})}}|},\tag{40}$$

where  $\overline{I_{GABA}^{(IO,VN)}}$  is the cycle-averaged inhibitory GABA receptormediated current from the VN neuron into the IO neuron, and  $\overline{I_{AMPA}^{(IO,DS)}}$  is the cycle-averaged excitatory AMPA receptor-mediated current into the IO neuron via the desired signal; no (excitatory) NMDA receptors exist on the IO neuron. [Note that the 4th term in Eq. (3) is given by  $-I_{Syn,i}^{(X)}(t)$ , because  $I_{GABA}^{(IO,CN)} > 0$  and  $I_{AMPA}^{(IO,DS)} < 0$ .] Fig. 13(a1) shows plots of  $\overline{I_{GABA}^{(IO,VN)}}$  (open circles) and  $|\overline{I_{AMPA}^{(IO,DS)}}|$  (crosses) versus cycle in the optimal case of  $p_c^* = 0.06$ . With increasing the cycle, the cycle-averaged inhibitory input from the VN neuron increases, and converges to the constant magnitude of the cycle-averaged excitatory input through the IO desired signal. Thus, as shown in Fig. 13(a2),  $\mathcal{L}_p$  increases with cycle, and at about the 300th cycle, it becomes saturated at  $\mathcal{L}_p = 1$ . In this saturated case, the cycle-averaged excitatory and inhibitory inputs into the IO neuron are balanced.

We also study the firing activity of IO neuron during learning process. Figs. 13(b1)–13(b5) and Figs. 13(c1)–13(c5) show cycle-evolutions of raster plots of spikes of the IO neuron (i.e., collection

of spike trains for all the realizations; number of realizations is 100) and the bin-averaged instantaneous individual firing rates  $f_{IO}$ [i.e., the number of spikes of the IO neuron in a bin divided by the bin width ( $\Delta t = 400 \text{ ms}$ )], respectively. In the 1st cycle, relatively dense spikes appear at the middle stage of the cycle in the raster plot of spikes, due to the effect of excitatory IO desired signal. However, with increasing the cycle, spikes at the middle stage become sparse, because of increased inhibitory input from the VN neuron. In this case, the bin-averaged instantaneous individual firing rate  $f_{IO}(t)$  of the IO neuron forms a bell-shaped curve due to the sinusoidally-modulating desired input signal into the IO neuron. With increasing the cycle, the amplitude of  $f_{IO}(t)$  decreases due to the inhibitory input from the VN neuron, and it becomes saturated at about the 300th cycle. Thus, the cycleaveraged individual firing rate  $\overline{f_{IO}(t)}$  is decreased from 1.51 Hz to 0.09 Hz, as shown in Fig. 13(d). The firing output of the IO neuron is fed into the PCs via the CFs. Hence, with increasing the cycle, the error-teaching CF instructor signals become weaker and saturated at about the 300th cycle.

While the saturated CF signals are fed into the PCs, saturation for the cycle-averaged bin-averaged synaptic weights  $\langle \tilde{J}(t) \rangle$  appears [see Fig. 8(c)]. Then, the subsequent learning process in the PC-VN system also becomes saturated, and we get the saturated learning gain degree  $\mathcal{L}_g^* \simeq 1.608$ ), which is shown in Fig. 12(d).

![](_page_19_Figure_1.jpeg)

**Fig. 14.** Highly-connected case of  $p_c = 0.6$ . (a) Raster plot of spikes of  $10^3$  randomly chosen GR cells. (b) Instantaneous whole-population spike rate  $R_{GR}(t)$  in the whole population of GR cells. Band width for  $R_{GR}(t)$ : h = 10 ms. (c) Distribution of conjunction indices  $\{C^{(l)}\}$  for the GR clusters in the whole population. (d) Fraction of spiking groups. Distributions of conjunction indices  $\{C^{(l)}\}$  for the (e1) in-phase and (e2) out-of-phase spiking groups. Bin size for the histograms in (c) and (e1)–(e2) is 0.1. (f) Ranges of  $\{C^{(l)}\}$  in the in-phase and the out-of-phase spiking groups.

## 3.5. Dependence of diversity degree D and learning gain degree $\mathcal{L}_g$ on $p_c$ (connection probability from GO to GR cells)

In the above subsections, we consider only the optimal case of  $p_c^* = 0.06$  (i.e., 6%) where the spiking patterns of the GR clusters are the most diverse. From now on, we vary the connection probability  $p_c$  from GO to GR cells, and investigate the dependence of the diversity degree  $\mathcal{D}$  for the spiking patterns of the GR clusters and the learning gain degree  $\mathcal{L}_g$  on  $p_c$ .

We first consider the highly-connected case of  $p_c = 0.6$ (i.e., 60%). Fig. 14(a) shows the raster plot of spikes of  $10^3$  randomly chosen GR cells, and the population-averaged firing activity in the whole population of GR cells may be well seen in the instantaneous whole-population spike rate  $R_{GR}(t)$  in Fig. 14(b). As shown in Fig. 2(b), each GR cluster is bounded by two glomeruli (corresponding to the terminals of the MFs) at both ends. Each glomerulus receives inhibitory inputs from nearby 81 GO cells with the connection probability  $p_c = 0.6$ . Hence, on average, about 49 GO cell axons innervate each glomerulus. Then, each GR cell in a GR cluster receives about 97 inhibitory inputs via two dendrites which contact the two glomeruli at both ends. Due to the increased inhibitory inputs from the pre-synaptic GO cells, spike density in the raster plot is decreased, and the top part of  $R_{GR}(t)$  becomes lowered and broadly flattened, in comparison with the optimal case in Fig. 4(b). Thus,  $R_{GR}(t)$  becomes more different from the firing rate  $f_{MF}$  for the MF signal in Fig. 1(b1).

GR cells in each GR cluster shares the same inhibitory and the excitatory inputs through their dendrites which synaptically contact the two glomeruli at both ends. Thus, GR cells in each GR cluster exhibit similar firing activity. Then, similar to the case of  $R_{GR}(t)$ , the cluster-averaged firing activity in the *I*th GR cluster ( $I = 1, ..., 2^{10}$ ) may be well described in terms of its instantaneous cluster spike rate  $R_{GR}^{(I)}(t)$  of Eq. (24). In this case, the conjunction index  $C^{(l)}$  of the *I*th GR cluster (representing the similarity degree between the spiking behavior  $[R_{GR}^{(l)}(t)]$  of the *I*th GR cluster and that of the whole population  $[R_{GR}(t)]$ ) is given by the cross-correlation at the zero time lag between  $R_{GR}^{(l)}(t)$  and  $R_{GR}(t)$  [see Eq. (25)].

Fig. 14(c) shows the distribution of conjunction indices  $\{C^{(I)}\}$ with a peak at 0.65. When compared with the optimal case in Fig. 6(a) with a peak at 0.55, the whole distribution is moved to the right, and the values of  $C^{(I)}$  for all the GR clusters are positive. Thus, all the anti-phase and complex out-of-phase spiking patterns with negative values of  $C^{(I)}$  disappear. Only the in-phase and out-of-phase spiking patterns with positive values of  $C^{(l)}$  persist. Consequently, the mean of the distribution  $\{C^{(I)}\}$  is increased to 0.613, while the standard deviation is decreased to 0.125, in comparison to the optimal case where the mean and the standard deviation are 0.320 and 0.516, respectively. Then, the diversity degree  $\mathcal{D}$  of the spiking patterns  $\{R_{GR}^{(l)}(t)\}$  in all the GR clusters, representing a quantitative measure for diverse recoding in the granular layer, is given by the relative standard deviation for the distribution  $\{C^{(l)}\}$  [see Eq. (26)]. In the highly-connected case of  $p_c = 0.6$ , its diversity degree is  $\mathcal{D} \simeq 0.204$  which is much smaller than  $\mathcal{D}^*$  ( $\simeq 1.613$ ) in the optimal case.

The reduction in  $\mathcal{D}$  for the spiking patterns (corresponding to the firing outputs) of the GR clusters arises due to decrease in differences between the total synaptic inputs into each GR cluster. As the connection probability  $p_c$  from the GO to GR cells is increased, differences between the total inhibitory synaptic inputs from the pre-synaptic GO cells into each GR cluster are decreased due to increase in the number of pre-synaptic GO cells. On the other hand, the excitatory inputs into each GR cluster via MFs are Poisson spike trains with the same firing rates, and hence they are essentially the same. Thus, differences between the total synaptic inputs (including both the inhibitory and the excitatory inputs) into each GR cluster become reduced. These less different inputs into the GR clusters produce less different outputs (i.e. spiking patterns) in the GR clusters, which leads to decreases in the diversity degree  $\mathcal{D}$  in the highly-connected case.

We decompose the whole GR clusters into the in-phase and the out-of-phase spiking groups. Unlike the optimal case of  $p_c^* =$ 0.06, no anti-phase spiking group appears. Fig. 14(d) shows the fraction of spiking groups. The in-phase spiking group is a major one with fraction 81.5%, while the out-of-phase spiking group is a minor one with fraction 18.5%. In comparison with the optimal case where the fraction of in-phase spiking group is 50.2%, the fraction of in-phase spiking group for  $p_c = 0.6$  is so much increased. In this highly-connected case, the spiking-group ratio, given by the ratio of the fraction of the in-phase spiking group to that of the out-of-phase spiking group, is  $\mathcal{R} \simeq 4.405$  which is much larger than that ( $\mathcal{R}^* \simeq 1.008$ ) in the optimal case. Thus, good balance between the in-phase and the out-of-phase spiking groups in the optimal case becomes broken up because the in-phase spiking group is a dominant one. In this unbalanced case, the diversity degree  $\mathcal{D}$  for the spiking patterns in the GR clusters is decreased so much to  $\mathcal{D}\simeq 0.204$ , in comparison with  $\mathcal{D}^*$  ( $\simeq$  1.613) in the optimal case of  $p_c^* = 0.06$ .

Figs. 14(e1) and 14(e2) also show plots of the fractions of conjunction indices  $C^{(l)}$  of the GR clusters in the in-phase and the out-of-phase spiking groups, respectively. The ranges for the distributions  $\{C^{(l)}\}$  in the two spiking groups are also given in the bar diagram in Fig. 14(f). As in the optimal case of  $p_c^* = 0.06$  in Figs. 5 and 6, we determine the threshold  $C_{th}$  ( $\simeq 0.41$ ) between the in-phase and the out-of-phase spiking patterns by making intensive examination of phase difference of  $R_{CR}^{(l)}(t)$  of the GR clusters relative to  $R_{CR}(t)$ . For  $C^{(l)} > C_{th}$ , in-phase spiking patterns with one or more peaks in the middle part of cycle exist. On the other hand, when passing the threshold  $C_{th}$  from the above

![](_page_20_Figure_2.jpeg)

**Fig. 15.** Change in synaptic weights of active PF-PC synapses and firing activity of PCs during learning in the highly-connected case of  $p_c = 0.6$ ; for comparison, data in the optimal case of  $p_c^* = 0.06$  are also given. (a1)–(a5) Cycle-evolution of bin-averaged (normalized) synaptic weights  $\overline{J}(t)$  of active PF signals (bin size:  $\Delta t = 100$  ms). Plots of (b1) cycle-averaged mean  $\overline{J}(t)$  and (b2) modulation  $\mathcal{M}_J$  for  $\langle \tilde{J}(t) \rangle$  versus cycle. (c1)–(c5) Realization-averaged instantaneous population spike rate  $\langle R_{PC}(t) \rangle_r$ ; the number of realizations is 100. Plots of (d1) cycle-averaged mean  $\overline{\langle R_{PC}(t) \rangle_r}$  and (d2) modulations  $\mathcal{M}_{PC}$  for  $\langle R_{PC}(t) \rangle_r$  versus cycle. In (a1)–(a5), (b1)–(b2), and (d1)–(d2), solid (open) circles represent data in the case of  $p_c = 0.6$  (0.06). In (c1)–(c5), solid (dotted) lines denote data in the case of  $p_c = 0.6$  (0.06).

(i.e., for  $C^{(l)} < C_{th}$ ) out-phase spiking patterns appear. These outof-phase spiking patterns have left-skewed (right-skewed) peaks near the 1st (3rd) quartile of cycle (i.e., near t = 500 (1500) ms).

In the case of in-phase spiking group, the distribution { $C^{(l)}$ } with a peak at 0.65 has positive values in the range of (0.41, 0.92). When compared with the optimal case where the range is (0.39, 0.85), the range in the highly-connected case of  $p_c = 0.6$  is shifted to the right and a little widened. Also, the mean and standard deviation for  $p_c = 0.6$  are 0.677 and 0.094, respectively. In comparison with the optimal case where the mean is 0.538 and the standard deviation is 0.181, the relative standard deviation for  $p_c = 0.6$  is much reduced.

Also, in the case of the out-of-phase spiking group, the distribution { $C^{(l)}$ } with a peak at 0.35 has only positive values in the range of (0.09, 0.41), and its mean and standard deviations are 0.325 and 0.067, respectively. Unlike the optimal case, there are no out-of-phase spiking patterns with negative values of  $C^{(l)}$ . Only out-of-phase spiking patterns which are developed from the inphase spiking patterns appear, and hence they have just positive values of  $C^{(l)}$ . Thus, the range for  $p_c = 0.6$  is moved to the positive region, and it becomes narrowed, in comparison to the optimal case where the range is (-0.20, 0.39). Also, the relative standard deviation for  $p_c = 0.6$  is much decreased, in comparison to the optimal case where the mean and the standard deviation are 0.102 and 0.328, respectively.

In the optimal case of  $p_c^* = 0.06$ , the in-phase and the outof-phase spiking groups are shown to play their own roles for synaptic plasticity at the PF-PC synapses, respectively. As a result of cooperation in their good-balanced state, effective depression (strong/weak LTD) at the PF-PC synapses occurs, which eventually leads to effective motor learning for the OKR adaptation in the PC-VN-IO system. On the other hand, in the highly-connected case of  $p_c = 0.6$ , the in-phase spiking group becomes a dominant one, and hence good balance between the in-phase and the out-ofphase spiking groups is broken up. In such an unbalanced state, contribution of the out-of-phase spiking group to the synaptic plasticity at the PF-PC synapses is decreased so much, which is clearly shown below.

Fig. 15 shows change in synaptic weights of active PF-PC synapses and firing activity of PCs during learning in the highlyconnected case of  $p_c = 0.6$ . Cycle-evolution of bin-averaged synaptic weights  $\langle \tilde{J}(t) \rangle$  (solid circles) of active PF signals is shown in Figs. 15(a1)–15(a5). For comparison, data for  $\langle \tilde{J}(t) \rangle$  (open circles) in the optimal case of  $p_c^* = 0.06$  are also given. In comparison with the optimal case, the bin-averaged synaptic weights  $\langle \tilde{J}(t) \rangle$  at the middle stage are less depressed, while at the initial and the final stage, they are more depressed. Thus, the modulation of  $\langle \tilde{J}(t) \rangle$  is reduced so much. This small modulation is distinctly in contrast to the big modulation in the optimal case.  $\langle \tilde{J}(t) \rangle$  also becomes saturated at about the 300th cycle, as in the optimal case.

In this highly-connected case of  $p_c = 0.6$ , less depression (in the bin-averaged synaptic weights  $\langle \tilde{J}(t) \rangle$ ) at the middle stage may be easily understood in the following way. We note that, at the middle stage, the in-phase spiking group makes a dominant contribution to  $\langle \tilde{J}(t) \rangle$  (i.e., the contribution of the out-ofphase spiking group to  $\langle \tilde{J}(t) \rangle$  may be negligible), as discussed in Fig. 10 in the optimal case of  $p_c = 0.06$ . However, the degree of depression changes depending on the relative phase difference between the in-phase PF (student) signals and the error-teaching (instructor) CF signals. The CF signals are in-phase ones with respect to the firing rate  $f_{DS}(t)$  of the Poisson spike trains for the IO desired signal (for a desired eye-movement) in Fig. 1(b2), and the in-phase PF signals are in-phase ones relative to the instantaneous whole-population spike rate  $R_{GR}(t)$ . We note that, depending on  $p_c$ , the reference signal  $R_{GR}(t)$  for the inphase PF signals has a varying "matching" degree relative to the sinusoidally-modulating IO desired signal  $f_{DS}(t)$ . Here, the matching degree  $\mathcal{M}_d$  is quantitatively given by the cross-correlation at the zero-time lag [*Corr*<sub>M</sub>(0)] between  $R_{GR}(t)$  and  $f_{DS}(t)$ :

$$Corr_{\rm M}(\tau) = \frac{\overline{\Delta f_{\rm DS}(t+\tau)\Delta R_{\rm GR}(t)}}{\sqrt{\Delta f_{\rm DS}^2(t)}\sqrt{\Delta R_{\rm GR}^2(t)}},\tag{41}$$

where  $\Delta f_{DS}(t) = f_{DS}(t) - \overline{f_{DS}(t)}$ ,  $\Delta R_{GR}(t) = R_{GR}(t) - \overline{R_{GR}(t)}$ , and the overline denotes the time average over a cycle.

With increasing  $p_c$ , the top part of  $R_{GR}(t)$  becomes more broadly flattened, and hence its matching degree with respect to  $f_{DS}(t)$  is decreased. For  $p_c = 0.6$ ,  $\mathcal{M}_d = 0.625$ , which is smaller than that ( $\mathcal{M}_d = 0.857$ ) in the optimal case of  $p_c^* = 0.06$ . Hence, due to decrease in the matching degree between  $R_{GR}(t)$ and  $f_{DS}(t)$ , on average, in-phase PF signals for  $p_c = 0.6$  are less depressed by the CF signals than those in the optimal case. Thus, in the highly-connected case of  $p_c = 0.6$ , the bin-averaged synaptic weights  $\langle \tilde{J}(t) \rangle$  (solid circles) at the middle stage (with a dominant in-phase spiking group) are less depressed than those (open circles) in the optimal case. In this way, at the middle stage where the contribution of the in-phase spiking group is dominant, the depression degree for  $\langle \tilde{J}(t) \rangle$  is determined by the matching degree  $\mathcal{M}_d$  between  $R_{GR}(t)$  and  $f_{DS}(t)$ .

More depression (in the bin-averaged synaptic weights  $\langle J(t) \rangle$ ) at the initial and the final stage for  $p_c = 0.6$  can also be clearly understood as follows: In the optimal case, at the initial and the final stages, the firing fractions  $F^{(G)}(t)$  of the in-phase (51%) and the out-of-phase (49%) spiking groups are nearly the same (see Eq. (35) and Fig. 10). Hence, at the initial and the final stage, somewhat less LTD occurs at the PF-PC synapses, in contrast to strong LTD at the middle stage, because both the out-of-phase spiking group (with weak LTD) and the in-phase spiking group make contributions together. However, in the highly-connected case of  $p_c = 0.6$ , the fraction of the in-phase spiking group is so much increased to 81.5%, in comparison to the optimal case (50.2%). Hence, the firing fraction of the in-phase spiking group becomes much larger even at the initial and the final stage  $(F^{(G)}(t) = 0.82)$ . Thus, the bin-averaged synaptic weights  $\langle I(t) \rangle$ (solid circles) at the initial and the final stage are more depressed than those (open circles) in the optimal case. In this way, at the initial and the final stage, the depression degree for  $\langle J(t) \rangle$  is determined by the relative fractions of the in-phase and the outof-phase spiking groups; as the fraction of the in-phase spiking group is increased,  $\langle J(t) \rangle$  is more depressed.

Figs. 15(b1) and 15(b2) show plots of cycle-averaged mean  $\langle \tilde{J}(t) \rangle$  and modulation  $\mathcal{M}_J$  for  $\langle \tilde{J}(t) \rangle$  versus cycle, respectively;  $p_c = 0.6$  (solid circles) and  $p_c^* = 0.06$  (open circles). Both the cycle-averaged mean  $\langle \tilde{J}(t) \rangle$  and the modulation  $\mathcal{M}_J$  for  $\langle \tilde{J}(t) \rangle$  become saturated at about the 300th cycle. With increasing the cycle, the cycle-averaged mean  $\langle \tilde{J}(t) \rangle$  decreases from 1 to 0.374 due to LTD at the PF-PC synapses, which is similar to the optimal case of  $p_c^* = 0.06$  where  $\langle \tilde{J}(t) \rangle$  decreases from 1 to 0.372. On the other hand, the modulation  $\mathcal{M}_J$  increases very slowly from 0 to 0.023, in contrast to the optimal case with a big modulation where it increases quickly from 0 to 0.112. When compared with the optimal case, bin-averaged synaptic weights  $\langle \tilde{J}(t) \rangle$  at the initial and the final stage come down more rapidly (i.e., they are more depressed), while at the middle stage, they come down

relatively slowly (i.e., they are less depressed). This kind of lesseffective synaptic plasticity at the PF-PC synapses arises due to the decreased matching degree of Eq. (41) between  $R_{GR}(t)$  and  $f_{DS}(t)$  (leading to less depression at the middle stage) and the much-increased fraction of in-phase spiking group (resulting in more depression at the initial and the final stage). As a result, the modulation  $\mathcal{M}_J$  makes a slow increase to its saturated value (= 0.023), which is markedly in contrast to the optimal case with a big modulation.

We next consider the effect of PF-PC synaptic plasticity with a reduced small modulation on the subsequent learning process in the PC-VN system. Figs. 15(c1)-15(c5) show cycle-evolution of realization-averaged instantaneous population spike rate  $\langle R_{PC}(t) \rangle_r$ of the PCs (number of realizations: 100);  $p_c = 0.6$  (solid line) and  $p_c^* = 0.06$  (dotted line). As a result of PF-PC synaptic plasticity,  $\langle \hat{R}_{PC}(t) \rangle_r$  becomes lower at the middle stage than at the initial and the final stage. Thus, like the case of  $\langle \tilde{J}(t) \rangle$ ,  $\langle R_{PC}(t) \rangle_r$  forms a well-shaped curve, and it becomes saturated at about the 300th cycle. At the middle stage,  $\langle R_{PC}(t) \rangle_r$  (solid line) for  $p_c = 0.6$  is larger than that (dotted line) in the optimal case of  $p_c^* = 0.06$  due to less-depressed  $\langle J(t) \rangle$ . On the other hand, at the initial and the final stages  $\langle R_{PC}(t) \rangle_r$  (solid line) for  $p_c = 0.6$  is smaller than that (dotted line) in the optimal case of  $p_c^* = 0.06$  because of moredepressed  $\langle J(t) \rangle$ . As a result of such less-effective PF-PC synaptic plasticity, the modulation of  $\langle R_{PC}(t) \rangle_r$  for  $p_c = 0.6$  becomes so small, in contrast to the optimal case with a big modulation (which arises from the effective PF-PC synaptic plasticity).

Figs. 15(d1) and 15(d2) show plots of cycle-averaged mean  $\overline{\langle R_{PC}(t) \rangle_r}$  and modulation  $\mathcal{M}_{PC}$  for  $\langle R_{PC}(t) \rangle_r$  versus cycle, respectively;  $p_c = 0.6$  (solid circles) and  $p_c^* = 0.06$  (open circles). Both  $\overline{\langle R_{PC}(t) \rangle_r}$  and  $\mathcal{M}_{PC}$  become saturated at about the <u>300th cycle</u>. With increasing the cycle, the cycle-averaged mean  $\overline{\langle R_{PC}(t) \rangle_r}$  decreases from 86.1 Hz to 51.8 Hz due to LTD at the PF-PC synapses, which is similar to the optimal case where  $\overline{\langle R_{PC}(t) \rangle_r}$  decreases from 86.1 Hz to 51.7 Hz. On the other hand, the modulations  $\mathcal{M}_{PC}$  increase slowly from 2.6 Hz to 5.6 Hz, which is distinctly in contrast to the optimal case where it increases rapidly from 2.6 Hz to 24.1 Hz. Such a small modulation in  $\langle R_{PC}(t) \rangle_r$  for  $p_c = 0.6$  arises due to the less-effective depression at the PF-PC synapses. These principal PCs of the cerebellar cortex also exert less-effective inhibitory coordination on the VN neuron which evokes OKR eye-movement.

Change in firing activity of the VN neuron during learning in the highly-connected case of  $p_c = 0.6$  (solid circles) is shown in Fig. 16; for comparison, data in the optimal case of  $p_c^* = 0.06$ (open circles) are also given. Figs. 16(a1)-16(a5) show cycleevolution of bin-averaged instantaneous individual firing rate  $f_{\rm VN}(t)$ . It seems to be saturated at about the 300th cycle. Due to the inhibitory coordination of PCs on the VN neuron, the maximum of  $f_{VN}(t)$  occurs at the middle stage, while the minima appear at the initial and the final stage. Thus,  $f_{VN}(t)$  forms a bellshaped curve, in contrast to the well-shaped curves of  $\langle R_{PC}(t) \rangle_r$ .  $f_{VN}(t)$  (solid circles) at the middle stage is smaller than that (open circles) in the optimal case of  $p_c^* = 0.06$  due to more inhibition of the PCs on the VN neuron, while at the initial and the final stage,  $f_{VN}(t)$  (solid circles) is larger than that (open circles) in the optimal case because of less inhibition of PCs on the VN neuron. As a result of such less-effective inhibitory coordination of the PCs, the modulation of  $f_{VN}(t)$  becomes smaller than that in the optimal case.

Figs. 16(b) and 16(c) show plots of cycle-averaged mean  $f_{VN}(t)$  and modulation  $\mathcal{M}_{VN}$  for  $f_{VN}(t)$ . Both  $\overline{f_{VN}(t)}$  and  $\mathcal{M}_{VN}$  become saturated at about the 300th cycle.  $\overline{f_{VN}(t)}$  increases from 44.3 Hz to 71.4 Hz, which is nearly the same as that in the optimal case where it increases from 44.3 Hz to 71.5 Hz.  $\mathcal{M}_{VN}$  also increases slowly from 20.2 Hz to 22.6 Hz, which is in contrast to the optimal

![](_page_22_Figure_2.jpeg)

**Fig. 16.** Change in firing activity of the VN neuron during learning in the highly-connected case of  $p_c = 0.6$ ; for comparison, data in the optimal case of  $p_c^* = 0.06$ are also given. (a1)-(a5) Cycle-evolution of bin-averaged instantaneous individual firing rates  $f_{VN}(t)$ ; the bin size is  $\Delta t = 100$  ms. Plots of (b) cycle-averaged mean  $\overline{f_{VN}(t)}$  and (c) modulation  $\mathcal{M}_{VN}$  for  $f_{VN}(t)$ , and (d) learning gain degree  $\mathcal{L}_g$  versus cycle. In (a1)-(a5) and (b)-(d), solid (open) circles represent data for  $p_c = 0.6$  (0.06).

case where  $\mathcal{M}_{\text{VN}}$  increases quickly from 20.2 Hz to 32.5 Hz. Then, the learning gain degree  $\mathcal{L}_g$ , given by the normalized gain ratio, is shown in Fig. 16(d).  $\mathcal{L}_g$  increases from 1 and becomes saturated at  $\mathcal{L}_g^* \simeq 1.118$  at about the 300th cycle which is smaller than the saturated learning gain degree  $\mathcal{L}_g^*$  ( $\simeq 1.608$ ) in the optimal case. Consequently, due to the less-effective coordination of PCs, the saturated learning gain degree becomes smaller.

To sum up the highly-connected case of  $p_c = 0.6$ , good balance between the in-phase and the out-of-phase spiking group in the optimal case ( $\mathcal{R}^* \simeq 1.008$ ) is broken up, because the fraction of in-phase spiking group becomes dominant ( $\mathcal{R} \simeq 4.405$ ). Thus, the diversity degree for the spiking patterns of the GR clusters is so much decreased ( $\mathcal{D} \simeq 0.204$ ), in comparison with  $\mathcal{D}^*$  ( $\simeq$ 1.613) in the optimal case. Due to increase in the fraction of the in-phase spiking group and decrease in the matching degree between  $R_{\rm GR}(t)$  and  $f_{\rm DS}(t)$ , synaptic plasticity at the PF-PC synapses becomes less effective, which also results in less-effective motor learning for the OKR eye-movement.

We next consider the lowly-connected cases of  $p_c = 0.006$ (i.e. 0.6%). Figs. 17(a) and 17(b) show the raster plot of spikes of 10<sup>3</sup> randomly-chosen GR cells and the instantaneous wholepopulation spike rate  $R_{GR}(t)$  in the whole population of GR cells for  $p_c = 0.006$ , respectively. In this lowly-connected case, the inhibitory inputs from GO cells into the GR clusters are so much reduced, and the excitatory MF signals into the GR clusters become dominant inputs. Hence, the raster plot of spikes becomes more dense, and  $R_{GR}(t)$  becomes more similar to the firing rate  $f_{MF}(t)$  of the Poisson spike train for the MF signal in Fig. 1(b1), in contrast to the highly-connected case of  $p_c = 0.6$  with broadly flattened top part in  $R_{GR}(t)$  [see Fig. 14(b)].

Due to so much decrease in inhibitory inputs from the GO cells into the GR clusters, only the in-phase spiking group [where spiking patterns are similar to  $f_{\rm MF}(t)$ ] appears (i.e., all the out-of-phase spiking group disappears), which is distinctly in contrast to the optimal case of  $p_c^* = 0.06$  where diverse spiking groups such as the in-phase, anti-phase, and complex out-of-phase spiking groups coexist. The distribution of conjunction indices { $C^{(I)}$ } of the

GR clusters ( $C^{(l)}$ : representing the similarity degree between the spiking behavior [ $R_{GR}^{(l)}(t)$ : instantaneous cluster spike rate] of the *I*th GR cluster and that [ $R_{GR}(t)$ ] in the whole population) is shown in Fig. 17(c). In comparison to the optimal case in Fig. 6(a), the distribution is shifted to the positive region, due to existence of only the in-phase GR clusters with positive values of  $C^{(I)}$ . Thus, it has a peak at 0.75, and its range is (0.32, 0.98).

In this lowly-connected case of  $p_c = 0.006$ , the mean and the standard deviation for the distribution  $\{C^{(l)}\}\$  are 0.737 and 0.129, respectively, which is in contrast to the optimal case with the smaller mean (= 0.320) and the larger standard deviation (= 0.516). Then, the diversity degree  $\mathcal{D}$  of the spiking patterns in all the GR clusters, given by the relative standard deviation for the distribution  $\{C^{(l)}\}\$  [see Eq. (26)], is  $\mathcal{D} \simeq 0.175$  which is much smaller than  $\mathcal{D}^*$  ( $\simeq$  1.613) in the optimal case. Consequently, the degree in diverse recoding in the granular layer is so much reduced in the lowly-connected case of  $p_c = 0.006$ , due to existence of only the in-phase spiking group without the out-of-phase spiking group.

We also compare the lowly-connected case of  $p_c = 0.006$  with the highly-connected case of  $p_c = 0.6$  in Fig. 14. The diversity degree  $\mathcal{D}$  ( $\simeq$  0.204) for  $p_c$  = 0.6 is also much reduced in comparison with  $\mathcal{D}^*$  ( $\simeq 1.613$ ) in the optimal case. However, it is a little larger than  $\mathcal{D} \simeq 0.175$  for  $p_c = 0.006$ , because for  $p_c = 0.6$ a minor out-of-phase spiking group with positive values of  $C^{(I)}$ exists, along with the major in-phase spiking group. We also note that, the in-phase spiking patterns in both the lowly- and the highly-connected cases have completely different waveforms. For  $p_c = 0.006$  the in-phase spiking patterns are more similar to the sinusoidally-modulating MF signal  $f_{MF}(t)$ , while those for  $p_c = 0.6$ are more different from  $f_{MF}(t)$  due to broad flatness in their top part. Thus, the matching degree  $M_d$  (= 0.981) of Eq. (41) for  $p_c = 0.006$  is larger than that (= 0.625) for  $p_c = 0.6$ . In this way, there are two independent ways via increase or decrease in  $p_c$ from the optimal value  $p_c^*$  (= 0.06) to break up the good balance between the in-phase and the out-of-phase spiking groups, which

![](_page_23_Figure_1.jpeg)

**Fig. 17.** Lowly-connected case of  $p_c = 0.006$ . (a) Raster plot of spikes of  $10^3$  randomly chosen GR cells. (b) Instantaneous whole-population spike rate  $R_{GR}(t)$  in the whole population of GR cells. Band width for  $R_{GR}(t)$ : h = 10 ms. (c) Distribution of conjunction indices  $\{C^{(l)}\}$  for the GR clusters in the whole population. (d1) Plot of matching degree  $\mathcal{M}_d$  versus  $p_c$ . (d2) Plot of the minimum  $\langle \tilde{J}(t) \rangle_{min}$  versus  $p_c$ . (e1)–(e5) Cycle-evolution of bin-averaged instantaneous individual firing rates  $f_{VN}(t)$ ; the bin size is  $\Delta t = 100$  ms. (f) Plot of learning gain degree  $\mathcal{L}_g$  (solid circles) versus cycle; for comparison, data (open circles) in the optimal case of  $p_c^* = 0.06$  are also given.

results in decrease in the diversity degree in recoding of the GR cells.

As a result of reduced diversity in recoding of the GR cells, the modulation for the bin-averaged synaptic weights  $\langle \tilde{J}(t) \rangle$  is much decreased, in contrast to the big modulation in the optimal case of  $p_c^* = 0.06$ . Fig. 17(d1) shows the plot of the matching degree  $\mathcal{M}_d$  between  $R_{GR}(t)$  and  $f_{DS}(t)$  versus  $p_c$ . With decreasing  $p_c$ ,  $\mathcal{M}_d$ increases monotonically. Hence,  $M_d$  (= 0.981) for  $p_c$  = 0.006 is larger than that (= 0.857) in the optimal case of  $p_c = 0.06$ . Due to increase in  $\mathcal{M}_d$ , at the middle stage in the lowly-connected case of  $p_c = 0.006$ , more depression in  $\langle \tilde{J}(t) \rangle$  is expected to occur, in comparison to the optimal case. Fig. 17(d2) shows the plot of the minimum  $\langle J(t) \rangle_{min}$  of the well-shaped curve for  $\langle J(t) \rangle$  (appearing at t = 1000 ms) versus  $p_c$ . We note that, as  $p_c$  is decreased from  $p_c^*$  (= 0.06),  $\langle \tilde{J}(t) \rangle_{min}$  decreases so much slowly. Hence,  $\langle \tilde{J}(t) \rangle_{min}$  (= 0.267) for  $p_c = 0.006$  becomes just a little smaller than that (= 0.274) in the optimal case. Thus, for  $p_c = 0.006$  $\langle \tilde{J}(t) \rangle$  is only a little more depressed than that in the optimal case of  $p_c = 0.06$ . At the initial and the final stage, only in-phase spiking group exists, unlike the optimal case where both the inphase and the out-of-phase spiking groups coexist. Hence, much more depression in  $\langle I(t) \rangle$  occurs in comparison to the optimal case. Thus, modulation in  $\langle \tilde{J}(t) \rangle$  for  $p_c = 0.006$  becomes much reduced, in comparison to the optimal case.

To make this point more clear, we divide the distribution of conjunction indices { $C^{(l)}$ } with a range (0.32, 0.98) in Fig. 17(c) into the two parts by taking the mean (= 0.737) as a reference.

Thus, the 1st in-phase spiking sub-group has its conjunction indices higher than the mean (= 0.737) [i.e., the range is (0.737), 0.98)], while the 2nd in-phase spiking sub-group has its conjunction indices lower than than the mean [i.e., the range is (0.32, 0.737)]. In this way, the whole in-phase spiking group is decomposed into the two sub-groups. Then, we obtain the firing fraction  $F_{i}^{(G)}$  of active PF signals in the G spiking group [see Eq. (35)], where G corresponds to the 1st or the 2nd in-phase spiking subgroup. Similar to Figs. 10(d1)–10(d5), the firing fraction  $F^{(G)}(t)$ for the 1st (2nd) in-phase spiking sub-group is found to form a bell-shaped (well-shaped) curve. At the initial and the final stages, the firing fractions  $F^{(G)}(t)$  for the 1st and the 2nd in-phase spiking sub-groups are equal (i.e., 50%). On the other hand, at the middle stage, the values of  $F^{(G)}(t)$  for the 1st and the 2nd in-phase spiking sub-group are 0.61 (61%) and 0.39 (39%), respectively. We note that more depression in  $\langle I(t) \rangle$  occurs for the 1st in-phase spiking group because their conjunction indices are larger than those in the 2nd in-phase spiking sub-group. Judging from the ratio of their firing fractions, at the middle stage (0.61 : 0.39),  $\langle \tilde{J}(t) \rangle$  is a little more depressed, in comparison to the initial and the final stage (0.5 : 0.5). Thus, in the lowly-connected case of  $p_c = 0.006$ , a small modulation appears in  $\langle \tilde{I}(t) \rangle$ , in contrast to the big modulation in the optimal case. The less-effective synaptic plasticity at the PF-PC synapses leads to reduced modulation in the realization-averaged instantaneous population spike rate  $\langle R_{\rm PC}(t) \rangle_r$  of the principal PCs in the cerebellar cortex which also exert less-effective inhibitory coordination on the VN neuron which produces the final output of the cerebellum (i.e., OKR eve-movement). Figs. 17(e1)-17(e5) show cycle-evolution of the bin-averaged instantaneous firing rate  $f_{VN}(t)$  (solid circles) of the VN neuron for  $p_c = 0.006$ ; for comparison, data (open circles) in the optimal case of  $p_c^* = 0.06$  are also given. With increasing the cycle,  $f_{VN}(t)$  seems to be saturated at about the 300th cycle.  $f_{\rm VN}(t)$  (solid circles) at the middle stage is a little larger than that (open circles) in the optimal case due to a little less inhibition of the PCs on the VN neuron for  $p_c = 0.006$ . On the other hand, at the initial and the final stage,  $f_{VN}(t)$  (solid circles) is much larger than that (open circles) in the optimal case because of much less inhibition of PCs on the VN neuron in comparison to the optimal case.

In this way, as a result of less-effective inhibitory coordination of the PCs, the modulation of  $f_{\rm VN}(t)$  becomes much smaller than that in the optimal case. Plots of the learning gain degree  $\mathcal{L}_g$ [corresponding to the modulation gain ratio for  $f_{\rm VN}(t)$ ] are shown in Fig. 17(f) in both the lowly-connected case of  $p_c = 0.006$  (solid circles) and the optimal case of  $p_c^* = 0.06$  (open circles). For  $p_c = 0.006 \mathcal{L}_g$  increases very slowly and becomes saturated at  $\mathcal{L}_g^* \simeq 1.099$  at about the 300th cycle. This saturated learning gain degree  $\mathcal{L}_g^* (\simeq 1.099)$  is much smaller than that ( $\mathcal{L}_g^* \simeq 1.608$ ) in the optimal case. Consequently, low diversity in recoding of the GR cells for  $p_c = 0.006$  results in less-effective motor learning for the OKR adaptation.

Before proceeding to the general variation of  $p_c$ , we now summarize the main points obtained in the above highly-connected ( $p_c = 0.6$ ) and lowly-connected ( $p_c = 0.006$ ) cases. In both the highly- and the lowly-connected cases, their diversity degrees  $\mathcal{D}$  [= 0.204 ( $P_c = 0.6$ ) and 0.175 ( $p_c = 0.006$ )] for spiking patterns of the GR clusters are so much decreased in comparison to that ( $\mathcal{D} = 1.613$ ) in the optimal case of  $p_c^* = 0.06$ , because the fraction of the in-phase spiking group is increased from 50.2% to 81.5% ( $p_c = 0.6$ ) and 100% ( $p_c = 0.006$ ). At the initial and the final stage, the degree of depression in the bin-averaged (normalized) synaptic weight  $\langle \tilde{J}(t) \rangle$  is determined by the firing fractions  $F^{(G)}$  of Eq. (35) of the in-phase and the out-of-phase spiking groups. For  $p_c = 0.6$  and 0.006,  $F^{(i)}$  of the in-phase spiking group is 0.82 (82%) and 1.0 (100%), respectively, in comparison to 0.51 (51%) in

![](_page_24_Figure_1.jpeg)

**Fig. 18.** Strong correlation between the diversity degree  $\mathcal{D}$  and the saturated learning gain degree  $\mathcal{L}_{g}^{*}$ . (a) Fraction of in-phase (solid circles) and out-of-phase (open circles) spiking groups. (b) Plot of diversity degree  $\mathcal{D}$  for the spiking patterns of all the GR clusters versus  $p_c$ . (c) Saturated learning gain degree  $\mathcal{L}_{g}^{*}$  versus  $p_c$ . (d) Plot of  $\mathcal{L}_{g}^{*}$  versus  $\mathcal{D}$ .

the optimal case of  $p_c^* = 0.06$ . Hence, for  $p_c = 0.6$  and 0.006, due to increased firing fraction of the in-phase spiking group (with strong LTD), more depression in  $\langle \tilde{J}(t) \rangle$  occurs at the initial and the final stage of cycle, which is in contrast to the optimal case (e.g., see Fig. 15).

In the middle stage, the in-phase spiking group make a dominant contribution, independently of  $p_c$ ;  $F^{(i)} = 0.94$  (94% for  $p_c^* = 0.06$ ), 0.98 (98% for  $p_c = 0.6$ ), and 1.0 (100% for  $p_c = 0.006$ ). However, the effect of the dominant in-phase spiking group on depression in  $\langle \tilde{J}(t) \rangle$  varies depending on  $p_c$ . The degree of depression in  $\langle \tilde{J}(t) \rangle$  is determined by the matching degree  $\mathcal{M}_d$  of Eq. (41) between the instantaneous whole-population spike rate  $R_{\rm GR}(t)$  and the IO desired signal  $f_{\rm DS}(t)$ ;  $R_{\rm GR}(t)$  plays a "reference" signal for the in-phase "student" PF signals and the error-teaching instructor CF signals are in-phase with  $f_{\rm DS}(t)$ . Depending on  $p_c$ , the reference signal  $R_{\rm GR}(t)$  has a changing matching degree  $\mathcal{M}_d$ with respect to  $f_{\rm DS}(t)$ . With decreasing  $p_c$ ,  $\mathcal{M}_d$  is monotonically increased [see Fig. 17(d1)] [i.e,  $R_{\rm GR}(t)$  becomes more similar to the sinusoidal IO desired signal  $f_{\rm DS}(t)$ ].

In the highly-connected case of  $p_c = 0.6$ ,  $\mathcal{M}_d = 0.625$ , which is smaller in comparison with  $\mathcal{M}_d = 0.857$  in the optimal case of  $p_c^* = 0.06$ . Due to decrease in  $\mathcal{M}_d$  for  $p_=0.6$ , in the middle stage (where the in-phase spiking group is dominant), less depression in  $\langle \tilde{J}(t) \rangle$  occurs (see Fig. 15). Consequently, for  $p_c = 0.6$  the modulation in  $\langle \tilde{J}(t) \rangle$  decreases, in comparison to that in the optimal case, due to more depression at the initial and the final stage and less depression at the middle stage.

In the lowly-connected-connected case of  $p_c = 0.006$ ,  $\mathcal{M}_d = 0.981$ , which is larger than that ( $\mathcal{M}_d = 0.857$ ) in the optimal case of  $p_c^* = 0.06$ . However, as shown in Fig. 17(d2), the minimum  $\langle \tilde{J}(t) \rangle_{min}$  of the well-shaped curve for  $\langle \tilde{J}(t) \rangle$  decreases so much slowly with decreasing  $p_c$ . Thus, for  $p_c = 0.006$  only a little more depression in  $\langle \tilde{J}(t) \rangle$  in comparison with the optimal case. As explained in the above, at the initial and the final stage, only

the in-phase spiking group (with strong LTD) exists, in contrast to the optimal case where both the out-of-phase (with weak LTD) and the in-phase spiking groups. Hence, much more depression takes place in comparison to the optimal case. As a result, for  $p_c = 0.006$  the modulation in  $\langle \tilde{J}(t) \rangle$  is decreased, in comparison with that in the optimal case, because of much more depression at the initial and the final stage and a little more depression at the middle stage.

Such decrease in modulation in  $\langle \bar{J}(t) \rangle$  for  $p_c = 0.6$  and 0.006 causes a small modulation in firing activity  $R_{PC}(t)$  of the PCs, which then exerts less-effective inhibitory coordination on the VN neuron (which evokes the OKR). Consequently, a small modulation in the firing activity  $f_{VN}(t)$  of the VN neuron arises [see Fig. 16 for  $p_c = 0.6$  and Fig. 17 for  $p_c = 0.006$ ]. Consequently, for  $p_c = 0.6$  and 0.006 where the diversity degree in recoding of GR cells is decreased, less-effective motor learning for the OKR adaptation occurs, in comparison to the optimal case of  $p_c = 0.06$ .

Finally, based on the above two examples for the highly- and the lowly-connected cases, we investigate dependence of the diversity degree  $\mathcal{D}$  for the spiking patterns of the GR clusters and the saturated learning gain degree  $\mathcal{L}_g^*$  on  $p_c$  by varying it from the optimal value ( $p_c^* = 0.06$ ). Fig. 18(a) shows plots of fractions of the in-phase and the out-of-phase spiking groups versus  $p_c$ . The fraction of the in-phase spiking group (solid circles) forms a well-shaped curve with a minimum at  $p_c = p_c^*$  (= 0.06), while the fraction of the out-of-phase spiking group (open circles) forms a bell-shaped curve with a maximum at the optimal value of  $p_c^* =$ 0.06. For sufficiently small  $p_c$ , we have two sample cases where the fraction of in-phase spiking group is 1 (i.e., the fraction of outof-phase spiking group is 0). We note that, in the optimal case of  $p_c^* = 0.06$ , fractions of the in-phase (50.2%) and the out-ofphase spiking (49.8%) groups are well balanced (i.e., good balance between the in-phase and the out-of-phase spiking groups). As  $p_c$  is changed (i.e., increased or decreased) from  $p_c^*$ , the fraction of the in-phase spiking group increases, and then the spikinggroup ratio  $\mathcal{R}$  (i.e., the ratio of the fraction of the in-phase spiking group to that of the out-of-phase spiking group) increases from the golden spiking-group ratio  $\mathcal{R}^*$  ( $\simeq 1.008$ ) in the optimal case.

Figs. 18(b) and 18(c) show plots of the diversity degree  $\mathcal{D}$  for the spiking patterns of the GR clusters and the saturated learning gain degree  $\mathcal{L}_{\sigma}^{*}$  versus  $p_{c}$ , respectively. The diversity degrees  $\mathcal{D}$ forms a bell-shaped curve with a maximum  $\mathcal{D}^*$  ( $\simeq$  1.613) in the optimal case of  $p_c^* = 0.06$  with the golden spiking-group ratio  $\mathcal{R}^* \simeq 1.008$  (i.e., good balance between the in- and the out-of-phase spiking group). We note that, in this optimal case where the recoding of the GR cells is the most diverse, the saturated learning gain degree  $\mathcal{L}^*$  also has its maximum ( $\mathcal{L}_g^* \simeq$ 1.608). As  $p_c$  is changed (i.e., increased or decreased) from the optimal value (= 0.06), the spiking-group ratio  $\mathcal{R}$  is increased, because of increase in the fraction of in-phase spiking group. Then, the diversity degree  $\mathcal{D}$  in recodings of the GR cells becomes decreased, which also results in decrease in the saturated learning gain degree  $\mathcal{L}_g^*$  from the maximum. Thus,  $\mathcal{L}_g^*$  also forms a bellshaped curve, as in the case of  $\mathcal{D}$ , and they have their maxima at the optimal values ( $p_c^* = 0.06$ ).

Fig. 18(d) shows a plot of  $\mathcal{L}_g^*$  versus  $\mathcal{D}$ . As shown clearly in Fig. 18(d), both  $\mathcal{L}_g^*$  and  $\mathcal{D}$  have a strong correlation with the Pearson's correlation coefficient  $r \simeq 0.9998$ . Consequently, the more diverse the recoding of the GR cells, the more effective the motor learning for the OKR adaptation.

#### 4. Summary and discussion

We are interested in the gain adaptation of OKR for the eye movement. Various experimental works on the OKR have been done in rabbits, mice, and zebrafishes (Harvey et al., 1997; Huang & Neuhauss, 2008; Iwashita et al., 2001; Matsuno et al., 2016; Nagao, 1983, 1988; Scheetz et al., 2017; Tabata et al., 2010). Moreover, some features of the OKR adaptation were successfully reproduced through computational simulations in the adaptive filter model (Gomi & Kawato, 1992) and the spiking network model (Yamazaki & Nagao, 2012). However, effects of diverse recoding of GR cells on the OKR adaptation in previous computational works are necessary to be more clarified in several dynamical aspects. Particularly, the previous works lacked complete dynamical classification of diverse spiking patterns of GR cells and their association with the error-teaching CF signals. We note that such dynamical classification of diverse recoding of GR cells may be a basis for clear understanding of the synaptic plasticity at the PF-PC synapses and the subsequent learning progress in the PC-VN-IO system.

Our cerebellar ring network is essentially the same as the square-lattice network in the previous works (Yamazaki & Nagao, 2012; Yamazaki & Tanaka, 2007a). The authors explained that the structural parameters of their square-lattice network model were selected on the basis of the anatomical observations for the cat (Palkovits, Magyar, & Szentágothai, 1971a, 1971b, 1972). The parameters for the single neuron models and the synaptic currents were also adopted from the known physiological data. Hence, the square-lattice network may be regarded as a biological neural network, based on the anatomical and the physiological data. For the effective study of OKR, we first introduced one-dimensional ring network. This ring network with simple architecture has advantage for computational and analytical efficiency, and its visual representation may be easily made. We note that the parameters for the architecture, the single LIF neuron models, and the synaptic currents in our ring network were chosen, based on those in the biological square-lattice network (Yamazaki & Nagao, 2012; Yamazaki & Tanaka, 2007a). Moreover, we also employed a refined rule for the synaptic plasticity, based on the experimental results (Safo & Regehr, 2008). In this sense, the ring network seems to be more effective for the study of OKR than the square-lattice network.

For the first time, we made complete quantitative classification of diverse spiking patterns in the GR clusters via introduction of the conjunction index  $\{C^{(l)}\}\$  and the diversity degree  $\mathcal{D}$ . Each spiking pattern in the Ith GR cluster may be characterized in terms of its conjunction index  $C^{(l)}$ , denoting the degree for the association of the spiking behavior of each Ith GR cluster [characterized by the instantaneous cluster spike rate  $R_{GR}^{(l)}(t)$ ] with the population-averaged firing activity in the whole population [given by the instantaneous whole-population spike rate  $R_{GR}(t)$ ]. Then, the whole spiking patterns are decomposed into the two types of in-phase and out-of-phase spiking groups which are inphase and out-of-phase with respect to the "reference" signal  $R_{GR}(t)$ , respectively. Furthermore, the degree of diverse recoding of the GR cells may be quantified in terms of the diversity degree  $\mathcal{D}$ , given by the relative standard deviation in the distribution of  $\{C^{(l)}\}$ . Thus,  $\mathcal{D}$  gives a quantitative measure for diverse recoding of GR cells. Dynamical origin for the appearance of diverse spiking patterns (i.e., outputs of the GR clusters) has also been investigated. It has thus been found that diverse total synaptic inputs (including both the excitatory inputs via MFs and the inhibitory inputs from the pre-synaptic GO cells) into the GR clusters lead to production of diverse spiking patterns in the GR clusters. Based on the dynamical classification of diverse spiking patterns in the GR clusters, we made intensive investigations on the effect of diverse recoding of GR cells on the OKR adaptation (i.e., its effect on the synaptic plasticity at the PF-PC synapses and the subsequent learning process in the PC-VN-IO system). To the best of our knowledge, this type of approach, based on the in-phase and the out-of-phase spiking groups, is unique for the study of OKR.

In our refined rule for the synaptic plasticity, we discuss the major LTD ( $\Delta$ LTD<sup>(1)</sup>), the minor LTD ( $\Delta$ LTD<sup>(2)</sup>), and  $\Delta$ LTP through comparison of their magnitudes. Synaptic plasticity (LTD or LTP) at PF-PC synapses occurs depending on the relative time difference  $\Delta t$  (=  $t_{CF} - t_{PF}$ ) between the activation times of the error-teaching instructor CF and the student PF signals. An instructor CF activation at a time t gives rise to the 1st type of major LTD of Eq. (14) in conjunction with earlier student PF signals in an effective range of  $0 < \Delta t < \Delta t_{\star}^{*}$  (= 277.5 ms). In the absence of such CF activation, a student PF firing at the time t may cause LTD or LTP, depending on the presence of earlier CF firings in an effective range. The PF activation at the time t causes another LTD in association with earlier CF activations in an effective range of  $\Delta t_1^*$  (= -117.5 ms) <  $\Delta t$  < 0. However, the probability of occurrence of earlier CF activations within the effective range is very low because the mean firing rates of CF signals are  $\sim$ 1.5 Hz). Hence, this 2nd type of LTD of Eq. (15) is a minor one in comparison to the 1st type of major LTD. In the absence of this kind of earlier CF firings,  $\triangle$ LTP of Eq. (16) takes place due to the PF activation alone. In the previous works (Yamazaki & Nagao, 2012; Yamazaki & Tanaka, 2007a), only the major LTD was considered along with  $\triangle$ LTP (i.e., minor LTD was ignored). Even in these cases, we expect that essentially the same results as ours could be obtained, because the major LTD is a dominant one.

In the synaptic plasticity rule of Eqs. (14)–(16), we note that the update rate  $\delta_{LTP}$  in Eq. (16) for the  $\Delta$ LTP is just  $\frac{1}{10}$  of the update rate  $\delta_{LTD}$  in Eq. (14) for the major LTD ( $\Delta$ LTD<sup>(1)</sup>). Hence, the effect of  $\triangle$ LTP is smaller than that of the major LTD, and the major LTD becomes the dominant synaptic modification at the PF-PC synapses. Due to the dominant effect of the major LTD, with increasing the cycle, the cycle-averaged bin-averaged (normalized) synaptic weight  $\langle \tilde{J}(t) \rangle$  decreases and becomes saturated at about the 300th cycle, as shown in Fig. 8(c). To examine the saturation of  $\langle \tilde{J}(t) \rangle$ , we consider the cycle-summed  $\Delta$ LTD ( $\langle \Delta$ LTD $\rangle_s$ ) [ $\Delta$ LTP  $(\langle \Delta LTP \rangle_s)$ ] in each cycle, given by the sum of  $\Delta LTD_{ii}$  ( $\Delta LTP_{ij}$ ) occurring at all the active PF-PC synapses during the whole cycle (0 < t < 2000 ms), where  $\Delta \text{LTD}_{ij} = \Delta \text{LTD}_{ii}^{(1)} + \Delta \text{LTD}_{ii}^{(2)}$ [see Eqs. (14)–(16)]. In the beginning cycles,  $|\langle \Delta LTD \rangle_s|$  (i.e., the magnitude of  $(\Delta LTD)_s$  is larger than  $(\Delta LTP)_s$ , and hence  $\Delta LTD$  is dominant. However, as the cycle is increased,  $|\langle \Delta LTD \rangle_s|$  decreases monotonically, and becomes saturated at about the 300th cycle, because with increasing the cycle the error-teaching CF instructor signal becomes weaker and saturated at about the 300th cycle [e.g., see Fig. 13(d)]. In this saturated case, the saturated value of  $|\langle \Delta LTD \rangle_s|$  becomes the same as that of  $\langle \Delta LTP \rangle_s$ . Consequently, cycle-level dynamical balance between  $\triangle$ LTD and  $\triangle$ LTP emerges, which leads to the saturation of the cycle-averaged bin-averaged (normalized) synaptic weight  $\langle I(t) \rangle$ , as shown in Fig. 8(c).

Diversely-recoded student PF signals from GR cells and the instructor error-teaching CF signals from the IO neuron are fed into the PCs. We note that the CF signals are always in-phase with respect to the reference signal  $R_{GR}(t)$ . During the whole learning process, the in-phase and the out-of-phase spiking groups have been found to play their own roles, respectively. The strength of LTD changes markedly depending on the type of spiking group (i.e., it varies distinctly depending on the phase difference between the instructor CF and the student PF signals), as shown in Fig. 9. The in-phase student PF signals have been found to be strongly depressed (i.e., strong LTD) by the instructor CF signals, because they are well-matched with the in-phase CF signals [e.g., see Figs. 9(a1)-9(a5)]. On the other hand, the out-ofphase student PF signals have been found to be weakly depressed (i.e., weak LTD) by the instructor CF signals, because they are ill-matched with the in-phase CF signals [e.g., see Figs. 9(b1)-9(b5)]. In this way, occurrence of strong or weak LTD depends just on the phase relation between the instructor CF and the student PF signals. We also expect that such occurrence is nearly independent of detailed form of the time window  $\Delta J_{LTD}(\Delta t)$  of Eq. (17) for the synaptic plasticity. [i.e., the same kind of strong and weak LTD may be expected to occur in the previous framework of synaptic plasticity rule (Yamazaki & Nagao, 2012; Yamazaki & Tanaka, 2007a)].

However, contributions of these in-phase and out-of-phase spiking groups vary depending on the stage of cycle. In the middle stage of each cycle, strong LTD takes place through dominant contributions of the in-phase spiking group, which results in emergence of a minimum of the bin-averaged (normalized) synaptic weight  $\langle \tilde{J} \rangle$  of active PF signals. In contrast, at the initial and the final stage of each cycle, less LTD occurs (i.e., maxima of  $\langle \tilde{J} \rangle$  appear) because both the out-of-phase spiking group with weak LTD and the in-phase spiking group with strong LTD make contributions together. Hence, the bin-averaged synaptic weights  $\langle \tilde{J} \rangle$  have been found to form a well-shaped curve (i.e., appearance of a minimum in the middle stage and maxima at both the initial and final stages). In this way, a big modulation in  $\langle \tilde{J} \rangle$  emerges through interplay of the in-phase and the out-of-phase spiking groups.

Due to this kind of effective depression (i.e., strong/weak LTD) at the PF-PC synapses, the (realization-averaged) instantaneous population spike rate  $\langle R_{PC}(t) \rangle_r$  of PCs (corresponding to the principal outputs of the cerebellar cortex) has been found to form a well-shaped curve with a minimum in the middle stage. Consequently, a big modulation occurs in  $\langle R_{PC}(t) \rangle_r$ . These PCs exert effective inhibitory coordination on the VN neuron (which evokes OKR eye-movement). Thus, the (realization-averaged) instantaneous individual firing rate  $f_{VN}(t)$  of the VN neuron has been found to form a bell-shaped curve with a maximum in the middle stage. The maximum in the middle stage of cycle is formed due to dominant contribution of the in-phase spiking group with strong LTD, while appearance of minima at the initial and the final stage is made via comparable contributions of the in-phase and the out-of-phase spiking groups. In this case, the learning gain degree  $\mathcal{L}_{g}$ , corresponding to the modulation gain ratio (i.e., normalized modulation divided by that at the 1st cycle for  $f_{\rm VN}$ ), has been found to increase with learning cycle and to be saturated at about the 300th cycle.

In the cerebellar ring network, we varied the connection probability  $p_c$  from the GO cells to the GR cells, and investigated dependence of the diversity degree  $\mathcal{D}$  of spiking patterns of GR cells and the saturated learning gain degree  $\mathcal{L}_g^*$  on  $p_c$ . Both  $\mathcal{D}$ and  $\mathcal{L}_{g}^{*}$  have been found to form bell-shaped curves with peaks at the same optimal value of  $p_c^* = 0.06$ . Hence, the average number of pre-synaptic GO cells for each GR cell in our optimal case is  $N_{\rm pre,GO}$  = 10, which seems to be close to that  $(N_{\rm pre,GO} = 8)$  in the square-lattice network whose structural parameters are based on the anatomical data of the cat (Palkovits et al., 1971a, 1971b, 1972; Yamazaki & Nagao, 2012; Yamazaki & Tanaka, 2007a). However, generally N<sub>pre,GO</sub> seems to vary depending on the species of animals (e.g., cat or rat); for the rat, refer to the works Brickley, Cull-Candy, and Farrant (1996) and Jakab and Hámori (1988). Furthermore, Npre, GO may also vary depending on the network models in computational works. Hence, to avoid direct comparison of N<sub>pre,GO</sub> in our computational optimal case with real experimental structural data seems to be reasonable. Here, the optimal case in our computational work corresponds to just the one where both  $\mathcal{D}$  and  $\mathcal{L}_g^*$  have their peak values. Moreover, both the diversity degree  $\mathcal{D}$  of spiking patterns

Moreover, both the diversity degree  $\mathcal{D}$  of spiking patterns of the GR cells and the saturated learning gain degree  $\mathcal{L}_g^*$  have been found to have a strong correlation with Pearson correlation coefficient  $r \simeq 0.9998$ . Consequently, the more diverse the spiking patterns of GR cells, the more effective the motor

learning for the OKR adaptation, which is the main result in our computational works. In the optimal case of  $p_c^* = 0.06$  with the maximum diversity degree  $\mathcal{D}^*$  (= 1.613), the in-phase and the out-of-phase spiking groups keep good balance (i.e., their fractions are nearly equal; spiking-group ratio  $\mathcal{R}^* = 1.008$ ). At the middle stage of each cycle, PF signals are strongly depressed by the instructor CF signals due to dominant contribution of the in-phase spiking group with strong LTD. On the other hand, at the initial and the final stage, both the in-phase (strong LTD) and the out-of-phase (weak LTD) spiking group make comparable contributions together, and hence PF signals are less depressed. This kind of effective depression (i.e., strong/weak LTD) causes a big modulation in synaptic plasticity, which eventually results in effective motor learning for the OKR adaptation. On the other hand, in the highly- or the lowly-connected case (e.g.,  $p_c = 0.6$ or 0.006), the in-phase spiking group becomes dominant, and hence good balance between the in-phase and the out-of-phase spiking groups becomes broken up, and the diversity degree for  $p_c = 0.6$  and 0.006 are so much decreased to  $\mathcal{D} = 0.204$  and 0.175, respectively. In such an unbalanced state, at the initial and the final stage of each cycle, the contribution of the out-of-phase spiking group to the synaptic plasticity at the PF-PC synapses is much decreased, and hence more depression occurs at the initial and the final stage due to major contribution of the in-phase spiking group. This kind of less effective depression at the PF-PC synapses gives rise to a small modulation in synaptic plasticity, which also leads to less effective motor learning. Consequently, more diversity of spiking patterns of GR cells results in more effective motor learning for the OKR adaptation

For examination of our main result, we propose a real experiment for the OKR. To control  $p_c$  in a given species of animals (e.g., a species of rabbit, mouse, or zebrafish) in an experiment seems to be practically difficult, unlike the case of computational neuroscience where  $p_c$  can be easily changed. Instead, we consider an experiment for several species of animals (e.g., 3 species of rabbit, mouse, and zebrafish). In each species, we consider a large number of randomly chosen GR cells (i = 1, ..., L). Then, through many learning cycles, one can obtain the peristimulus time histogram (PSTH) for each ith GR cell [i.e., (bin-averaged) instantaneous individual firing rate  $f_{GR}^{(i)}(t)$  of the *i*th GR cell]. GR cells are expected to exhibit diverse PSTHs. Then, in the case of each *i*th GR cell, we obtain its conjunction index  $C_i$  between its PSTH  $f_{GR}^{(i)}(t)$  and the CF signal from the IO neuron [i.e., the PSTH of the IO neuron  $f_{IO}(t)$ ]. In this case, the conjunction index  $C_i$  is given by the cross-correlation at the zero-time lag between  $f_{CR}^{(i)}(t)$ and  $f_{10}(t)$ . Thus, we get the diversity degree  $\mathcal{D}$  of PSTHs of GR cells, given by the relative standard deviation in the distribution of  $\{C_i\}$ , for the species.

Besides the PSTHs of GR cells, under the many learning cycles, we can also get a bell-shaped PSTH of a VN neuron [i.e., (binaveraged) instantaneous individual firing rate  $f_{VN}(t)$  of the VN neuron]. The normalized modulation of  $f_{VN}(t)$  (divided by that at the 1st cycle) corresponds to the learning gain degree  $\mathcal{L}_g$ . Thus, a set of  $(\mathcal{D}, \mathcal{L}_g)$  can be experimentally obtained for each species, and the set of  $(\mathcal{D}, \mathcal{L}_g)$  may vary depending on the species. Then, for example in the case of 3 different species of rabbit, mouse, and zebrafish, with the three different data sets for  $(\mathcal{D}, \mathcal{L}_g)$ , one can examine our main result (i.e., whether more diversity in PSTHs of GR cells results in more effective motor learning for the OKR).

In recent experimental works, dense and similar recodings of the GR cells were observed (Badura & De Zeeuw, 2017; Bengtsson & Jörntell, 2009; Giovannucci et al., 2017; Knogler, Markov, Dragomir, Štih, & Portugues, 2017; Wagner, Kim, Savall, Schnitzer, & Luo, 2017), in contrast to the classical Marr–Albus theory (Albus, 1971; Marr, 1969) where sparse and dissimilar recodings of the GR cells are postulated. At the population level, firing

activities of the GR cells were found to be dense (Badura & De Zeeuw, 2017; Giovannucci et al., 2017; Knogler et al., 2017; Wagner et al., 2017), and in Bengtsson and Jörntell (2009), the MF input codes were found to be preserved in the response of the GR cells (i.e., similar recoding). Based on our computational works, we also discuss how such dense and similar recodings may occur. We first note that, in the GR layer in Fig. 2(a), the inhibitory GO cells make inhibitory control on the firing activity of the GR cells. Hence, the spiking activity of the GR cells may vary depending on the strength of inhibitory control from the GO cells. In our work, we were focused on the inhibitory effect of the GO cells on the firing activity of the GR cells by varying the connection probability  $p_c$  from the GO to the GR cells. The population-level firing activity is well visualized in the raster plot of spikes and the corresponding IPSR  $R_{GR}(t)$  shows collective firing activity of the GR cells. In the optimal case of  $p_c^* = 0.06$ , the cycle-averaged activation degree  $\overline{A(t)}$  (= 0.772) is high, which leads to the spike density in the raster plot of spikes becomes high [Fig. 4(a)]. Hence, the recoding of the GR cells in this optimal case may be regarded as dense recoding. Due to the effect of inhibitory GO cells, the top part of  $R_{GR}(t)$  becomes lowered and flattened [see Fig. 4(b)], and hence the IPSR  $R_{GR}(t)$  has a different waveform with a central plateau, in comparison with the sinusoidally-modulated MF input in Fig. 1(b1). Consequently, the population-level firing activity of the GR cells may be regarded as dissimilar recoding.

We also studied another lowly-connected case  $p_c = 0.006$ . In this case, the inhibitory inputs from the GO cells to the GR cells are so much decreased. Hence, the raster plot of spikes becomes more dense (the cycle-averaged activation degree is A(t) = 0.875 [Fig. 17(a)], but the waveform of the IPSR  $R_{GR}(t)$ becomes more similar to the sinusoidally-modulated MF signal [Fig. 17(b)]. Thus, for  $p_c = 0.006$ , dense and similar recodings seem to occur, in contrast to the optimal case of  $p_c^* = 0.06$ with dense and dissimilar recodings. If the strength of inhibitory control of the GO cells is sufficiently strong (i.e, corresponding to very large  $p_c$ ), sparse recoding may be expected to occur. Judging based on our computational works, dense or sparse recoding and similar or dissimilar recoding of GR cells may appear depending on the strength of inhibitory control of the GO cells. Hence, we speculate that the experimental works on diverse and similar recoding (Badura & De Zeeuw, 2017; Bengtsson & Jörntell, 2009; Giovannucci et al., 2017; Knogler et al., 2017; Wagner et al., 2017) might correspond to the case where the inhibitory effect of GO cells might be so low, so that the possibility for occurrence of dense and similar recodings of the GR cells might become high.

We now discuss our results briefly in comparison with other previous works (Cayco-Gajic, Clopath, & Silver, 2017; Gao, van Beugen, & De Zeeuw, 2012; Inagaki & Hirata, 2017). It was discussed in Gao et al. (2012) that various forms of synaptic plasticity occur at different sites in the cerebellum, and they work synergistically to create optimal outputs for behavior. In the case of synaptic plasticity at the PF-PC synapse, LTD occurs when firing of the PF signal is in-phase (in conjunction) with that of the CF signal. On the other hand, in the absence of the CF signal, the PF signal becomes out-of-phase, and LTP takes place. This mechanism for the synaptic plasticity at the PF-PC synapse is essentially similar to that in our work. However, in our case, we employed a refined rule for the synaptic plasticity with a time window for the LTD (see Fig. 3), based on the experimental result in Safo and Regehr (2008), which has more quantitative advantages. In the presence of a CF firing, a major LTD ( $\Delta$ LTD<sup>(1)</sup>) occurs in conjunction with earlier PF firings, while a minor LTD  $(\Delta LTD^{(2)})$  takes place in association with later PF firings. Outside the effective range of LTD, PF firings alone result in LTP. The GR cells receive both the excitatory synaptic inputs via the MFs and the inhibitory synaptic inputs from the GO cells. It was shown in

Cayco-Gajic et al. (2017) that sparse excitatory synaptic connectivity via  $N_{syn}$  (= 3 ~ 5) MFs is crucial for pattern separation of the MF inputs. In contrast to the work in Cayco-Gajic et al. (2017), we controlled the inhibitory synaptic inputs into the GR cells by changing the connection probability  $p_c$  from the GO cells to the GR cells, and investigated the effect of diverse recoding of GR cells on the motor learning. It was thus found that the saturated learning gain degree  $\mathcal{L}_g^*$  for the OKR is maximum for an optimal value of  $p_c$  (= 0.06).

In Inagaki and Hirata (2017), Inagaki and Hirata studied the motor learning for the vestibulo-ocular reflex in the case that each GR cell receives 6 excitatory MF inputs and 3 inhibitory GOcell inputs. There are 3 types of vestibular, efference-copy, and retinal-slip input signals from the MF. Thus, 9 diverse types of GR cells were prepared depending on the fractions of the vestibular, the efference-copy, and the retinal-slip signals (conveyed via MF inputs). In this case, motor learning for the vestibular ocular reflex was induced by a combination of LTD and LTP at the PF-PC synapses. Moreover, different types of signal processing were required for high- and low-gain motor learning. In contrast to this work, we considered a single type of sinusoidal MF input. Then, diverse spiking patterns of GR cells were shown to arise due to inhibitory coordination of the GO cells. Based on the in-phase and the out-of-phase spiking groups, we investigated the effect of diverse recoding of the GR cells on the motor learning for the OKR.

Finally, we discuss limitations of our present work and future works. In the present work, we did not take into consideration intra-population synaptic connections, and hence no motor rhythms appear in the presence of just inter-population synaptic connections. When we add intra-population couplings between inhibitory GO cells, a granular motor rhythm of 7  $\sim$  25 Hz may appear in the granular layer (i.e., GR-GO feedback system) (D'Angelo et al., 2009). Ultrafast motor rhythm of  $\sim$  200 Hz may also appear in the Purkinje (molecular) layer by adding intra-population synaptic connections between PCs (BCs) (Solages et al., 2008). In the system of IO neurons,  $\sim$  10 Hz motor rhythm appears in the presence of electric gap junctions between IO neurons (Llinás, 2011). Hence, in a future work, it would be interesting to investigate the effect of motor rhythms on diverse recoding of GR cells and learning process in the PC-VN-IO system by adding intra-population couplings. Beyond the synaptic plasticity at PF-PC synapses (considered in this work), diverse synaptic plasticity occurs at other synapses in the cerebellum (Gao et al., 2012; Hansel, Linden, & D'Angelo, 2001) such as synaptic plasticity at PF-BC and BC-PC synapses (Lennon, Yamazaki, & Hecht-Nielsen, 2015), at MF-cerebellar nucleus and PC-cerebellar nucleus synapse (Zheng & Raman, 2010), and at MF-GR cells synapses (D'Angelo & De Zeeuw, 2008). Therefore, as a future work, it would be interesting to study the effect of diverse synaptic plasticity at other synapses on cerebellar motor learning. In addition to variation in  $p_c$ , another possibility to change synaptic inputs into the GR cells is to vary NMDA receptor-mediated maximum conductances  $\bar{g}_{\rm NMDA}^{\rm (GR)}$  and  $\bar{g}_{\rm NMDA}^{\rm (GO)}$ , associated with persistent long-lasting firing activities. It would also be interesting to investigate the effect of NMDA receptormediated synaptic inputs on diverse recoding of GR cells and motor learning in the OKR adaptation by changing  $\bar{g}_{\rm NMDA}^{\rm (GR)}$  and  $\bar{g}_{NMDA}^{(GO)}$ . This work is beyond the scope of the present work, and hence it is left as a future work.

We also discuss another interesting future work. In the present work, we varied the connection probability  $p_c$  (from the GO cells to the GR cells) while the synaptic weight  $J^{(GR,GO)}$  (= 10.0) per synapse is unchanged. Hence, with increasing (decreasing)  $p_c$ , the total inhibitory synaptic strength  $K^{(GR,GO)}$  on each GR cell also increases (decreases). Instead, one may consider another case

### Table A.1

Parameter values for LIF neuron models with AHP currents for the granule (GR) cell and the Golgi (GO) cell in the granular layer, the Purkinje cell (PC) and the basket cell (BC) in the Purkinje-molecular layer, and the vestibular nucleus (VN) and the inferior olive (IO) neurons.

X-population		Granular layer		Purkinje-molecular layer		VN neuron	IO neuron
		GR cell	GO cell	PC	BC		io neuron
C <sub>X</sub>		3.1	28.0	107.0	107.0	122.3	10.0
$I_L^{(X)}$	$g_L^{(X)}$	0.43	2.3	2.32	2.32	1.63	0.67
	$V_L^{(X)}$	-58.0	-55.0	-68.0	-68.0	-56.0	-60.0
	$\bar{g}^{(X)}_{AHP}$	1.0	20.0	100.0	100.0	50.0	1.0
$I_{AHP}^{(X)}$	$ au_{AHP}^{(X)}$	5.0	5.0	5.0	2.5	2.5	10.0
	$V_{AHP}^{(X)}$	-82.0	-72.7	-70.0	-70.0	-70.0	-75.0
	$v_{th}^{(X)}$	-35.0	-52.0	-55.0	-55.0	-38.8	-50.0
$I_{ext}^{(X)}$		0.0	0.0	250.0	0.0	700.0	0.0

#### Table A.2

Parameter values for synaptic currents  $I_R^{(T,S)}(t)$  into the granule (GR) and the Golgi (GO) cells in the granular layer. The GR cells receive excitatory inputs via mossy fibers (MFs) and inhibitory inputs from GO cells. The GO cells receive excitatory inputs via parallel fibers (PFs) from GR cells.

Target cells (T)	GR			GO	GO	
Source cells (S) Receptor (R)	MF AMPA	MF NMDA	GO GABA	PF AMPA	PF NMDA	
$\bar{g}_{R}^{(T)}$	0.18	0.025	0.028	45.5	30.0	
$J_{ij}^{(T,S)}$	8.0	8.0	10.0	0.00004	0.00004	
$V_R^{(S)}$	0.0	0.0	-82.0	0.0	0.0	
$ au_R^{(T)}$	1.2	52.0	7.0, 59.0	1.5	31.0, 170.0	
<i>A</i> <sub>1</sub> , <i>A</i> <sub>2</sub>			0.43, 0.57		0.33, 0.67	

### Table A.3

Parameter values for the synaptic currents  $I_R^{(T,S)}(t)$  into the vestibular nucleus (VN) and the inferior olive (IO) neurons. The VN neuron receives excitatory inputs via MFs and inhibitory inputs from PCs. The IO neuron receives excitatory input via the desired signal (DS) and inhibitory input from the VN neuron.

Target cells (T)	PC			BC
Source cells (S) Receptor (R)	PF AMPA	CF AMPA	BC GABA	PF AMPA
$\bar{g}_{R}^{(T)}$	0.7	0.7	1.0	0.7
$J_{ij}^{(T,S)}$	0.006	1.0	5.3	0.006
$V_R^{(S)}$	0.0	0.0	-75.0	0.0
$ au_R^{(T)}$	8.3	8.3	10.0	8.3

### Table A.4

Parameter values for the synaptic currents  $I_R^{(T,S)}(t)$  into the vestibular nucleus (VN) and the inferior olive (IO) neurons. The VN neuron receives excitatory inputs via MFs and inhibitory inputs from PCs. The IO neuron receives excitatory input via the desired signal (DS) and inhibitory input from the VN neuron.

Target cells (T)	VN			IO		
Source cells (S) Receptor (R)	MF AMPA	MF NMDA	PC GABA	DS AMPA	VN GABA	
$ar{g}_R^{(T)}$	50.0	25.8	30.0	1.0	0.18	
$J_{ij}^{(T,S)}$	0.002	0.002	0.008	1.0	5.0	
$V_R^{(S)}$	0.0	0.0	-88.0	0.0	-75.0	
$ au_R^{(T)}$	9.9	30.6	42.3	10.0	10.0	

where  $p_c$  is changed while maintaining a constant  $K^{(GR,GO)}$ . For keeping the value of  $K^{(GR,GO)}$  to be a constant, the synaptic weight  $J^{(GR,GO)}$  must also change depending on the variation of  $p_c$ ; when  $p_c$  is increased (decreased),  $J^{(GR,GO)}$  should decrease (increase) such that  $K^{(GR,GO)}$  is a constant. We made a preliminary work on this issue by fixing the constant value of  $K^{(GR,GO)}$  at  $K^{(GR,GO)*}$  in the optimal case of  $p_c^* = 0.06$ . Particularly, we were interested in how the spiking patterns in the GR clusters, characterized by  $R_{GR}^{(l)}(t)$ , and the population-averaged firing activity, given by the instantaneous whole-population spike rate  $R_{GR}(t)$ , change when varying  $p_c$ . We first considered the highly-connected case of  $p_c = 0.6$  (with  $K^{(GR,GO)} = K^{(GR,GO)*}$ ). In comparison with the highly-connected case in Fig. 14 where  $K^{(GR,GO)} > K^{(GR,GO)*}$ , shapes of  $R_{GR}(t)$  and  $R_{GR}^{(l)}(t)$  in the case of  $K^{(GR,GO)} = K^{(GR,GO)*}$  were nearly unchanged, while their amplitudes were increased because  $K^{(GR,GO)}$  was decreased to  $K^{(GR,GO)*}$ . We note that the conjunction index  $C^{(l)}$  between  $R_{GR}^{(l)}(t)$  and  $R_{GR}(t)$  represents the degree for similarity between their shapes. Hence, the distribution of  $\{C^{(l)}\}$  in the case of  $K^{(GR,GO)} = K^{(GR,GO)*}$  was found to be nearly the same as that in Fig. 14(c). We also get the diversity degree  $\mathcal{D} (\simeq 0.195)$  for the spiking patterns, which is close to that ( $\mathcal{D} \simeq 0.204$ ) in Fig. 14(c). In this highly-connected case, decrease in the synaptic weight J for maintaining the value of  $K^{(GR,GO)}$  seems to have effect mainly on the amplitudes of spiking patterns without much alteration of their shapes, and hence the diversity for the spiking patterns seems to be determined mainly by the high connectivity from the GO to the GR cells. Consequently, in both cases with and without maintaining the constant  $K^{(GR,GO)*}$ , the diversity degree  $\mathcal{D}$  for the spiking patterns seems to be nearly the same.

Next, we considered the lowly-connected case of  $p_c = 0.006$ where  $K^{(GR,GO)} = K^{(GR,GO)^*}$ . In comparison to the lowly-connected case in Fig. 17 with  $K^{(GR,GO)} < K^{(GR,GO)^*}$ , both the shapes and the amplitudes of  $R_{GR}^{(I)}(t)$  and  $R_{GR}(t)$  were changed. Because  $K^{(GR,GO)}$  was increased to  $K^{(GR,GO)*}$ , amplitudes of  $R_{GR}^{(I)}(t)$  and  $R_{GR}(t)$  were lowered and their top shapes became flattened. Thus, the spiking patterns in the case of  $K^{(GR,GO)} = K^{(GR,GO)*}$  tended to be somewhat similar to those in the optimal case of  $p_c^* = 0.06$ . We note that, in this lowly-connected case, both the low-connectivity  $p_c$ and the increase in synaptic weight I seem to determine the diversity for the spiking patterns, unlike the highly-connected case. Mainly due to increased synaptic weight J, conjunction indices for the spiking patterns were broadly distributed in a range of (-0.17,0.75), in contrast to that in Fig. 17(c). Out-of-phase spiking patterns with negative conjunction indices also appear, unlike the case in Fig. 17 where only in-phase spiking patterns exist. In this case, we obtain the diversity degree is  $\mathcal{D}$  ( $\simeq$  1.192), which is much larger than that ( $\mathcal{D} \simeq 0.175$ ) in the case of Fig. 17. However, it is still smaller than that ( $\mathcal{D}^*~\simeq~1.613$ ) in the optimal case. To get the effect of low connectivity, we start from the optimal case of  $p_c^* = 0.06$  in Fig. 6, and decrease  $p_c$  from 0.06 to 0.006 (lowly-connected case) while maintaining the constant  $K^{(GR,GO)*}$ The range in the distribution of conjunction indices in the optimal case is (-0.57, 0.85). We note that this range became narrowed in the case of lowly-connected case of  $p_c = 0.006$ . Through decrease from  $p_c^*$  to 0.006, in-phase and anti-phase spiking patterns with higher magnitudes of conjunction indices seems to be "degraded", and accordingly their magnitude of conjunction indices seem to be lowered. We conjecture that the connection

### Table B.5

Glossary for various terms which characterize the cerebellar model.				
Terms	Description			
$A_i(t)$ : activation degree	Fraction of active GR cells			
$A_i^{(G)}(t)$ : activation degree in the <i>G</i> spiking group	Fraction of active GR cells in the $G$ spiking group			
$C^{(l)}$ : output conjunction index	The degree for the conjunction of the spiking behavior in each <i>I</i> th GR cluster with the population-averaged spiking behavior in the whole population			
$\mathcal{C}_{in}^{(l)}$ : input conjunction index	The degree for the conjunction of the total synaptic input into each <i>I</i> th GR cluster with the cluster-averaged total synaptic input			
$\mathcal{D}$ : output diversity degree	Relative standard deviation for the distribution of $\{C^{(l)}\}$			
$\mathcal{D}_{in}$ : input diversity degree	Relative standard deviation for the distribution of $\{C_{in}^{(l)}\}$			
$f_{GR}^{(i)}(t)$ : instantaneous individual firing rate	Individual firing activity of active GR cells			
$f_{GR}^{(p)}(t)$ : instantaneous population firing rate	Population firing activity of the whole GR cells			
$f_X(t)$ : firing rate $\mathcal{L}_g$ : learning gain degree	Firing activity of the X cell (X = VN and IO) Modulation gain ratio. Normalized modulation of the individual firing rate $f_{VN}(t)$ of the VN neuron divided by that at the 1st cycle			
$\mathcal{L}_p$ : learning progress degree	Ratio of the cycle-averaged inhibitory input from the VN neuron to the cycle-averaged excitatory input via the IO desired signal			
$\mathcal{M}_d$ : matching degree	Matching degree between $R_{GR}(t)$ (instantaneous whole- population spike rate in the whole population of GR cells) and $f_{DS}(t)$ (IO desired signal)			
$R_{GR}^{(l)}(t)$ : instantaneous cluster spike rate	Spiking behavior in each Ith GR cluster			
$R_X(t)$ : instantaneous whole- population spike rate	Population behavior of X cells			
$\langle W_J^{(G)}(t) \rangle$ : weighted synaptic weight in the <i>G</i> spiking group	Contribution of the <i>G</i> spiking group to $\langle \tilde{J}(t) \rangle$ of active PF signals in the whole population			

probability of  $p_c = 0.006$  would be too small to warrant the highdegree conjunction for the in-phase and the anti-phase spiking patterns. Based on the results of this preliminary work, we hypothesize a possibility that the diversity degree  $\mathcal{D}$  might have its maximum at the same optimal value of  $p_c^* = 0.06$  even in the case of maintaining the constant  $K^{(GR,GO)^*}$ ; in the lowlyconnected case of  $p_c < p_c^*$ ,  $\mathcal{D}$  would decrease in a much slow way in comparison with the case without maintaining the constant  $K^{(GR,GO)^*}$ . To confirm this hypothesis, more intensive future work is necessary by decreasing  $p_c$  from  $p_c^*$  in the lowly-connected case with the constant  $K^{(GR,GO)^*}$  where changes in both  $p_c$  and J have effects on spiking patterns.

#### **Declaration of competing interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

### Acknowledgment

This research was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education (Grant No. 20162007688).

# Appendix A. Parameter values for the LIF neuron models and the synaptic currents

In this appendix, we list four tables which show parameter values for the LIF neuron models in Section 2.3 and the synaptic currents in Section 2.4. These values are adopted from physiological data (Yamazaki & Nagao, 2012; Yamazaki & Tanaka, 2007a).

For the LIF neuron models, the parameter values for the capacitance  $C_X$ , the leakage current  $I_L^{(X)}$ , the AHP current  $I_{AHP}^{(X)}$ , and the external constant current  $I_{ext}^{(X)}$  are given in Table A.1.

For the synaptic currents, the parameter values for the maximum conductance  $\bar{g}_{R}^{(T)}$ , the synaptic weight  $J_{ij}^{(T,S)}$ , the synaptic reversal potential  $V_{R}^{(S)}$ , the synaptic decay time constant  $\tau_{R}^{(T)}$ , and the amplitudes  $A_{1}$  and  $A_{2}$  for the type-2 exponential-decay function in the granular layer, the Purkinje-molecular layer, and the other parts for the VN and IO are given in Tables A.2, A.3, and A.4, respectively.

### Appendix B. Glossary

In this appendix, glossary for various terms characterizing the cerebellar model is given in Table B.5.

### References

- Achard, P., & De Schutter, E. (2008). Calcium, synaptic plasticity and intrinsic homeostasis in Purkinje neuron models. *Frontiers in Computational Neuroscience*, 2, 8.
- Albus, J. S. (1971). A theory of cerebellar function. *Mathematical Biosciences*, 10, 25-61.
- Badura, A., & De Zeeuw, C. I. (2017). Cerebellar granule cells: Dense, rich and evolving representations. *Current Biology*, 27, R415–R418.
- Bengtsson, F., & Jörntell, H. (2009). Sensory transmission in cerebellar granule cells relies on similarly coded mossy fiber inputs. Proceedings of the National Academy of Sciences of the United States of America, 106, 2389–2394.
- Bouvier, G., Aljadeff, J., Clopath, C., Bimbard, C., Ranft, J., Blot, A., et al. (2018). Cerebellar learning using perturbations. *eLife*, 7, Article e31599.
- Brickley, S. G., Cull-Candy, S. G., & Farrant, M. (1996). Development of a tonic form of synaptic inhibition in rat cerebellar granule cells resulting from persistent activation of GABA<sub>A</sub> receptors. *Journal Physiology*, 497, 753–759.
- Brindley, G. S. (1964). The use made by the cerebellum of the information that it receives from sense organs. *IBRO Bulletin*, 3, 80.
- Brunel, N. (2000). Dynamics of sparsely connected networks of excitatory and inhibitory spiking neurons. Journal of Computational Neuroscience, 8, 183–208.

- Brunel, N., & Hakim, V. (1999). Fast global oscillations in networks of integrate-and-fire neurons with low firing rates. *Neural Computation*, 11, 1621–1671.
- Brunel, N., & Hakim, V. (2008). Sparsely synchronized neuronal oscillations. Chaos, 18, Article 015113.
- Brunel, N., & Hansel, D. (2006). How noise affects the synchronization properties of recurrent networks of inhibitory neurons. *Neural Computation*, *18*, 1066–1110.
- Brunel, N., & Wang, X.-J. (2003). What determines the frequency of fast network oscillations with irregular neural discharges? I. Synaptic dynamics and excitation-inhibition balance. *Journal of Neurophysiology*, 90, 415–430.
- Buonomano, D. V., & Mauk, M. D. (1994). Neural network model of the cerebellum: Temporal discrimination and the timing of motor responses. *Neural Computation*, 6, 38–55.
- Cayco-Gajic, N. A., Clopath, C., & Silver, A. (2017). Sparse synaptic connectivity is required for decorrelation and pattern separation in feedforward networks. *Nature Communications*, 8, 1116.
- Chen, C., & Thompson, R. F. (1995). Temporal specificity of long-term depression in parallel fiber-Purkinje synapses in rat cerebellar slice. *Learning and Memory*, 2, 185–198.
- Christian, K. M., & Thompson, R. F. (2003). Neural substrates of eyeblink conditioning: Acquisition and retention. *Learning and Memory*, 11, 427-455.
- Coesmans, M., Weber, J. T., De Zeeuw, C. I., & Hansel, C. (2004). Bidirectional parallel fiber plasticity in the cerebellum under climbing fiber control. *Neuron*, 44, 691–700.
- D'Angelo, E., & De Zeeuw, C. I. (2008). Timing and plasticity in the cerebellum: Focus on the granular layer. *Trends in Neurosciences*, 32, 30–40.
- D'Angelo, E., Koekkoek, S. K. E., Lombardo, P., Solinas, S., Ros, E., Garrido, J., et al. (2009). Timing in the cerebellum: Oscillations and resonance in the granular layer. *Neuroscience*, 162, 805–815.
- De Schutter, E. (1995). Cerebellar long-term depression might normalize excitation of Purkinje cells: A hypothesis. *Trends in Neurosciences*, 18, 291–295.
- Dean, P., Porrill, J., Ekerot, C.-F., & Jörntell, H. (2010). The cerebellar microcircuit as an adaptive filter: Experimental and computational evidence. *Nature Reviews. Neuroscience*, *11*, 30–43.
- Fujita, M. (1982). Adaptive filter model of the cerebellum. *Biological Cybernetics*, 45, 195–206.
- Gallimore, A. R., Kim, T., Tanaka-Yamamoto, K., & De Schutter, E. (2018). Switching on depression and potentiation in the cerebellum. *Cell Reports*, 22, 722–733.
- Gao, Z. Z., van Beugen, B. J., & De Zeeuw, C. I. (2012). Distributed synergistic plasticity and cerebellar learning. *Nature Reviews. Neuroscience*, 13, 619–635.
- Geisler, C., Brunel, N., & Wang, X.-J. (2005). Contributions of intrinsic membrane dynamics to fast network oscillations with irregular neuronal discharges. *Journal of Neurophysiology*, 94, 4344–4361.
- Gerstner, W., & van Hemmen, J. L. (1992). Associative memory in a network of 'spiking' neurons. Network, 3, 139–164.
- Gerstner, W., & Kistler, W. (2002). Spiking neuron models. New York: Cambridge University Press.
- Gilman, S., Bloedel, J., & Lechtenberg, R. (1981). Disorders of the cerebellum. Philadelphia: F.A. Davis Company.
- Giovannucci, A., Badura, A., Deverett, B., Najafi, F., Pereira, T. D., Gao, Z., et al. (2017). Cerebellar granule cells acquire a widespread predictive feedback signal during motor learning. *Nature Neuroscience*, 20, 727–734.
- Gomi, H., & Kawato, M. (1992). Adaptive feedback control models of the vestibulocerebellum and spinocerebellum. *Biological Cybernetics*, 68, 105–114.
- Hansel, C., Linden, D. J., & D'Angelo, E. (2001). Beyond parallel fiber LTD: The diversity of synaptic and non-synaptic plasticity in the cerebellum. *Nature Neuroscience*, 4, 467–475.
- Harvey, R. J., De'Sperati, C., & Strata, P. (1997). The early phase of horizontal optokinetic responses in the pigmented rat and the effects of lesions of the visual cortex. *Vision Research*, *37*, 1615–1625.
- Häusser, M., & Clark, B. A. (1997). Tonic synaptic inhibition modulates neuronal output pattern and spatiotemporal synaptic integration. *Neuron*, 19, 665–678.
   Hebb, D. O. (1949). The organization of behavior; A neuropsychological theory. New
- York: Wiley & Sons. Heine, S. A., Highstein, S. M., & Blazquez, P. M. (2010). Golgi cells operate as
- state-specific temporal filters at the input stage of the cerebellar cortex. *The Journal of Neuroscience*, 30, 17004–17014.
- Huang, Y.-Y., & Neuhauss, S. C. F. (2008). The optokinetic response in zebrafish and its applications. Frontiers in Bioscience, 13, 1899–1916.
- Inagaki, K., & Hirata, Y. (2017). Computational theory underlying acute vestibuloocular reflex motor learning with cerebellar long-term depression and long-term potentiation. *Cerebellum*, 16, 827–839.
- Ito, M. (1984). The cerebellum and neural control. New York: Raven Press.
- Ito, M. (1989). Long-term depression. Annual Review of Neuroscience, 12, 85–102.
  Ito, M. (1998). Cerebellar learning in the vestibulo-ocular reflex. Trends in Cognitive Sciences, 2, 313–321.
- Ito, M. (2000). Mechanisms of motor learning in the cerebellum. *Brain Research*, 886, 237-245.

- Ito, M. (2001). Cerebellar long-term depression: Characterization, signal transduction, and functional roles. *Physiological Reviews*, 81, 1143–1195.
- Ito, M. (2002a). Historical review of the significance of the cerebellum and the role of Purkinje cells in motor learning. *Annals of the New York Academy of Sciences*, 978, 273–288.
- Ito, M. (2002b). The molecular organization of cerebellar long-term depression. Nature Reviews Neuroscience, 3, 896–902.
- Ito, M. (2012). The cerebellum: Brain for an implicit self. New Jersey: Pearson Education Inc.
- Ito, M., & Kano, M. (1982). Long-lasting depression of parallel fiber-Purkinje cell transmission induced by conjunctive stimulation of parallel fibers and climbing fibers in the cerebellar cortex. *Neuroscience Letters*, 33, 253–258.
- Ito, M., Sakurai, M., & Tongroach, P. (1982). Climbing fibre induced depression of both mossy fibre responsiveness and glutamate sensitivity of cerebellar Purkinje cells. *The Journal of Physiology*, 324, 113–134.
- Iwashita, M., Kanai, R., Funabiki, K., Matsuda, K., & Hirano, T. (2001). Dynamic properties, interactions and adaptive modifications of vestibulo-ocular reflex and optokinetic response in mice. *Neuroscience Research*, 39, 299–311.
- Jakab, R. L., & Hámori, J. (1988). Quantitative morphology and synaptology of cerebellar glomeruli in the rat. Anatomy and Embryology, 179, 81–88.
- Kenyon, G. T., Medina, J. F., & Mauk, M. D. (1998). A mathematical model of the cerebellar-olivary system I: Self-regulating equilibrium of climbing fiber activity. *Journal of Computational Neuroscience*, 5, 17–33.
- Kim, S.-Y., & Lim, W. (2014). Realistic thermodynamic and statistical-mechanical measures for neural synchronization. *Journal of Neuroscience Methods*, 226, 161–170.
- Knogler, L. D., Markov, D. A., Dragomir, E. I., Štih, V., & Portugues, R. (2017). Sensorimotor representations in cerebellar granule cells in larval zebrafish are dense, spatially organized, and non-temporally patterned. *Current Biology*, 27, 1288–1302.
- Lennon, W., Yamazaki, T., & Hecht-Nielsen, R. (2015). A model of in vitro plasticity at the parallel fiber Molecular layer interneuron synapses. *Frontiers in Computational Neuroscience*, 9, 150.
- Lev-Ram, V., Mehta, S. B., Kleinfeld, D., & Tsien, R. Y. (2003). Reversing cerebellar long-term depression. Proceedings of the National Academy of Sciences of the United States of America, 100, 15989–15993.
- Llinás, R. R. (2011). Cerebellar motor learning versus cerebellar motor timing: The climbing fiber story. *The Journal of Physiology*, 589, 3423–3432.
- Llinás, R. R. (2014). The olivo-cerebellar system: A key to understanding the functional significance of intrinsic oscillatory brain properties. Frontiers in Neural Circuits, 7, 96.
- Maass, W., Natschläger, T., & Markram, H. (2002). Real-time computing without stable states: A new framework for neural computation based on perturbations. *Neural Computation*, 14, 2531–2560.
- Manto, M. U. (2010). Cerebellar disorders; A practical approach to diagnosis and managements. Cambridge: Cambridge University Press.
- Marr, D. (1969). A theory of cerebellar cortex. The Journal of Physiology, 202, 437-470.
- Mathy, A., Ho, S. S. N., Davie, J. T., Duguid, I. C., Clark, B. A., & Häusser, M. (2009). Encoding of oscillations by axonal bursts in inferior olive neurons. *Neuron*, *62*, 388–399.
- Matsuno, H., Kudoh, M., Watakabe, A., Yamamori, T., Shigemoto, R., & Nagao, S. (2016). Distribution and structure of synapses on medial vestibular nuclear neurons targeted by cerebellar flocculus Purkinje cells and vestibular nerve in mice: Light and electron microscopy studies. *PLoS One*, *11*, Article e0164037.
- Mauk, M. D., & Donegan, N. H. (1997). A model of Pavlovian eyelid conditioning based on the synaptic organization of the cerebellum. *Learning and Memory*, 3, 130–158.
- Medina, J. F., Garcia, K. S., Nores, W. L., Taylor, N. M., & Mauk, M. D. (2000). Timing mechanisms in the cerebellum: Testing predictions of a large-scale computer simulation. *Journal of Neuroscience*, 20, 5516–5525.
- Molnár, E. (2014). Motor learning and long-term plasticity of parallel fibre-Purkinje cell synapses require post-synaptic Cdk5/p35. Journal of Neurochemistry, 131, 1–3.
- Nagao, S. (1983). Effects of vestibulocerebellar lesions upon dynamic characteristics and adaptation of vestibulo-ocular and optokinetic responses in pigmented rabbits. *Experimental Brain Research*, 53, 36–46.
- Nagao, S. (1988). Behavior of floccular Purkinje cells correlated with adaptation of horizontal optokinetic eye movement response in pigmented rabbits. *Experimental Brain Research*, 73, 489–497.
- Palkovits, M., Magyar, P., & Szentágothai, J. (1971a). Quantitative histological analysis of the cerebellar cortex in the cat. I. Number and arrangement in space of the Purkinje cells. *Brain Research*, 32, 1–13.
- Palkovits, M., Magyar, P., & Szentágothai, J. (1971b). Quantitative histological analysis of the cerebellar cortex in the cat. II. Cell numbers and densities in the granular layer. *Brain Research*, *32*, 13–32.
- Palkovits, M., Magyar, P., & Szentágothai, J. (1972). Quantitative histological analysis of the cerebellar cortex in the cat. IV. Mossy fiber-Purkinje cell numerical transfer. *Brain Research*, 45, 15–29.

- Pearson, K. (1895). Notes on regression and inheritance in the case of two parents. *Proceedings of The Royal Society of London*, 58, 240–242.
- Roberts, P. D. (2007). Stability of complex spike timing-dependent plasticity in cerebellar learning. *Journal of Computational Neuroscience*, 22, 283–296.
- Safo, P., & Regehr, W. G. (2008). Timing dependence of the induction of cerebellar LTD. Neuropharmacology, 54, 213–218.
- Sakurai, M. (1987). Synaptic modification of parallel fibre-Purkinje cell transmission in vitro guinea-pig cerebellar slices. *The Journal of Physiology*, 394, 463–480.
- Scheetz, S. D., Shao, E., Zhou, Y., Cario, C. L., Bai, Q., & Burton, E. A. (2017). An open-source method to analyze optokinetic reflex responses in larval zebrafish. *Journal of Neuroscience Methods*, 293, 329–337.
- Sejnowski, T. J. (1977). Storing covariance with nonlinearly interacting neurons. Journal of Mathematical Biology, 4, 303-321.
- Shimazaki, H., & Shinomoto, S. (2010). Kernel bandwidth optimization in spike rate estimation. Journal of Computational Neuroscience, 29, 171–182.
- Solages, C. de, Szapiro, G., Brunel, N., Hakim, V., Isope, P., Buisseret, P., et al. (2008). High-frequency organization and synchrony of activity in the Purkinje cell layer of the cerebellum. *Neuron*, 58, 775–788.
- Steuber, V., Mittmann, W., Hoebeek, F. E., Silver, R. A., De Zeeuw, C. I., Häusser, M., et al. (2007). Cerebellar LTD and pattern recognition by Purkinje cells. *Neuron*, 54, 121–136.
- Strata, P. (2009). David Marr's theory of cerebellar learning: 40 years later. The Journal of Physiology, 587, 5519–5520.
- Strogatz, S. H. (2001). Exploring complex networks. Nature, 410, 268-276.

- Tabata, H., Shimizu, N., Wada, Y., Miura, K., & Kawano, K. (2010). Initiation of the optokinetic response (OKR) in mice. *Journal of Vision*, *10*, 13.
- Thach, W. T. (1968). Discharge of Purkinje and cerebellar nuclear neurons during rapidly alternating arm movements in the monkey. *Journal of Neurophysiology*, 31, 785–797.
- Wagner, M. J., Kim, T. H., Savall, J., Schnitzer, M. J., & Luo, L. (2017). Cerebellar granule cells encode the expectation of reward. *Nature*, 544, 96–100.
- Wang, X.-J. (2010). Neurophysiological and computational principles of fscortical rhythms in cognition. *Physiological Reviews*, 90, 1195–1268.
- Wang, S.-H., Denk, W., & Häusser, M. (2000). Coincidence detection in single dendritic spines mediated by calcium release. *Nature Neuroscience*, 3, 1266–1273.
- Watts, D. J., & Strogatz, S. H. (1998). Collective dynamics of 'small-world' networks. Nature, 393, 440–442.
- Yamazaki, T., & Nagao, S. (2012). A computational mechanism for unified gain and timing control in the cerebellum. *PLoS One*, 7, Article e33319.
- Yamazaki, T., & Tanaka, S. (2007a). A spiking network model for passage-oftime representation in the cerebellum. *European Journal of Neuroscience*, 26, 2279–2292.
- Yamazaki, T., & Tanaka, S. (2007b). The cerebellum as a liquid state machine. Neural Networks, 20, 290–297.
- Yang, Y., & Lisberger, S. G. (2014). Purkinje-cell plasticity and cerebellar motor learning are graded by complex-spike duration. *Nature*, 510, 529–532.
- Zheng, N., & Raman, I. M. (2010). Synaptic inhibition, excitation, and plasticity in neurons of the cerebellar nuclei. *Cerebellum*, *9*, 56–66.